

Oral management of adult patients undergoing hematopoietic stem cell transplantation

P. BOLLERO¹, P.C. PASSARELLI², A. D'ADDONA², G. PASQUANTONIO¹,
M. MANCINI¹, R. CONDÒ¹, L. CERRONI¹

¹Department of Clinical Science and Translational Medicine, University of Rome, Tor Vergata, Rome, Italy

²Oral Surgery and Implantology Unit, Catholic University of Sacred Heart, Gemelli Foundation, School of Medicine, Rome, Italy

Abstract. – Chemotherapy and hematopoietic stem cell transplantation (HSCT) are the current treatments for patients with hematological diseases; they result in myelosuppression, and increase the susceptibility of patients to severe infections. The oral cavity is a potential site of complications in HSCT patients, because it is the entrance for agents that can cause systemic infections; it is one of the most frequent locations for side effects deriving from conditioning therapy. The importance of dental pre-chemotherapy and transplant prescription is often stressed, since both therapies depress the immune system and platelets, making each intervention at this stage a high risk. The aim of this article is to review the potential complications of HSCT, and to extrapolate from the scientific literature the treatments and timeframes in which dental therapies can be performed, avoiding important risks for patients.

Key Words:

Stem cell, Transplantation, Oral management, Complications of HSCT, Chemotherapy, Hematopoietic.

Introduction

The feasibility of performing certain dental procedures in patients undergoing hematopoietic stem cell transplantation (HSCT) depends on the overall state of the patient's health, as well as the stage of the disease and/or antineoplastic therapy or HSCT. HSCT is widely used in the treatment of disorders of the hematopoietic system or, most commonly, of malignancies such as lymphoma, myeloma, and leukemia^{1,2}. It represents a valid therapeutic option for aplastic anemia congenital immune deficiency states, lysosomal accumulation disorders (Gaucher's disease) and congenital leukocyte affections (Kostmann's syndrome). This procedure has been successfully extended to patients with solid tumors and autoimmune diseases such as multiple sclerosis^{3,4}.

HSCT is a complex therapeutic procedure that involves the infusion of healthy hematopoietic cells, following a preconditioning therapy that destroys diseased cells and those no longer functioning; the conditioning therapy is an indispensable step in the transplantation procedure. Conditioning involves the administration of high-dose chemo/radiotherapy associated with antiangiogenic, cortisone or monoclonal antibody drugs, according to standardized protocols. High doses of therapeutic drugs and/or radiotherapy cause complications and side effects both direct (due to the same administration) and indirect or secondary to the organ toxicity characteristic of these drugs. The indirect effects of preconditioning occur in the gastrointestinal tract (mucositis), liver, lung, and bladder, as well as in other manifestations such as hemorrhagic cystitis and veno-occlusive liver disease, or severe secondary respiratory deficits such as the development of idiopathic pneumonia. In the most severe cases, there is irreversible multi-organ dysfunction. The aplasia phase is characterized by a severe reduction in immune defenses, as well as a decrease in the concentration of hemoglobin and in peripheral blood-cell counts, which exposes the patient to a high risk of hemorrhage and infection⁵. The aplasia phase of variable duration is followed by the digestion phase, consisting of the resumption of bone marrow function: polymorphonucleate nucleation is achieved at neutrophil counts greater than 500/mm³ (for at least three consecutive days) and at platelet counts greater than 20,000-50,000/mm³ of blood⁶.

Hematopoietic Stem Cells (HSC)

In most countries where allogeneic HSCT is a therapeutic procedure available to most patients in need, families have been so small for many decades that a human leukocyte antigen (HLA)-i-

dental sibling donor is available for less than a third of patients⁷. Some patients cannot be transplanted because of the lack of an HLA-identical donor. Hence, the vast majority of patients have to rely on alternatives: peripheral blood stem cells (PBSC) or bone marrow from unrelated adult donors or frozen umbilical cord blood units (CBUs)⁷. For several years, bone marrow was the most widely used source of HSC; HSC (CD34+) in the bone marrow represent 1.1% of total nucleated cells, equivalent to about $2-3 \times 10^6/\text{kg}$ body weight of a healthy individual⁸. Today, peripheral blood is the most used HSC source for harvesting stem cells in adults, after administration of granulocyte colony-stimulating growth factor. PBSC has been shown to produce earlier engraftment and recovery of granulocytes and platelets compared with bone marrow^{6,8}.

The use of CB provides many theoretical advantages, both for PBSC and spinal cord stem cells, due to the immunity of the cells contained therein. Infant cells are, in fact, primitive stem cells with high regenerative potential when compared with adult stem cells. The main practical advantages of using CB as an alternative source of stem cells are the relative ease with which they can be obtained and cryopreserved, the absence of risks for mother or baby, the reduced likelihood of transmitting infections, particularly CMV (cytomegalovirus), and the ability to store fully tested and HLA-typed transplants, available for immediate use. The volunteer donor, once identified by the register, must still be available for donation, must undergo examination for infection and completion of HLA typing^{9,10}. Pooling CB from two donors whose HLA is closely matched to the recipient has resulted in more rapid engraftment in adults for whom unrelated donors are unavailable⁸.

HSC transplantation is distinguished as one of two types: autologous or allogeneic. Autologous transplantation is utilized to treat chemosensitive malignancies, such as multiple myeloma, non-Hodgkin and Hodgkin lymphoma⁸. Autologous transplantation consists of the reinfusion, after chemotherapy, of a high dose of HSC from the same patient, previously withdrawn and cryopreserved. Emphasis is increasingly placed on the identification of new phenotypic markers that might be relevant for optimizing discrimination between leukemic and normal cells, and, perhaps more importantly, for the detection of minimal residual disease, as well as post-transplantation monitoring of relapse occurrence¹¹. The objective of autotran-

splantation is to treat a malignant disease in the blood or a solid organ, allowing the patient to survive the administration of myeloablative therapy that eradicates the pathology itself. Benefits arise from administering high doses of chemotherapy without compromising bone marrow function irreversibly: reinfusion allows bone marrow recovery in about 15 days. Autologous transplantation restores immunological tolerance by replacing an immune system reactive against self-antigens, and therefore, no longer functioning¹².

Spleen HSCs are extracted by an extracorporeal procedure consisting of multiple aspirations of medullary blood at the level of the iliac ridges (bilaterally), performed under general or spinal anesthesia. Before the infusion is performed, CD34+ cells are counted because the medullary blood undergoes filtration procedures that could reduce the number of HSC in the product being infused. To increase the number of peripheral stem cells, it is possible to resort to a mobilization procedure involving the use of chemotherapies such as cyclophosphamide and/or growth factors. The growth factors used to mobilize HSC in peripheral venous blood are: recombinant human granulocyte colony-stimulating factor (rhG-CSF), recombinant human granulocyte-macrophage colony-stimulating factor (rhGM-CSF) and Flt3 ligand (positive lymphopoietic development modulator)¹³⁻¹⁵. The side effects associated with mobilization with G-CSF are of little magnitude, and generally represented by headache, bone pain and asymptomatic upsurge of alkaline phosphatase and gamma GT¹⁶.

