

MiRNA-146b-5p inhibits the malignant progression of gastric cancer by targeting TRAF6

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Abstract. – OBJECTIVE: MicroRNAs (miRNAs) are 22 nucleotides long that are extensively expressed in eukaryotes. They are vital regulators in pathological processes. This study aims to illustrate the role of miRNA-146b-5p in the development of gastric cancer (GC).

PATIENTS AND METHODS: MiRNA-146b-5p levels in 62 GC species and matched paracancerous ones were detected. Influences of miRNA-146b-5p level on clinical parameters of GC patients were assessed. Phenotype changes of AGS and SGC-7901 cells overexpressing miRNA-146b-5p were evaluated by Cell Counting Kit-8 (CCK-8) and transwell assay, respectively. Luciferase assay and rescue experiments were conducted to uncover the mechanism of miRNA-146b-5p in regulating the development of GC.

RESULTS: MiRNA-146b-5p was downregulated in GC species than paracancerous ones. Low level of miRNA-146b-5p was observed in patients combined lymphatic metastasis and distant metastasis than those without metastases. *In vitro* overexpression of miRNA-146b-5p attenuated proliferative and migratory potentials of GC cells. TRAF6 was the target of miRNA-146b-5p, which was responsible for the development of GC regulated by miRNA-146b-5p.

CONCLUSION: MiRNA-146b-5p is negatively correlated to lymphatic metastasis and distant metastasis rates of GC. It suppresses the malignant development of GC by targeting TRAF6.

Keywords:

MiRNA-146b-5p, TRAF6, Gastric cancer (GC), Malignant development.

Introduction

Gastric cancer (GC) is a popular digestive system tumor. Its incidence has been sharply reduced in developed countries. In the United States, the incidence of GC reduced about 20% in the past two decades. However, Asia is the area with high morbidity and mortality of GC. GC remains to be the

second leading fatal disease in many countries¹⁻⁵. It is estimated that there are about 100 million newly onsets of GC. More than 8 million people die of this tumor each year. In China, about 400,000 GC patients are initially diagnosed as advanced GC. Seriously, more than 50% GC patients develop postoperative recurrence with a median survival of shorter than 16 months⁶. Low detective rate of early-stage GC, insensitive chemotherapy and radiotherapy, as well as deficiency of effective biomarkers, all result in the poor prognosis of GC^{7,8}. It is urgent to clarify the pathogenesis and etiology of GC, thus develop novel biomarkers of GC⁹⁻¹¹.

MicroRNAs (miRNAs) are non-coding RNAs widely distributed in eukaryotes^{12,13}. They exert post-transcriptional regulation on target gene silencing through degrading or inhibiting translation of mRNAs after recognizing their 3'UTRs^{14,15}. MiRNAs are featured by high conservation and tissue specificity, which are involved in cell behavior regulations^{16,17}. By targeting oncogenes or tumor-suppressor genes, miRNAs are able to influence tumor development¹⁸. Previous studies^{19,20} have shown the potentials of miRNAs as diagnostic and therapeutic targets of GC.

In this paper, a total of 62 GC species and paired paracancerous ones were collected. The role of miRNA-146b-5p/TRAF6 axis in mediating the malignant progression of GC was explored.

Patients and Methods

Patients and Samples

62 paired GC species and paracancerous ones (5 cm away from tumor edge) were collected and stored at -80°C. None of enrolled subjects were pre-operatively treated with anti-tumor therapy. Their clinical data were recorded. Patients and their families in this study have been fully informed. This study was approved by Ethics Committee of the Second Hospital of Shandong University.

Cell Culture

GC cell lines (AGS, BGC-823, SGC-7901, MKN28, and MKN45) and epithelial cells of gastric mucosa (GES-1) were cultured in Dulbecco's Modified Eagle's Medium (DMEM; Gibco, Rockville, MD, USA) containing 10% fetal bovine serum (FBS; Gibco, Rockville, MD, USA), 100 UI/mL penicillin, and 100 µg/mL streptomycin in a 5% CO₂ incubator at 37°C. Cell passage was conducted in 1×typsin+EDTA (ethylenediaminetetraacetic acid) at 80-90% confluence.

Transfection

Cells were grown at 50-70% confluence in 6-well plates and transfected with plasmids constructed by GenePharma (Shanghai, China), using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA). Transfected cells for 48 h were harvested for functional experiments.

Cell Proliferation Assay

Cells were inoculated in a 96-well plate with 2×10³ cells per well. At the appointed time points, absorbance value at 490 nm of each sample was recorded using the Cell Counting Kit-8 (CCK-8) kit (Dojindo Molecular Technologies, Kumamoto, Japan) for plotting the viability curves.

Transwell Assay

200 µL of suspension (1.0×10⁵ cells/mL) was applied in the upper side of the transwell chamber (Millipore, Billerica, MA, USA) inserted in a 24-well plate with 700 µL medium containing 10% FBS in the bottom. After 48 h, the cells in the bottom were fixed with methanol for 15 min, dyed with crystal violet for 10 min, and counted using microscope. Migrator cell number was counted in 5 randomly selected fields per sample (magnification 40×).

Quantitative Real Time-Polymerase Chain Reaction (qRT-PCR)

Extractions RNAs by TRIzol reagent (Invitrogen, Carlsbad, CA, USA) were purified by DNase treatment, and reversely transcribed into complementary deoxyribose nucleic acids (cDNAs) using PrimeScript RT Reagent (TaKaRa, Otsu, Shiga, Japan). Purified cDNAs underwent qRT-PCR using SYBR[®] Premix Ex Taq[™] (TaKaRa, Otsu, Japan). β-actin and U6 were used as the internal references. Each sample was performed in triplicate, and the relative level was calculated by 2^{-ΔΔCt}. miRNA-146b-5p: forward: 5'-TGACCCATCCTGGGCCTCAA-3', reverse: 5'-CCAGT-

GGGCAAGATGTGGGCC-3'; U6: forward: 5'-CGCTTCGGCAGCACATATAC-3', reverse: 5'-TTCACGAATTTGCGTGTTCAT-3'; TRAF6: forward: 5'-TGCTTGATGGCATTACGACAT-3', reverse: 5'-CATTGGACATTTCAAT-3'; β-actin: forward: 5'-CCTGCCACCCAGCACAAAT-3', reverse: 5'-TGCCAGGGTGTCCTTTG-3'.

Western Blot

Cells were lysed for separating cellular proteins and electrophoresed. Protein samples were loaded on polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). Subsequently, non-specific proteins were blocked by 5% skim milk for 2 h. Membranes were reacted with primary and secondary antibodies for indicated time. Band exposure and analyses were finally conducted.

Luciferase Assay

Cells inoculated in a 24-well plate were cotransfected with pmirGLO-TRAF6-WT/pmirGLO-TRAF6-Mut/pmirGLO and NC/miRNA-146b-5p, respectively. 48 hours later, the cells were lysed for determining relative Luciferase activity (Promega, Madison, WI, USA).

Statistical Analysis

GraphPad Prism 5 V6.01 (La Jolla, CA, USA) was used for data analyses. Data were expressed as mean ± standard deviation (SD). The differences between the two groups were analyzed by the *t*-test. Chi-square analysis was used to assess the relationship between miRNA-146b-5p level and clinical parameters of GC patients. *p*<0.05 was considered as statistically significant.