The biological assumptions underlying the clinical outcome of allogeneic transplantation are the eradication of the underlying disease, followed by the recovery of hematopoiesis by infusing the donor HSC. Eradication is entrusted to the pre-transplant conditioning regimes, which determine the total disappearance of the totipotent stem cell compartment of the patient that will be replaced by donor hematopoietic progenitor cells. The purpose of allogeneic transplantation is the complete replacement of the patient's hematopoietic patency with healthy cells obtained from the donor. Allogeneic stem cell donors may be related (sibling) or unrelated, and should have a matching HLA type. Allogeneic transplantation is a potentially curative cellular therapy for hematologic malignancies, many neoplastic and non-systemic hepatic disorders and bone marrow failure syndromes (congenital or acquired medullary aplasia and hemoglobinopathies)^{6,8,16-18}.

Graft Rejection

Extreme polymorphism of the HLA system within the general population makes it possible to find a compatible donor in the household in only 25% of cases, extending to 33% if family members are incompatible with only one locus¹⁹. The main complications of allogeneic HSCT are graft rejection and graft-vs.-host disease (GvHD). Rejection is a rare complication that occurs only in 1% of cases, due to the effect of conditioning regimens that make the patient accept the transplant by depressing the immune system heavily¹. The ever-increasing indication for allogeneic transplantation, and the use of incompatible HLA donors for patients without a compatible donor, make GvHD one of the major causes of transplant-related morbidity and mortality in HSCT recipients. It is more likely to occur if the host receives a graft from an unrelated donor, or if the host or donor is older^{8,20}.

After hematopoietic recovery, donor T-lymphocytes in the graft recognize the host tissues as foreign, and initiate an immune-mediated inflammatory response. This gives rise to GvHD, which is a potentially fatal complication if not treated adequately. This is, in addition, to the direct effects of the disease which are also due to immunosuppressive medicines used for treatment. A certain aspect known as graft-vs.-neoplasia attacks and destroys any remaining cancer cells, and accounts for the highly curative potential of allo-HSCT²¹. One of the major complications of hematopoietic cell transplantation is GvHD differentiation into acute (aGvHD) and chronic (cGvHD) disease in relation to post-transplantation time and various clinical manifestations. By convention, GvHD is considered acute if it arises after the implantation phase (recovery of bone marrow function) within 100 days of transplantation. It is possible to diagnose aGvHD forms even before complete neutrophilization; for this reason, aGvHD is best characterized by clinical manifestations rather than onset timing⁸. cGvHD develops in 25-45% of patients who have received HLA-identical bone marrow transplant, and rises to 50-80% of recipients undergoing allo-HSCT from unrelated donors or HLA-partially compatible family members^{8,18}. The clinical presentation of cGvHD is variable, and may include lichen planus-like changes of skin and mucosa, sclerosis of the skin, sicca syndrome secondary to lacrimal and/or salivary gland damage, liver involvement with cholestasis, and a decrease in pulmonary function secondary to bronchiolitis obliterans²⁰.

Oral involvement may be seen in approximately 80% of patients with cGvHD, and can be a significant source of patient morbidity. Common areas of oral involvement include the tongue and buccal and labial mucosa; oral lesions are often hyperkeratotic, and are considered evident signs for the presence of systemic disease^{22,23}. Patients with salivary gland cGvHD report symptoms of xerostomia, hyposalivation, altered taste, limited opening of the mouth, and difficulty eating and tooth brushing. In addition, salivary gland cGvHD can predispose to the risk of caries determined by reduced antimicrobial activity, compromised buffering capacity and a decreased rate of enamel and dentine remineralization^{18,24}. Another complication of salivary gland cGvHD is the formation of mucoceles²⁵. Immunological reconstitution, starting with the HSC of the donor, takes place in months or years, prolonging the risk of infection over time⁵. In the HSC transplant phase, it being a programmable procedure, the role of the dentist is to evaluate possible odontostomatological pathologies that may interfere with the outcome of the transplant itself, and then to treat all those infectious loci that may cause systemic bacterial transmission, or those pathologies that cause pain and/or discomfort of the oral cavity. GvHD represents, to date, the major complication of allo-HSCT. As a result, timely diagnosis and a multidisciplinary treatment programme are essential for early detection of the oral signs of potential systemic complications, and to improve the quality of life in both adult and paediatric patients^{26,27}.

Materials and Methods

English language literature was searched for references published from August 1997 up to July 2017 in MEDLINE and PubMed, using the following medical subject headings (MeSH) and terms or combinations: leukemia, stem cell transplantation dental, oral, periodontal, hematological patients, chemotherapy, dental treatment, mucositis. Terms were used together with all their known synonyms.

Results

Patients undergoing immunosuppressive and conditioning therapy have an increased risk of developing oral complications: opportunistic infections, gingivitis, mucositis, and ulcerations

of the oral mucosa. No dental treatment before HSCT is related to a higher rate of these complications²⁸. It has been shown that dental treatment prior to chemotherapy/HSCT may prevent the additional death of 1.8/1,000 patients, and reduce systemic infections by 20-25%²⁹. The role of the dentist in HSCT mainly occurs in three different phases. The first is preconditioning dental screening; dental care at this stage is based on treatment priority, and is geared to acute phase problems. Conditioning follows the so-called aplasia phase in which, in addition to a reduction in hemoglobin concentration and the number of circulating platelets, there is a drastic reduction in the patient's immune defenses.

Dental care should be planned based on pre-treatment dental evaluation, and management of complications during immunosuppression and after HSCT. There are considerations to be made and limitations regarding certain dental treatments, especially invasive ones, during the various stages of antineoplastic treatment. A simplified myelosuppression grading is considered a useful tool for understanding the myelosuppressive state caused by chemotherapy. It also facilitates communication between medical and dental staff³⁰. Considering the risk of bleeding and serious infections associated with invasive oral cavity procedures, there are protocols that underline the importance of evaluating some hematologic indices, especially neutrophil and platelet counts. All potential sources of oral infection should be eliminated before the conditioning phase, but time limitations and the patient's condition can interfere with the treatment protocol. In these cases, some treatments should be postponed until the patient is in a good clinical and hematological condition^{1,5,22-24,31}. The US National Cancer Institute³² argues that oral cavity lesions, endodontic lesions, caries, periodontal disease, and implants should be treated at this stage.

Time Limitations

The appropriate duration between referral for assessment and transplant is under discussion, and may differ between authors³³. In the preconditioning phase, all dental treatment needs to be completed before the patient initiates over-peak chemotherapy and immunosuppressive therapy. Neutropenia and thrombocytopenia create a limited period for dental treatment.

All dental treatments should be carried out at least 3 days before the onset of chemotherapy (approximately 10 days before granulocyte count

falls below 500 cells/mm³). When hematologic parameters have values lower than those considered minimal, dental treatments should be postponed. Surgical procedures should be as atraumatic as possible and should be completed at least 7-10 days before the initiation of radio/chemotherapy^{8,33-36}.

A neutrophil count of at least 1000-1500/mm³ and a platelet count of at least 40,000-50,000 cells/mm³ are required to perform invasive procedures (periodontal probing or dental extraction)^{32,37,38}.