Results

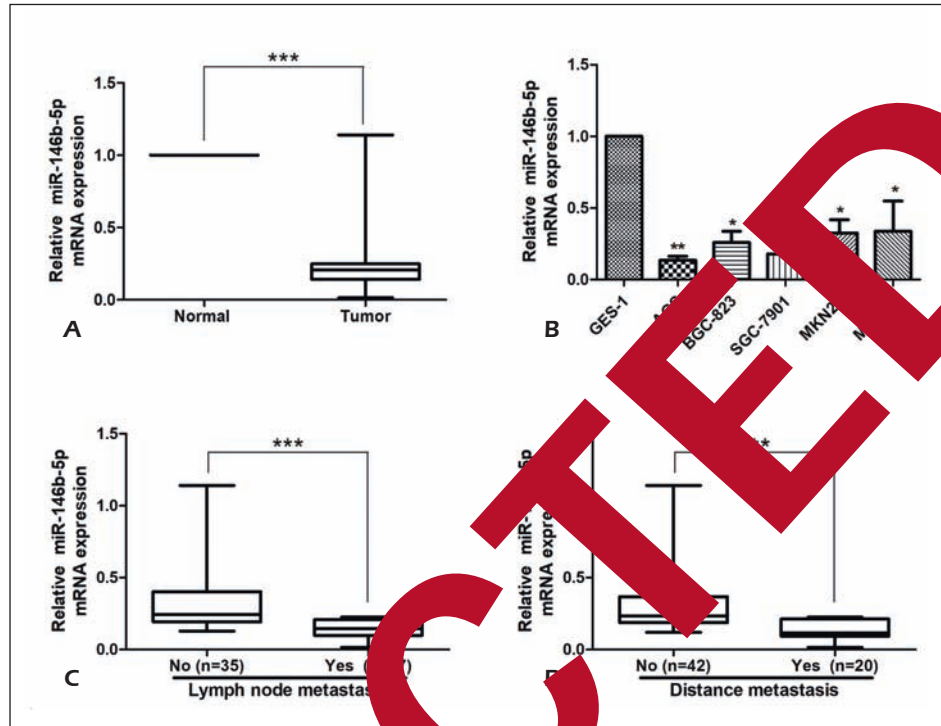
Downregulated MiRNA-146b-5p in GC

Compared with paracancerous species, miRNA-146b-5p was downregulated in GC tissues (Figure 1A). Similarly, miRNA-146b-5p was lowly expressed in GC cell lines than that of GES-1 cell line (Figure 1B).

MiRNA-146b-5p Level was Linked to Lymphatic Metastasis and Distant Metastasis of GC

Based on median level of miRNA-146b-5p in enrolled GC patients, they were assigned into

Figure 1. Downregulated miRNA-146b-5p in GC. **A**, MiRNA-146b-5p levels in GC species and paracancerous ones. **B**, MiRNA-146b-5p levels in GC cell lines. **C**, MiRNA-146b-5p levels in GC patients either with lymph node metastasis or not. **D**, MiRNA-146b-5p levels in GC patients either with distant metastasis or not. Data were expressed as mean±SD. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.



high or low-level groups. Chi-square analysis showed that miRNA-146b-5p level was linked to rates of lymphatic metastasis and distant metastasis, while it was not correlated to age, sex, and tumor grade of GC patients (Table I). Moreover, lower level of miRNA-146b-5p was observed in GC patients combined lymphatic metastasis and distant metastasis than those without metastases (Figure 1C, 1D). MiRNA-146b-5p may be a novel biomarker of GC.

Overexpression of MiRNA-146b-5p Suppressed Proliferative and Migratory Potentials of GC

The inhibition efficacy of miRNA-146b-5p mimics was first verified in AGS and SGC-7901 cells (Figure 2A). Viability of GC cells was markedly reduced after overexpression of miRNA-146b-5p (Figure 2B). Besides, reduced migratory cell number was seen in GC cells overexpressing miRNA-146b-5p (Figure 2C).

Table I. LncRNA H19 and miR-146b-5p expression and clinical features of patients with ovarian cancer.

Parameters	Number of cases	miR-146b-5p expression		p-value
		High (%)	Low (%)	
Age (years)	<60	11	16	0.200
	≥60	20	15	
Gender	Female	18	21	0.529
	Male	12	10	
Tumor size (cm)	<5	10	12	0.596
	≥5	21	19	
Lymph node metastasis	No	22	13	0.021
	Yes	9	18	
Distance metastasis	No	25	17	0.030
	Yes	6	14	