Given the high proportion of interproximal and rampant caries in patients with cGvHD, it is recommended that patients should undergo routine dental examinations with bitewing radiographs in the early follow-up period, no later than 6 months to 1 year after allo-HSCT¹⁸. Lastly, in the long-term survival phase, it is necessary to have a video protocol for dentistry inserted in the context of the multidisciplinary approach to the transplant patient. After treatment of acute phases, other procedures may be performed, such as restoring restorations, restoring fractured teeth or prosthetic assessments.

The US National Cancer Institute emphasizes that immune system reconstitution time in transplanted patients may vary from 6 to 12 months, and that routine dental care, including calculus removal procedures, should not be carried out during this period. This is because ultrasounds and other instruments that produce an aerosol can cause pneumonia in these patients, by accidental aspiration of bacteria and debris^{39,40}. Preventive manoeuvres should be used, and antibiotic prophylaxis is required. The use of corticosteroids and/or platelet transfusion is recommended before invasive procedures⁴⁰.

Dental Treatment Needs

Dentists and oral hygienists have important roles in managing the oral health of myelosuppressed patients^{8,30}. The aim of dental assessment and treatment prior to HSCT is to identify potential sources of infection and trauma, and to reduce the morbidity derived from oral complications which themselves can compromise chemotherapy protocols. Many dentists have incomplete knowledge of the disease and the HSCT procedure, the correct dental care that should be provided before and after HSCT, and the dental care provided in hospital⁴¹.

Toljanic et al³⁹ conducted a prospective study to evaluate a pre-chemotherapy dental treatment protocol. The study was conducted on 48 patients

with odontogenic pathologies classified as mild, moderate or severe, considering the probability of developing acute processes during chemotherapy. Acute processes were analyzed on the basis of clinical signs such as edema, abscess and radiographic alterations, whereas the clinical symptoms mainly considered were pain and fever. Acute pathologies were removed before chemotherapy, and in cases of chronic disease, no treatment was done. Odontogenic chronic lesions were present in 79% of patients (of these, 44% were considered serious); 4% showed fever due to odontogenic infection, and antibiotic therapy was administered without interruption of chemotherapy. The authors postulated that patients with mild or moderate chronic lesions can safely undergo chemotherapy without dental treatment, as it has been shown that exacerbation is rare. When it has occurred, patients have been treated effectively without interruption of chemotherapy. The authors argued that it is more important to initiate chemotherapy rapidly because tooth extraction could be a source of a possible infection that would delay the repair of the wound. The authors concluded that the treatment of chronic odontogenic lesions can be safely postponed until the end of chemotherapy, considering the therapeutic benefits.

Melkos et al²⁸ conducted a prospective study in 58 patients undergoing allogeneic or autologous HSCT, analyzing pre-existing odontogenic lesions, dental care and the effect of both on the medical procedure. Patients were divided into two groups: Group I = absence of infectious outbreaks or patients fully treated prior to transplantation, and Group II = presence of infectious outbreaks while undergoing transplantation, without dental surgery. All patients were subjected to dental screening before transplantation, through objective and radiographic examination (panoramic radiographs and periapical radiographs of symptomatic teeth). Dental elements with periapical lesions and periodontal pockets were considered infectious foci. The type of dental treatment prior to transplantation, and the appearance of post-transplant complications (mucositis, infections, GvHD development and disease relapse) were evaluated for an average of 25-75 weeks after the date of transplantation. The protocol before HSCT consisted of conservative treatment of carious lesions, and extraction of non-recoverable teeth. Non-vital teeth and those with periapical lesions were treated by performing endodontic therapy, apicectomy or extraction. Post-HSCT complications were obser-

ved in 75% of patients in Group I and in 95.4% of patients in Group II. The main post-operative infections were cytomegalovirus reactivation and *Staphylococcus epidermidis* bacteraemia. aGvHD was significantly associated with teeth inclusions or with periapical lesions. A higher number of complications was observed in Group II patients, highlighting the importance of oral examination and dental treatment prior to transplantation. The authors concluded that dental treatment prior to HSCT should not be radical, but it should be an integration of restorative and preventive protocols that should be tailored to each individual case.

Yamagata et al¹⁵ conducted a prospective study on 41 patients with hematological malignancies and who were scheduled for HSCT. The authors designed a protocol for minimal intervention, in which the treatment modality was decided according to the severity of the disease, and only symptomatic and heavily compromised teeth were extracted. The dental status was evaluated between 7 and 240 days before HSCT therapy by clinical and radiographic examination. Thirty-six patients required one or more kinds of dental treatment. All therapies were performed up to 10 days before transplantation, without alteration or delay in transplant planning. No patient had signs or symptoms of odontogenic infection during the immunosuppression period. The authors concluded that a conservative protocol seems to be suitable for patients who are to undergo HSCT.

Durey et al³⁶ assessed the treatment needs of 116 patients undergoing pre-HSCT; 93.6% of patients had signs of mucosal and/or dental disease. There was an overall incidence of periodontal disease of 79.5%, ranging from gingivitis to periodontal pockets greater than 6 mm. The authors concluded that assessment within a specialized hospital center has the advantage of providing direct access to an expert dentist and sedation facilities.

Endodontic Treatment

Periapical lesions should be treated because they are a risk factor for acquisition of viridians bacteremia after transplant⁴². The oral bacterial flora changes before and following chemotherapy; an increase of oral colonization with potentially pathogenic microorganisms (*Enterococcus faecalis* and *Candida* spp.) is observed. Root canal infections among autologous and allogeneic HSCT patients can be associated with a substantial increase in morbidity, and with significant impairment of the patient's quality of life²⁵. The

research of Braga-Diniz et al⁴³ investigated the relationship between the epidemiological and clinical profiles of 188 patients before and after HSCT, and the need for endodontic treatment. Patients were subjected to an HSCT conditioning dental regimen based on a thorough clinical and radiographic evaluation. Intraoral periapical and bitewing X-rays were obtained; after evaluation, specific dental treatment was planned and performed. A total of 103 patients were in the pre-transplantation stage, and 85 were in the post-transplantation stage. The systemic conditions of the patients referred for dental treatment were compromised, especially for those in the pre-transplantation stage. The frequencies of endodontic treatment were 24.3% and 24.7% before and after HSCT, respectively, corresponding to 23.2% of the targeted sample.

Periodontal Treatment

Assessment of periodontal and oral hygiene status should be performed in patients undergoing HSCT, because prolonged neutropenia may be associated with oral complications. Laine et al⁴⁰ have shown a higher incidence of febrile episodes during chemotherapy in patients with untreated periodontal disease compared to those with healthy periodontal status⁴⁴. Teeth with advanced periodontal disease may potentially cause bacteremia during neutropenia. In a retrospective study on 77 subjects who received a pre-transplant dental evaluation, Akintoye et al⁴⁵ reported that the high incidence of septicemia (64%) was related to bacteria typically found in the periodontal crevice or oral cavity, and many patients had poor periodontal health during transplantation. Raber-Durlacher et al⁴⁴ studied the correlation between gingivitis/periodontitis and the development of bacteremia during the neutropenia period following HSCT. Eighteen patients were examined and classified into two groups: the first group (28% of total) without periodontal disease (PPD \leq 4 mm and BOP \leq 10%), and the second group consisting of patients with gingivitis (PPD \leq 4 mm and BOP $>$ 10%) or periodontitis (PPD $>$ 4 mm and BOP \geq 10%). Two-thirds (67%) of patients developed bacteremia (diagnosed by blood samples taken twice a week), and Group 2 had more frequent episodes during the neutropenia phase than Group 1. The authors suggested that gingivitis and periodontitis may represent a risk factor for the development of bacteremia, also demonstrated in other studies^{46,47}. Dental/periodontal infections can be misdiagnosed, particularly

when inflammation symptoms are masked by neutropenia.