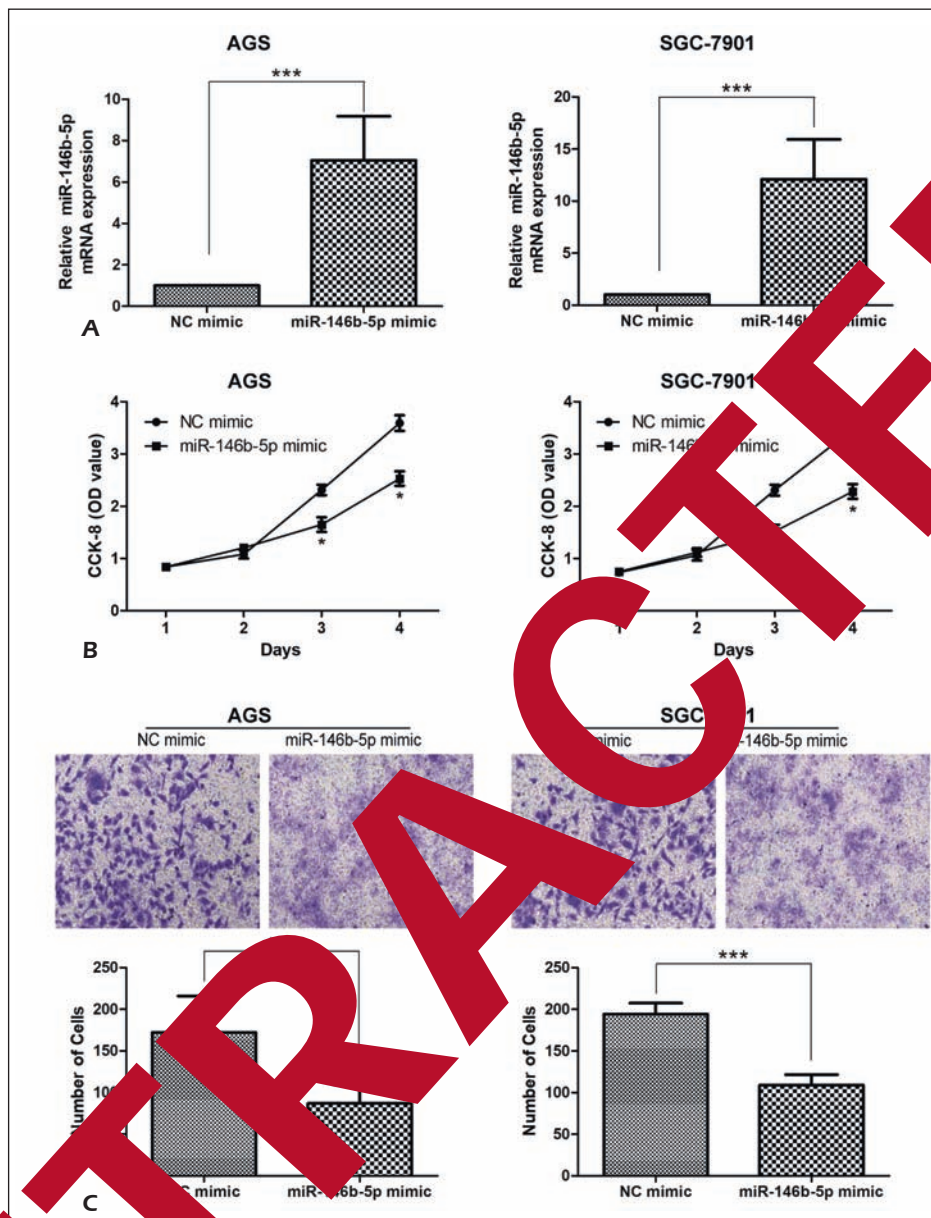


Figure 7 Overexpression of miRNA-146b-5p suppressed proliferative and migratory potentials of GC. **A**, Transfection efficacy of miRNA-146b-5p mimics. **B**, Viability in AGS and SGC-7901 cells transfected with NC mimics and miRNA-146b-5p mimics. **C**, Migration in AGS and SGC-7901 cells transfected with NC mimics and miRNA-146b-5p mimics (magnification 40 \times). Data were expressed as mean \pm SD. * p <0.05, *** p <0.001.

miRNA-146b-5p Targeted TRAF6
 Potential binding sequences in 3'UTR of miRNA-146b-5p and TRAF6 were predicted (Figure 2). Luciferase activity in GC cells cotransfected with miRNA-146b-5p mimics and pGL3-TRAF6-WT further showed the binding between miRNA-146b-5p and TRAF6. Protein and mRNA levels of TRAF6 were downregulated in AGS and SGC-7901 cells overexpressing miRNA-146b-5p (Figure 3A). In addition, TRAF6

was highly expressed in GC species and cell lines (Figure 3B, 3C).