Septicemia frequently occurs during HSCT, and can be fatal. A loss of integrity in the alveolar bone around the teeth suggests that an intact alveolar barrier is also important in preventing viridans streptococcal bacteremia, which occurs in 61% of bacteremia patients during the period of post-transplant neutropenia⁴². Domestic oral hygiene should be based on the use of chlorhexidine-based mouthwashes, in addition to dental floss and toothbrushes, for the prevention of possible infections.

Protocol

Non-invasive procedures do not require an additional cure, and can be performed at any stage of the disease or chemotherapy; they are classified as Type I (objective examination, X-rays, and oral hygiene instructions) or Type II (simple and atraumatic restorations, prophylaxis and supra-gingival scaling)¹. There are non-surgical procedures (Type III), such as complex restorations, scaling/root planning and endodontic treatments, whose implementation requires special attention before and during chemotherapy, considering the patient's overall health status and the risk/benefit ratio^{46,47}. Execution of invasive treatments such as simple (Type IV) and multiple extractions (Type V) may be performed, but always considering the risk/benefit ratio and the patient's state of health⁴⁸.

In high-risk patients (active phase leukemia with bone marrow suppression), dental surgery is limited to emergency care. In patients at moderate risk (maintenance phase), the peak of myelosuppression is most apparent usually after 14 days of chemotherapy administration. At this stage, dental treatments should be avoided; before or 21 days after the onset of chemotherapy, treatments may be performed.

1. Eliminate potential sources of traumatic injury to the mucous membrane.
 - a) Orthodontic appliances, ill-fitting prostheses, deficient/rough restorations and fractured and/or chapped teeth^{8,49}. Orthodontic devices that do not irritate soft tissue can be maintained if the patient maintains a good oral hygiene status. After 2 years of health from completing the therapy, orthodontic treatment that was discontinued can be restarted^{8,50}.
2. Teeth with mild or moderate caries: restore 5.
3. Residual roots and unrestorable teeth: extract.

4. Symptomatic periodontal tooth or probing on depth > 8 mm or severe periodontitis: extract.
 - a) It is important that the dentist is aware of the signs and symptoms of periodontal disease, since these may be difficult to highlight when the patient is immunosuppressed⁴⁷.
5. Asymptomatic tooth with probing on depth < 8 mm: scaling.
 - a) The patient should undergo scaling and root planning, to check for periodontal disease and related bacterial infection^{38,44,51,52}. Some authors suggest that bleeding caused by scaling and root planning can cause bacteremia^{31,46,47}.
6. Pulpitis: endodontic treatment.
 - a) It should be performed at least 1 week before chemotherapy begins, so that there is enough time to evaluate periapical infection resolution¹.
7. Asymptomatic apical periodontitis > 5 mm: endodontic treatment or extraction (if there is insufficient time)⁵.
8. Symptomatic partially erupted third molar (PEM): extract.
 - a) PEMs can be a source of infection due to pericoronitis; if the gingival tissue partially covering the tooth is a potential factor for infection, it should be surgically removed if hematologic levels allow it. Asymptomatic PEMs do not increase the risk of developing a local infection, even though in 36% of patients local infections frequently occur during HSCT treatment. Complications may be avoided by frequent monitoring of the patient⁵³.

Antibiotic prophylaxis: For invasive procedures such as extraction and periodontal causal treatment that can cause significant bleeding and spread of bacteria in the blood, antibiotic coverage must be performed. Systemic antibiotic prophylaxis reduces the occurrence of febrile episodes, clinically or microbiologically documented infection and bacteremia, without significantly affecting all-cause mortality or infection-related mortality⁵⁴. Antibiotic prophylaxis during oral and maxillofacial surgery should be performed until 6 months after the completion of chemotherapy, even though some authors do not recommend routine antibiotic prophylaxis in patients undergoing chemotherapy^{1,38,52}. Invasive procedures should, therefore, be planned and agreed upon with a multidisciplinary approach³⁸, and patients should be treated with 2 g amoxicillin orally 1 h before treatment³³. Antimicrobial prophylaxis is recommended when the neutrophil count is less

than 2,000 cells/mm³ of blood, or in patients with increased susceptibility to endocarditis^{32,46,55}. When there is an odontogenic infectious outbreak, intravenous antibiotics should be administered to high-risk patients^{48,49}.

Management of oral bleeding: A platelet count of at least 60,000 cells/mm³ is acceptable for oral surgery. When spontaneous bleeding is observed, the dentist should perform oral hygiene motivation procedures. If these measures are not sufficient, platelet transfusion can be considered³². Oral bleeding can be controlled by the use of vasoconstrictor agents (adrenaline), clot-forming agents (topical thrombin and/or hemostatic collagen agents) and tissue protectants: cyanoacrylate products help seal bleeding sites and protect organized clots³². Epsilon-Aminocaproic acid (EACA) may be useful in patients with unstable clots; a notable reduction in post-operative bleeding is evidenced following dental extraction when either EACA or tranexamic acid is used⁵⁶. Tranexamic acid as a topical hemostatic agent is considered effective in reducing the incidence of post-operative bleeding in patients receiving oral anticoagulants; 500 mg tranexamic acid/5 ml saline solution mouthwash minimizes gingival bleeding^{33,52}.

Oral complications of HSCT

Mucositis

Oral mucositis (OM) occurs in most patients treated with high-dose therapy and stem cell transplantation. It has been associated with an increased need for total parenteral nutrition, opioid analgesics and prolonged hospitalization with intensive attendant care^{57,58}. OM is an inflammatory-driven process of the oral mucosa; it is induced by radiation therapy and/or chemotherapy, and is characterized clinically by mucosal damage ranging from mild inflammation (erythema) to extensive ulcerations penetrating the submucosa. Mucositis is associated with an increased risk of systemic infection resulting from bacteremia associated with the breakdown of mucosal barriers. In general, these alterations occur within 2-3 weeks. After completion of radiotherapy, they heal over a similar period of time; resolution coincides with engraftment. Vascular damage and epithelial atrophy make the mucosa susceptible to trauma, and even mild trauma can cause the onset of an ulcer that can take months to heal⁵⁹. The main initiating factor is the presence of reactive oxygen species (ROS) that damage cells, tissues and

blood vessels directly. It is postulated that histamine attenuates tissue damage by decreasing the generation and release of ROS, and suppresses the release of pro-inflammatory cytokines⁶⁰.