Overexpression of TRAF6 Abolished Inhibitory Effect of overexpressed MiRNA-146b-5p on Malignant Phenotypes of GC

Rescue experiments were designed for clarifying the involvement of TRAF6 in GC development regulated by miRNA-146b-5p. First of all, the over-

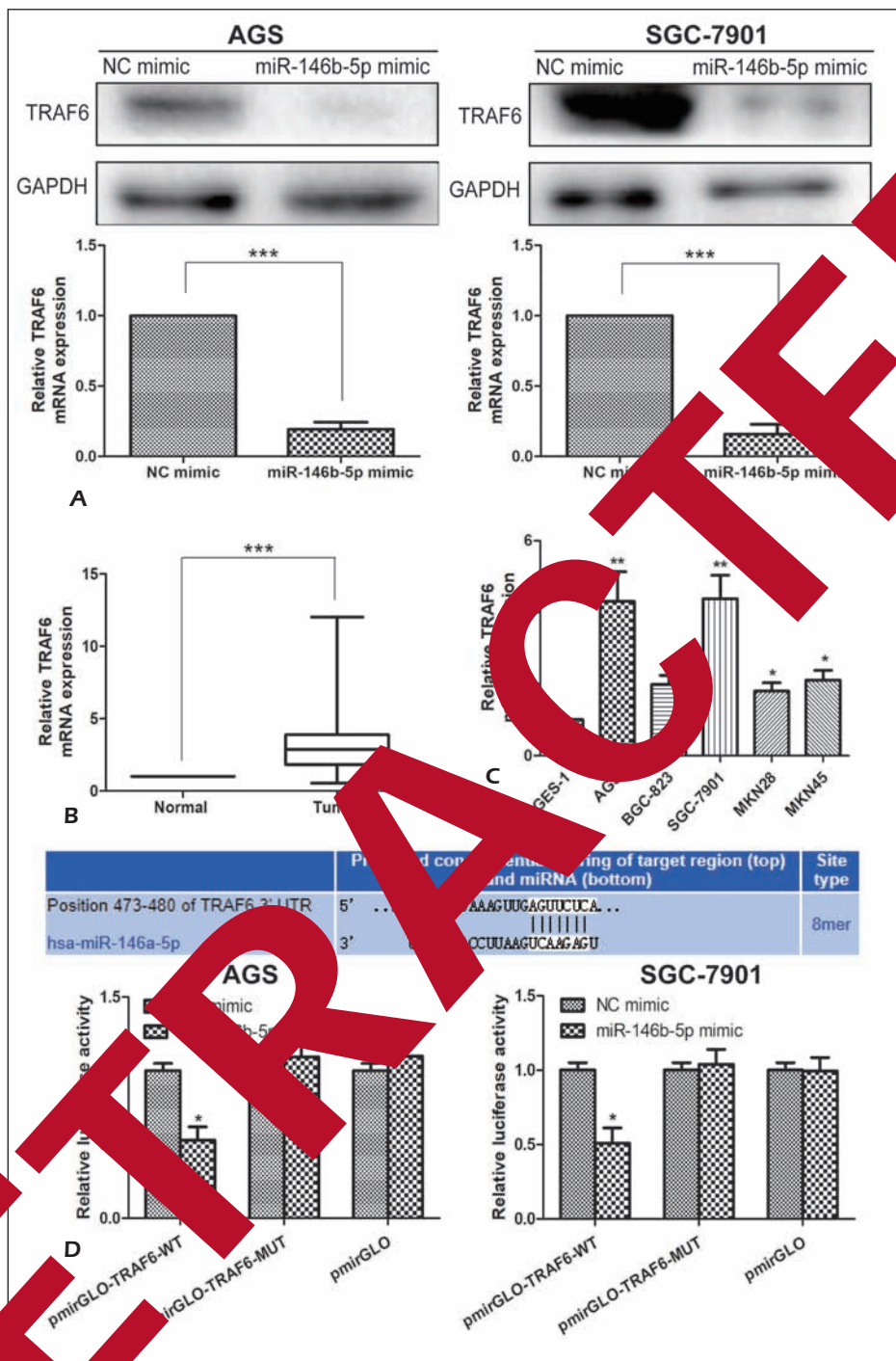


Figure 3. MiRNA-146b-5p targeted TRAF6. **A**, Protein and mRNA levels of TRAF6 in AGS and SGC-7901 cells transfected with NC mimics and miRNA-146b-5p mimics. **B**, TRAF6 levels in GC species and paracancerous ones. **C**, TRAF6 levels in GC cell lines. **D**, Potential binding sequences in 3'UTR of miRNA-146b-5p and TRAF6. Luciferase activity in AGS and SGC-7901 cells transfected with pmirGLO-TRAF6-WT/pmirtGLO-TRAF6-MUT/pmirtGLO and NC/miRNA-146b-5p mimics. Data are expressed as mean±SD. **p*<0.05, ***p*<0.01, ****p*<0.001.

expression of TRAF6 was able to downregulate miRNA-146b-5p in GC cells overexpressing miRNA-146b-5p (Figure 4A) and upregulate TRAF6 as well (Figure 4B). Notably, the overexpression of

TRAF6 reversed the reduced migratory potential in GC cells overexpressing miRNA-146b-5p (Figure 4C). Hence, TRAF6 was responsible for malignant development of GC regulated by miRNA-146b-5p.