In HSCT patients, mucositis is not limited to the oral cavity but may occur along the entire oro-digestive tract²⁵. The prevalence of OM is between 30-70% after chemotherapy, and can reach 90% after HSCT^{61,62}. Conditioning regimens are the most important parameters determining OM risk, but patient-related factors are also involved. Genetic polymorphisms associated with the expression of inflammatory mediators such as tumor necrosis factor (TNF)- α have been implicated in OM risk in patients undergoing allogeneic HSCT⁶³. Identification of populations at higher risk for OM is needed to begin intense prevention for patients undergoing conditioning for allo-HSCT. It is hypothesized that patients who carry the MTHFR 677TT genotype are associated with a higher risk of developing OM, and the use of multivitamins before transplantation has a potential protective effect on the development of OM. Concurrent folic acid supplementation does not change the effectiveness or toxicity of methotrexate in this patient population^{58,64}.

Oral ulcerative mucositis is a significant, common and important risk factor for alpha-hemolytic streptococcal bacteremia in autologous transplant patients with myelosuppression. The extent and severity of OM is significantly correlated with the number of days for which injectable narcotics, total parenteral nutrition, and injectable antibiotics are administered, as well as the risk of significant infection, days spent in hospital and mortality^{57,65}.

A descriptive study was carried out by Recolons et al⁶⁶ on 97 patients who received treatment with chemotherapy or conditioning prior to HSCT; oral hygiene and dental, plaque and gingival index were evaluated and related to mucositis. Patients with a high plaque index presented a high percentage of mucositis. Patients with healthy gums, good oral hygiene and no visible plaque presented a reduced prevalence of mucositis (26.7%). In the study of Coracin et al⁶⁷ on 97 patients (56% male and 44% female) who underwent HSCT, dental plaque and periodontal status, causative factors in oral inflammation, were determinant of the incidence of OM. The grade of mucositis was seen to be influenced by the number of missing teeth and by the DMFS index; this might be due to the trauma determined by tooth friction⁶⁸. The presence of *Porphyromonas gingivalis*, a strictly anaerobic Gram-negative bacterium associated with perio-

dontitis, has been shown to have a predictive value for mucosal ulcerations in HSCT patients⁴⁹.

The presence of plaque and calculus increases the risk of bacteremia, even in neutropenic patients, while good oral hygiene reduces the occurrence and severity of mucositis^{66,69}. Sepsis in 25–50% of immunosuppressed patients derives from the oral cavity, and tooth brushing does not increase bacteremia.

Indications

Dental brushing 2-3 times a day⁶⁶. Some authors recommend replacement of the toothbrush after each neutropenic cycle. Toothpaste containing sodium dodecyl sulfate should be avoided if it irritates the mucosa⁴⁹.

Chlorhexidine mouthwash: not recommended, due to the possibility of irritation of the oral mucosa. In cases of oral infection, non-alcoholic and non-flavored chlorhexidine may be tolerated during neutropenia⁴⁹.

Aloe vera solution: a statistically significant reduction in severe mucositis is found⁷⁰.

Caphosol® + 1% fluoride gel: reduces the frequency and intensity of OM⁷¹.

High-power laser therapy: faster healing of lesions, reduced inflammation, decrease in pain 3 days after the first application, and complete healing after 11 days^{62,72}.

Mucormycosis

Zygomycoses are infections caused by fungi of the class Zygomycetes, comprised of the orders Mucorales and Entomophthorales. Mucormycosis is a fungal infection due to fungi of Mucorales species that may become opportunistic pathogens in immunocompromised patients⁷³, as for example in the case of HSCT. Mucormycosis confined to the periodontium is uncommon, and few cases have been reported⁷⁴. Prolonged and profound neutropenia due to chemotherapy for an underlying hematologic malignancy, and HSCT in recipients with severe GvHD are other significant risk factors for this form of the disease. Mucormycosis is an aggressive and frequently fatal opportunistic fungal infection in these patients, with mortality rates ranging from 69% to 77%⁷⁵. The most common oral sign of mucormycosis is ulceration of the palate. Differential diagnosis may include odontogenic or periodontal infection and maxillary sinusitis. Organisms may be identified on biopsy using periodic acid Schiff stain or methenamine silver stain⁷⁶. Recently, Epstein et al⁷⁶ described

a case of gingival mucormycosis after 25 days of HSCT. The authors emphasize the importance of early detection and treatment of post-transplant oral infections by a multidisciplinary team.

Indications

Amphotericin B deoxycholate: 1-1.5 mg/kg/day.
Granulocyte transfusion: at least 1×10^{10} .

Osteonecrosis and Osteoporosis

Long-term corticosteroid therapy may contribute to the loss of bone density, which may affect alveolar bone and temporomandibular joints, and may be associated with an increased risk of avascular necrosis of bone. Bisphosphonate therapy can increase osteonecrosis of the jaw⁸.

Conclusions

The purpose of this work was to evaluate when perform dental interventions in patients waiting for transplantation, and what interventions can be performed safely. Even if large-scale longitudinal studies are absent, dental treatment is preferred to the strategy of no dental treatment, as underlined by Elad et al²⁹. The oral health of patients undergoing HSCT should be evaluated for oral infections that should be eliminated, before initiation of the conditioning phase. The role of the dental practitioner occurs during the chemotherapy phase and during the phase preceding the transplant, to allow the patient to achieve good oral health, and increase the success rate of the transplant.

The conditions most reported in different studies are about periodontal pathology. This highlights the importance of the dental hygienist collaborating with the dentist; both of them must educate the patient about the importance of maintaining oral health for the prevention of unwanted complications during and after treatment with chemotherapeutic agents. The patient must be informed about any possible dental-related complications that may occur after transplantation. During the various phases of chemotherapy, emergency dental interventions are always possible; the dentist should be assisted by specialist medical staff. A dedicated dental service within the hospital appears to be effective for better communication between the dentist and the oncologist. Dental care cannot always be done in the hospital in a short time, hence the need to create guidelines for the private dentist.

Conflict of Interest

The Authors declare that they have no conflict of interest.

References

- 1) ZIMMERMANN C, MEURER MI, GRANDO LJ, GONZAGA DEL MORAL JA, DA SILVA RATH IB, SCHAEFER TAVARES S. Dental treatment in patients with leukemia. *J Oncol* 2015; 2015: 571739.
- 2) GRATWOHL A, BALDOMERO H, DEMIRER T, ROSTI G, URBANO-ISPIZUA A. Hematopoietic stem cell transplantation for solid tumors in Europe. *Ann Oncol* 2004; 15: 653-600.
- 3) TYNDALL A, GRATWOHL A. Hematopoietic stem and progenitor cells in the treatment of severe autoimmune diseases. *Ann Rheum Dis* 1996; 55: 149-151.
- 4) FASSAS A, PASSWEG JR, ANAGNOSTOPOULOS A, KAZIS A, KOZAK T, HAVRDOVA E, CARRERAS E, GRAUS F, KASHYAP A, OPENSHAW H, SCHIPPERUS M, DECONINCK E, MANCARDI G, MARMONT A, HANSZ J, RABUSIN M, ZUAZU NAGORE FJ, BESALDUCH J, DENTAMARO T, FOUILLARD L, HERTENSTEIN B, LA NASA G, MUSSO M, PAPINESCHI F, ROWE JM, SACCARDI R, STECK A, KAPPOS L, GRATWOHL A, TYNDALL A, SAMIJN J; Autoimmune Disease Working Party of the EBMT (European Group for Blood and Marrow Transplantation). Hematopoietic stem cell transplantation for multiple sclerosis. A retrospective multicentre study. *J Neurol* 2002; 249: 1088-1097.
- 5) YAMAGATA K, ONIZAWA K, YANAGAWA T, HASEGAWA Y, KOJIMA H, NAGASAWA T, YOSHIDA H. A prospective study to evaluate a new dental management protocol before hematopoietic stem cell transplantation. *Bone Marrow Transplant* 2006; 38: 237-242.
- 6) ESH-EBMT Handbook on haematopoietic stem cell transplantation. ESH-EBMT, 2014.
- 7) SAULNIER N, DI CAMPLI C, ZOCCO MA, DI GIOACCHINO G, NOVI M, GASBARRINI A. From stem cell to solid organ. Bone marrow, peripheral blood or umbilical cord blood as favorable source? *Eur Rev Med Pharmacol Sci* 2005; 9: 315-324.
- 8) EPSTEIN JB, RABER-DURLACHER JE, WILKINS A, CHAVARRIA MG, MYINT H. Advances in hematologic stem cell transplant: an update for oral health care providers. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2009; 107: 301-312.
- 9) ZHANG P, FENG K, XUE Y, ZHANG CX, WANG Y, LI XL. Clinical applications of haploidentical hematopoietic stem cell transplantation in severe aplastic anemia. *Eur Rev Med Pharmacol Sci* 2017; 21: 155-161.
- 10) RUBINSTEIN P, CARRIER C, SCARADAVOU A, KURTZBERG J, ADAMSON J, MIGLIACCIO AR, BERKOWITZ RL, CABBAD M, DOBRILA NL, TAYLOR PE, ROSENFELD RE, STEVENS CE. Outcomes among 562 recipients of placental-blood transplants from unrelated donors. *N Engl J Med* 1998; 339: 1565-1577.
- 11) GOBBI G, MIRANDOLA P, MALINVERNO C, SPONZILLI I, CARUBBI C, RICCI F, BINAZZI R, BASSO G, GIULIANI-PICCARI G, RAMAZZOTTI G, PASQUANTONIO G, COCCO L, VITALE M. Aberrant expression of B203.13 antigen in acute

- lymphoid leukemia of B-cell origin. *Int J Oncology* 2008; 33: 371-374.
- 12) RICHARD KB, SHIMON S, WILLIAM HB, ALBERTO M. Mar-mont induction of tolerance in autoimmune diseases by hematopoietic stem cell transplantation: getting closer to a cure? *Blood* 2002; 99: 768-784.
 - 13) ZUMBERG MS, LEATHER HL, NEJAME C, MEYER C, WINGARD JR. GM-CSF versus G-CSF: engraftment characteristics, resource utilization, and cost following autologous PBSC transplantation. *Cytotherapy* 2002; 4: 531-538.
 - 14) GAZITT Y. Comparison between granulocyte colony-stimulating factor and granulocyte macrophage colony stimulating factor in the mobilization of peripheral blood stem cell. *Curr Opin Hematol* 2002; 9: 190-198.
 - 15) SUDO Y, SHIMAZAKI C, ASHIHARA E, KIKUTA T, HIRAI H, SUMIKUMA T, YAMAGATA N, GOTO H, INABA T, FUJITA N, NAKAGAWA M. Synergistic effect of FLT 3 ligand on the granulocyte colony stimulating factor-induced mobilization of hemopoietic stem cells and progenitor cells into blood in mice. *Blood* 1997; 89: 3186-3191.
 - 16) SAUER M, GREWAL S, PETERS C. Hematopoietic stem cell transplantation for mucopolysaccharidoses and leukodystrophies. *Klin Padiatr* 2004; 216: 163-168.
 - 17) ALI F, TARESH S, AL-NUZAILY M, MOK PL, ISMAIL A, AHMAD S. Stem cells differentiation and probing their therapeutic applications in haematological disorders: a critical review. *Eur Rev Med Pharmacol Sci* 2016; 20: 4390-4400.
 - 18) CASTELLARIN P, STEVENSON K, BIASOTTO M, YUAN A, WOO SB, TREISTER NS. Extensive dental caries in patients with oral chronic graft-versus-host disease. *Biol Blood Marrow Transplant* 2012; 18: 1573-1579.
 - 19) SPRANGERS B, VAN WIJMEERSCH B, FEVERY S, WAER M, BILLIAU AD. Experimental and clinical approaches for optimization of the graft versus-leukemia effect. *Nat Clin Pract Oncol* 2007; 4: 404-414.
 - 20) IMANGULI MM, ALEVIZOS I, BROWN R, PAVLETIC SZ, ATKINSON JC. Oral graft-versus-host disease. *Oral Dis* 2008; 14: 396-412.
 - 21) ZAHID MF, ALI N, SHAIKH MU, ADIL SN. Outcome of allogeneic hematopoietic stem cell transplantation in patients with hematological malignancies. *Int J Hematol Oncol Stem Cell Res* 2014; 8: 30-38.
 - 22) SCHUBERT MM, CORREA ME. Oral graft-versus-host disease. *Dent Clin North Am* 2008; 52: 79-109.
 - 23) ELAD S, ZEEVI I, OR R, RESNICK IB, DRAY L, SHAPIRA MY. Validation of the National Institutes of Health (NIH) scale for oral chronic graft-versus-host disease (cGVHD). *Biol Blood Marrow Transplant* 2010; 16: 62-69.
 - 24) BRAND HS, BOTS CP, RABER-DURLACHER JE. Xerostomia and chronic oral complications among patients treated with haematopoietic stem cell transplantation. *Br Dent J* 2009; 207: E17.
 - 25) HAVERMAN TM, RABER-DURLACHER JE, RADEMACHER WM, VOKURKA S, EPSTEIN JB, HUISMAN C, HAZENBERG MD, DE SOET JJ, DE LANGE J, ROZEMA FR. Oral complications in hematopoietic stem cell recipients: the role of inflammation. *Mediators Inflamm* 2014; 2014: 378281.
 - 26) MATURO P, CONDÒ R, COSTACURTA M, DOCIMO R. Oral manifestations in the graft versus host disease in paediatric patients: case report. *Oral Implantol (Rome)* 2009; 2: 34-41.
 - 27) CONDÒ R, MATURO P, PERUGIA C, DOCIMO R. Oral lesions in paediatric patients with graft-versus-host disease. *Eur J Paediatr Dent* 2011; 12: 50-54.
 - 28) MELKOS AB, MASSENKEIL G, ARNOLD R, REICHART PA. Dental treatment prior to stem cell transplantation and its influence on the posttransplantation outcome. *Clin Oral Investig* 2003; 7: 113-115.
 - 29) ELAD S, SHAPIRA MY, MCNEAL S, OR R, GARFUNKEL AA, HIRSCHHORN A, BITAN M, RESNICK I, BENJAMIN G, BARASCH A. Oral effects of nonmyeloablative stem cell transplantation: a prospective observational study. *Quintessence Int* 2008; 39: 673-678.
 - 30) AKASHI M, SHIBUYA Y, KUSUMOTO J, FURUDOI S, INUI Y, YAKUSHIJIN K, OKAMURA A, MATSUOKA H, KOMORI T. Myelosuppression grading of chemotherapies for hematologic malignancies to facilitate communication between medical and dental staff: lessons from two cases experienced odontogenic septicemia. *BMC Oral Health* 2013; 13: 41.
 - 31) HORLIANA AC, CHAMBRONE L, FOZ AM, ARTESE HP, RABELO MDE S, PANNUTI CM, ROMITO GA. Dissemination of periodontal pathogens in the bloodstream after periodontal procedures: a systematic review. *PLoS One* 2014; 9: e98271.
 - 32) NIH. National Cancer Institute (US). Oral complications of chemotherapy and head/neck radiation for health professionals. NIH, 2016.
 - 33) ELAD S, GARFUNKEL AA, OR R, MICHAELI E, SHAPIRA MY, GALILI D. Time limitations and the challenge of providing infection-preventing dental care to hematopoietic stem-cell transplantation patients. *Support Care Cancer* 2003; 11: 674-677.
 - 34) SONIS S, KUNZ A. Impact of improved dental services on the frequency of oral complications of cancer therapy for patients with non-head and neck malignancies. *Oral Surg Oral Med Oral Pathol* 1988; 65: 19-22.
 - 35) BARKER JG. Current practices in the oral management of the patient undergoing chemotherapy or bone marrow transplantation. *Support Care Cancer* 1999; 7: 17-20.
 - 36) DUREY K, PATTERSON H, GORDON K. Dental assessment prior to stem cell transplant: treatment need and barriers to care. *Br Dent J* 2009; 206: E19.
 - 37) HAYTAC MC, DOGAN MC, ANTMEN B. The results of a preventive dental program for pediatric patients with hematologic malignancies. *Oral Health Prev Dent* 2004; 2: 59-65.
 - 38) KOULOCHERIS P, METZGER MC, KESTING MR, HOHLWEG-MAJERT B. Life-threatening complications associated with acute monocytic leukaemia after dental treatment. *Aust Dent J* 2009; 54: 45-48.