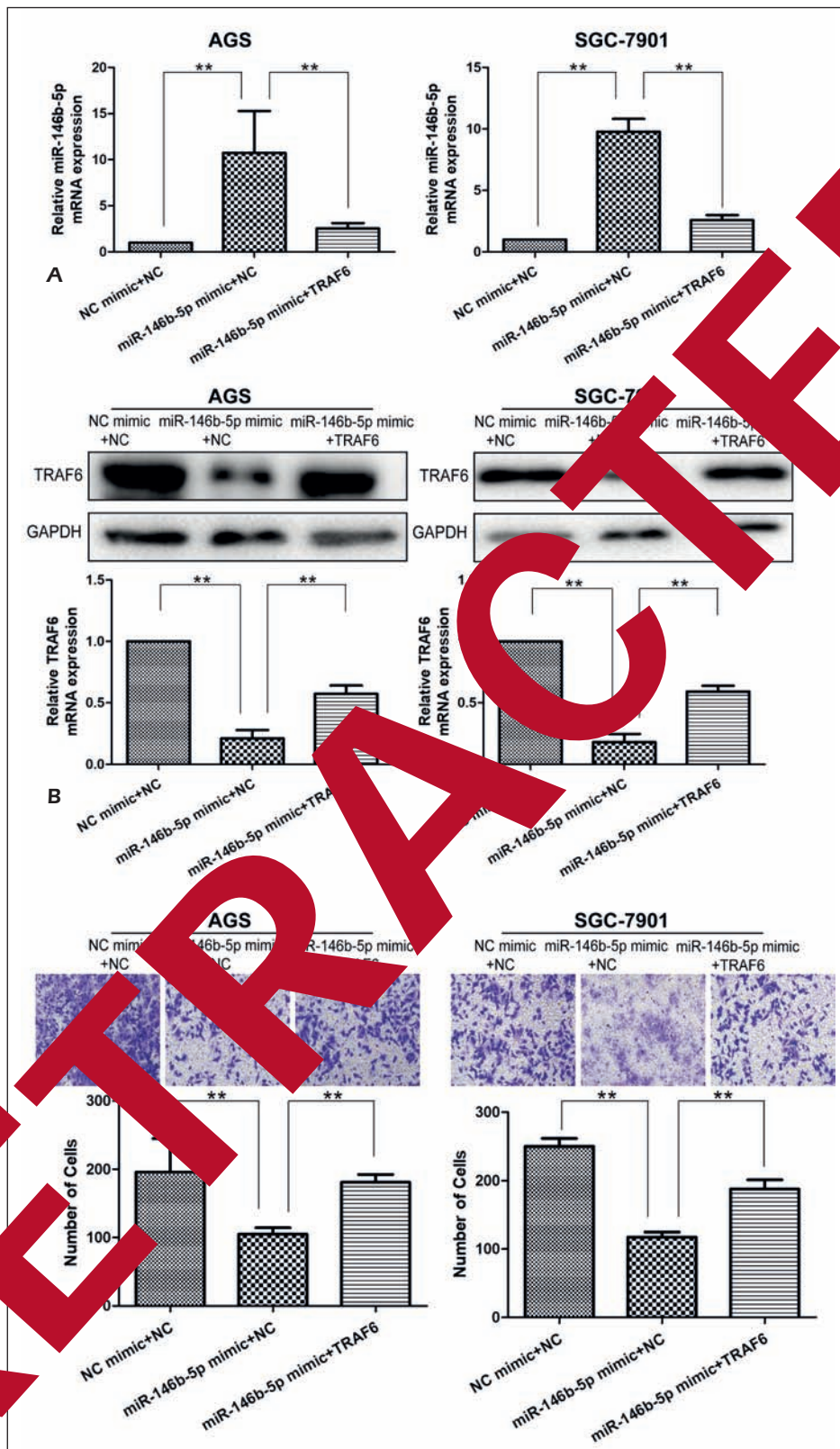


Figure 4. Overexpression of TRAF6 abolished inhibitory effect of overexpressed miRNA-146b-5p on malignant phenotypes of GC. AGS and SGC-7901 cells were transfected with NC mimics+NC, miRNA-146b-5p mimics+NC or miRNA-146b-5p mimics+pcDNA-TRAF6, respectively. **A**, The mRNA level of miRNA-146b-5p. **B**, Protein and mRNA levels of TRAF6. **C**, Migration (magnification 40 \times). Data were expressed as mean \pm SD. ** p <0.01.

Discussion

Despite great advances have been made in the treatment and improved prognosis of GC, its mortality in China remains high^{4,5}. Postoperative recurrence and metastasis are the main causes of GC-induced death. Therefore, it is of great significance to elucidate the mechanism underlying metastasis and recurrence of GC⁶⁻⁸. The occurrence and development of GC are complicated and regulated by epigenetics and genetics, involving multiple factors and genes⁷⁻⁹. In the past, abnormally expressed oncogenes and tumor-suppressor genes were believed to be the key links in the pathogenesis of GC^{9,10}.

MiRNAs are highly conserved non-coding RNAs^{12,13}. They extensively participate in GC development¹⁷. Differences in pathological subtypes and differentiation levels of GC may be attributed to different pathways in which miRNAs are involved^{17,18}. It is reported that miRNA-146b-5p dysregulation is closely linked to malignant development of tumor cells^{21,22}. The strong invasive and metastatic potentials of GC result in the poor prognosis of affected patients. In this paper, our