- 39) TOLJANIC JA, BEDARD JF, LARSON RA, FOX JP. A prospective pilot study to evaluate a new dental assessment and treatment paradigm for patients scheduled to undergo intensive chemotherapy for cancer. *Cancer* 1999; 85: 1843-1848.
- 40) LAINE PO, LINDOVIST JC, PYRHÖNEN SO, STRAND-PETTINEN IM, TEERENHOVI LM, MEURMAN JH. Oral infection as a reason for febrile episodes in lymphoma patients receiving cytostatic drugs. *Eur J Cancer B Oral Oncol* 1992; 28B: 103-107.
- 41) BOS-DEN BRABER J, POTTING CM, BRONKHORST EM, HUYSMANS MC, BLIJLEVENS NM. Oral complaints and dental care of haematopoietic stem cell transplant patients: a qualitative survey of patients and their dentists. *Support Care Cancer* 2015; 23: 13-19.
- 42) GRABER CJ, DE ALMEIDA KN, ATKINSON JC, JAVAHERI D, FUKUDA CD, GILL VJ, BARRETT AJ, BENNETT JE. Dental health and viridans streptococcal bacteremia in allogeneic hematopoietic stem cell transplant recipients. *Bone Marrow Transplant* 2001; 27: 537-542.
- 43) BRAGA-DINIZ JM, SANTA-ROSA CC, MARTINS RC, SILVA MESE, VIEIRA LO, RIBEIRO SOBRINHO AP. The need for endodontic treatment and systemic characteristics of hematopoietic stem cell transplantation patients. *Braz Oral Res* 2017; 31: e50.
- 44) RABER-DURLACHER JE, LAHEIJ AM, EPSTEIN JB, EPSTEIN M, GEERLIGS GM, WOLFFE GN, BLIJLEVENS NM, DONNELLY JP. Periodontal status and bacteremia with oral viridans streptococci and coagulase negative staphylococci in allogeneic hematopoietic stem cell transplantation recipients: a prospective observational study. *Support Care Cancer* 2013; 21: 1621-1627.
- 45) AKINTOYE SO, BRENNAN MT, GRABER CJ, MCKINNEY BE, RAMS TE, BARRETT AJ, ATKINSON JC. A retrospective investigation of advanced periodontal disease as a risk factor for septicemia in hematopoietic stem cell and bone marrow transplant recipients. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2002; 94: 581-588.
- 46) DALY CG, MITCHELL DH, HIGHFIELD JE, GROSSBERG DE, STEWART D. Bacteremia due to periodontal probing: a clinical and microbiological investigation. *J Periodontol* 2001; 72: 210-214.
- 47) KINANE DF, RIGGIO MP, WALKER KF, MACKENZIE D, SHEARER B. Bacteraemia following periodontal procedures. *J Clin Periodontol* 2005; 32: 708-713.
- 48) LITTLE JW, FALACE DA, MILLER CS, RHODUS NL. Dental management of the medically compromised patient. *Elsevier* 2012; 23: 403-406.
- 49) ELAD S, RABER-DURLACHER JE, BRENNAN MT, SAUNDERS DP, MANK AP, ZADIK Y, QUINN B, EPSTEIN JB, BLIJLEVENS NM, WALTIMO T, PASSWEG JR, CORREA ME, DAHLÖF G, GARMING-LEGERT KU, LOGAN RM, POTTING CM, SHAPIRA MY, SOGA Y, STRINGER J, STOKMAN MA, VOKURKA S, WALLHULT E, YAROM N, JENSEN SB. Basic oral care for hematology-oncology patients and hematopoietic stem cell transplantation recipients: a position paper from the joint task force of the Multinational Association of Supportive Care in Cancer/International Society of Oral Oncology (MASCC/ISOO) and the European Society for Blood and Marrow Transplantation (EBMT). *Support Care Cancer* 2015; 23: 223-236.
- 50) SHELLER B, WILLIAMS B. Orthodontic management of patients with hematologic malignancies. *Am J Orthod Dentofacial Orthop* 1996; 109: 575-580.
- 51) TONG DC, ROTHWELL BR. Antibiotic prophylaxis in dentistry: a review and practice recommendations. *J Am Dent Assoc* 2000; 131: 366-374.
- 52) COSTA FW, RODRIGUES RR, SOUSA LH, CARVALHO FS, CHAVES FN, FERNANDES CP, PEREIRA KM, SOARES EC. Local hemostatic measures in anticoagulated patients undergoing oral surgery: a systematized literature review. *Acta Cir Bras* 2013; 28: 78-83.
- 53) OHMAN D, BJÖRK Y, BRATEL J, KRISTIANSSON C, JOHANSSON P, JOHANSSON JE, BRUNE M, HASSÉUS B. Partially erupted third molars as a potential source of infection in patients receiving peripheral stem cell transplantation for malignant diseases: a retrospective study. *Eur J Oral Sci* 2010; 118: 53-58.
- 54) KIMURA S, AKAHOSHI Y, NAKANO H, UGAI T, WADA H, YAMASAKI R, ISHIHARA Y, KAWAMURA K, SAKAMOTO K, ASHIZAWA M, SATO M, TERASAKO-SAITO K, NAKASONE H, KIKUCHI M, YAMAZAKI R, KAKO S, KANDA J, TANIHARA A, NISHIDA J, KANDA Y. Antibiotic prophylaxis in hematopoietic stem cell transplantation. A meta-analysis of randomized controlled trials. *J Infect* 2014; 69: 13-25.
- 55) ZIMMER A, FREIFELD A. When to use prophylactic antibiotics in neutropenic patients. *Oncology (Williston Park)* 2016; 30: 838-840.
- 56) WATTERSON C, BEACHER N. Preventing perioperative bleeding in patients with inherited bleeding disorders. *Evid Based Dent* 2017; 18: 28-29.
- 57) SONIS ST, OSTER G, FUCHS H, BELLM L, BRADFORD WZ, EDELSBERG J, HAYDEN V, EILERS J, EPSTEIN JB, LEVEQUE FG, MILLER C, PETERSON DE, SCHUBERT MM, SPIJKERVET FK, HOROWITZ M. Oral mucositis and the clinical and economic outcomes of hematopoietic stem-cell transplantation. *J Clin Oncol* 2001; 19: 2201-2205.
- 58) CHAUDHRY HM, BRUCE AJ, WOLF RC, LITZOW MR, HOGAN WJ, PATNAIK MS, KREMERS WK, PHILLIPS GL, HASHMI SK. The incidence and severity of oral mucositis among allogeneic hematopoietic stem cell transplantation patients: a systematic review. *Biol Blood Marrow Transplant* 2016; 22: 605-616.
- 59) SONIS ST, ELTING LS, KEEFE D, PETERSON DE, SCHUBERT M, HAUSER-JENSEN M, BEKELE BN, RABER-DURLACHER J, DONNELLY JP, RUBENSTEIN EB. Perspectives on cancer therapy-induced mucosal injury: pathogenesis, measurement, epidemiology, and consequences for patients. Mucositis Study Section of the Multinational Association for Supportive Care in Cancer; International Society for Oral Oncology. *Cancer* 2004; 100: 1995-2025.
- 60) ELAD S, ACKERSTEIN A, BITAN M, SHAPIRA MY, RESNICK I, GESUNDHEIT B, COHEN Y, DISS O, BARAK D, DRAY L, OR R. A prospective, double-blind phase II study evaluating the safety and efficacy of a topical histamine gel for the prophylaxis of oral mucositis in patients post hematopoietic stem cell transplantation. *Bone Marrow Transplant* 2006; 37: 757-762.