findings uncovered that miRNA-146b-5p was downregulated in GC species than paracancerous ones. GC patients combined lymphatic metastasis and distant metastasis expressed a lower level of miRNA-146b-5p than those without metastases. *In vitro* overexpression of miRNA-146b-5p attenuated proliferation and migratory potentials of GC cells.

MiRNAs negatively regulate gene expression through complementary base pairing. Mature miRNAs specifically recognize target mRNA, and thus result in inhibition of gene expression¹⁵. Differentially expressed miRNAs in different types of tumor may be potential biomarkers^{16,17}. Our results showed that TRAF6 was the direct target binding miRNA-146b-5p. TRAF6 is a member of the adaptor protein family that couples the TNF receptor family to the signaling pathway^{23,24}. It is reported that the activation of the downstream signaling, including NF- κ B pathway, is involved in TRAF6-induced tumorigenesis²⁵. Here, TRAF6 was the target of miRNA-146b-5p, which was responsible for the development of GC regulated by miRNA-146b-5p. Collectively, miRNA-146b-5p suppressed the malignant development of GC through targeting TRAF6.

Conclusions

MiRNA-146b-5p level is negatively correlated to lymphatic metastasis and distant metastasis

rates of GC. It suppresses the malignant development of GC by targeting TRAF6.

Funding Acknowledgements

Key R & D programs in Shandong, China (No. 2017GSF108229).

Conflict of Interests

The authors declare that they have no conflict of interest.

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