- 61) SCULLY C, EPSTEIN J, SONIS S. Oral mucositis: a challenging complication of radiotherapy, chemotherapy and radiochemotherapy: part I, pathogenesis and prophylaxis of mucositis. *Head Neck* 2003; 25: 1057-1070.
- 62) VITALE MC, MODAFFARI C, DECEMBRINO N, ZHOU FX, ZECCA M, DEFABIANIS P. Preliminary study in a new protocol for the treatment of oral mucositis in pediatric patients undergoing hematopoietic stem cell transplantation (HSCT) and chemotherapy (CT). *Lasers Med Sci* 2017; 32: 1423-1428.
- 63) BOGUNIA-KUBIK K, POLAK M, LANGE A. TNF polymorphisms are associated with toxic but not with aGVHD complications in the recipients of allogeneic sibling haematopoietic stem cell transplantation. *Bone Marrow Transplant* 2003; 32: 617-622.
- 64) ROBIEN K, SCHUBERT MM, YASUI Y, MARTIN P, STORB R, POTTER JD, ULRICH CM. Folic acid supplementation during methotrexate immunosuppression is not associated with early toxicity, risk of acute graft-versus-host disease or relapse following hematopoietic transplantation. *Bone Marrow Transplant* 2006; 37: 687-692.
- 65) RUESCHER TJ, SODEIFI A, SCRIVANI SJ, KABAN LB, SONIS ST. The impact of mucositis on alpha-hemolytic streptococcal infection in patients undergoing autologous bone marrow transplantation for hematologic malignancies. *Cancer* 1998; 82: 2275-2281.
- 66) SABATER RECOLONS MDEL M, LÓPEZ J, RODRÍGUEZ DE RIVERA CAMPILLO ME, CHIMENOS KÜSTNER E, CONDE VIDAL JM. Buccodental health and oral mucositis. Clinical study in patients with hematological diseases. *Med Oral Patol Oral Cir Bucal* 2006; 11: E497-E502.
- 67) CORACIN FL, SANTOS PS, GALLOTTINI MH, SABOYA R, MUISQUEIRA PT, BARBAN A, CHAMONE DDE A, DULLEY FL, NUNES FD. Oral health as a predictive factor for oral mucositis. *Clinics (Sao Paulo)* 2013; 68: 792-796.
- 68) HERNÁNDEZ-FERNÁNDEZ A, OÑATE-SÁNCHEZ RE, CABRERIZO-MERINO MC, DE ARRIBA-DE LA-FUENTE F, HERAS-FERNANDO I, VICENTE-GARCÍA V. Influence of oral health on mucositis in patients undergoing hematopoietic progenitor cell transplantation (HPCT). *Med Oral Patol Oral Cir Bucal* 2012; 17: e94-e101.
- 69) EPSTEIN JB, HANCOCK PJ, NANTEL S. Oral candidiasis in haematopoietic cell transplantation patients: an outcome-based analysis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003; 96: 154-163.
- 70) SU CK, MEHTA V, RAVIKUMAR L, SHAH R, PINTO H, HALPERN J, KOONG A, GOFFINET D, LE QT. Phase II double-blind randomized study comparing oral aloe vera versus placebo to prevent radiation-related mucositis in patients with head-and-neck neoplasms. *Int J Radiat Oncol Biol Phys* 2004; 60: 171-177.
- 71) PAPAS AS, CLARK RE, MARTUSCELLI G, O'LOUGHLIN KT, JOHANSEN E, MILLER KB. A prospective, randomized trial for the prevention of mucositis in patients undergoing hematopoietic stem cell transplantation. *Bone Marrow Transplant* 2003; 31: 705-712.
- 72) GOBBO M, OTTAVIANI G, BUSSANI R, POZZATO G, BIASOTTO M. Methotrexate-induced oral mucositis in rheumatoid arthritis disease: therapeutic strategy in a case report photonics. *Laser Med* 2013; 2: 71-76.
- 73) SPELLBERG B, EDWARDS J JR, IBRAHIM A. Novel perspectives on mucormycosis: pathophysiology, presentation, and management. *Clin Microbiol Rev* 2005; 18: 556-569.
- 74) McDERMOTT NE, BARRETT J, HIPPI J, MERINO MJ, RICHARD LEE CC, WATERMAN P, DOMINGO DL, WALSH TJ. Successful treatment of periodontal mucormycosis: report of a case and literature review. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2010; 109: e64-e69.
- 75) AL AKHRASS F, DEBIANE L, ABDALLAH L, BEST L, MULANOVICH V, ROLSTON K, KONTTOYIANNIS DP. Palatal mucormycosis in patients with hematologic malignancy and stem cell transplantation. *Med Mycol* 2011; 49: 400-405.
- 76) EPSTEIN JB, KUPFERMAN SB, ZABNER R, REJALI A, HOPP ML, LILL M, TZACHANIS D. Early diagnosis and successful management of oral mucormycosis in a hematopoietic stem cell transplant recipient: case report and literature review. *Support Care Cancer* 2016; 24: 3343-3346.