

# Polyoxometalate SbW9 regulates proliferation and apoptosis of NSCLC cells via PTEN-dependent AKT signaling pathway

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**Abstract.** – **OBJECTIVE:** To explore the influence of polyoxometalate SbW9 on proliferation and apoptosis of non-small cell lung cancer (NSCLC) cells and its mechanism.

**MATERIALS AND METHODS:** NSCLC cell lines A549 and PC9 were treated with 50  $\mu$ M polyoxometalate. Then, the proliferation of NSCLC cells was detected via 2,3-bis(2-methoxy-4-nitro-5-sulfophenyl)-5-[(phenylamino)carbonyl] tetrazolium hydroxide (XTT) assay and colony formation assay; the apoptosis of NSCLC cells was detected via flow cytometry and terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL); and the expression of apoptosis-related proteins, B-cell lymphoma 2 (Bcl-2) and Bcl-2 associated X protein (Bax), was detected via Western blotting. Moreover, the protein expression levels of phosphatase and tensin homolog deleted on chromosome ten (PTEN), phosphorylated-protein kinase B (p-AKT) and total AKT (T-AKT) were detected via Western blotting.

**RESULTS:** The polyoxometalate inhibited the proliferation of A549 and PC9 cells in a concentration-dependent manner (50  $\mu$ M) ( $p < 0.05$ ), and it (50  $\mu$ M) also inhibited the proliferation of both cells in a time-dependent manner (0/2 h) ( $p < 0.05$ ). The results of colony formation assay revealed that the polyoxometalate (50  $\mu$ M) could significantly inhibit the colony formation of A549 and PC9 cells ( $p < 0.05$ ). The results of flow cytometry and TUNEL staining showed that the polyoxometalate (50  $\mu$ M) significantly induced the apoptosis of A549 and PC9 cells ( $p < 0.05$ ). According to further Western blotting, polyoxometalate (50  $\mu$ M) inhibited the expression of anti-apoptotic gene Bcl-2 and promoted the expression of pro-apoptotic gene Bax. Besides, the Western blotting results manifested that the polyoxometalate could activate the expression of PTEN and inhibit the phosphorylation of downstream AKT ( $p < 0.05$ ).

**CONCLUSIONS:** The polyoxometalate can activate the expression of PTEN to inhibit the phos-

phorylation of AKT, ultimately inhibiting the proliferation and inducing the apoptosis of NSCLC cells. Therefore, the polyoxometalate is expected to become a novel drug for the clinical treatment of NSCLC.

**Keywords:**

NSCLC, Proliferation, Apoptosis, Polyoxometalate.

## Introduction

Lung cancer is one of the most common causes of cancer death in the world<sup>1,2</sup>. The number of deaths from non-small cell lung cancer (NSCLC) accounts for about 85-90% in the total from all types of lung cancer<sup>3</sup>. The therapeutic strategies of NSCLC include operation, radiotherapy, chemotherapy, targeted therapy, and combined therapy<sup>4</sup>. Although the potential chemotherapeutic efficacy of novel compounds on NSCLC has been confirmed in many studies, the sensitivity of high-grade NSCLC to chemotherapeutic drugs remains poor, and the exact mechanism of such a phenomenon has not been fully clarified<sup>5</sup>.

The expression of phosphatase and tensin homolog deleted on chromosome ten (PTEN) is inhibited in a variety of tumor tissues, and it has been proved to be a cancer suppressor gene<sup>6</sup>. PTEN is a major inhibitor of phosphoinositide 3-kinase (PI3K), which, therefore, exerts an important regulatory effect on the PI3K/AKT signaling pathway and subsequent cellular biological behaviors<sup>7</sup>. The PI3K/AKT signaling pathway plays an important role in regulating cell survival, growth, differentiation, apoptosis, and autophagy<sup>8</sup>. The activated PI3K can mediate the phos-

phorylation of AKT Thr 308 (catalytic domain) and Ser 473 (regulatory domain), and once AKT is activated, it can regulate a variety of important life activities, such as apoptosis and proliferation<sup>9</sup>.

The polyoxometalate, as a transition metal-oxygen cluster, has the structural charges and size that can be regulated and possesses the potential to produce organic-inorganic hybrids<sup>10</sup>. Based on these chemical structures, researchers combined some biomolecules with polyoxometalate to form specific biomacromolecules, obtaining good effects in the treatment of disease. In this study, the effects of polyoxometalate SbW9 on proliferation and apoptosis of NSCLC cells were detected, and the molecular mechanism of SbW9 in affecting the proliferation and apoptosis of NSCLC was explored.

## Materials and Methods

### Materials and Cells

NSCLC cell lines A549 and PC9 were purchased from Shanghai Kanglang Biological Technology Co., Ltd. (Shanghai, China), and SbW9 was synthesized by the Chemical Research Institute of the Chinese Academy of Science. Fetal bovine serum (FBS) was purchased from Gibco (Rockville, MD, USA). Roswell Park Memorial Institute-1640 (RPMI-1640) medium containing penicillin and streptomycin was purchased from Gibco (Rockville, MD, USA). The cells were cultured in the medium at 37°C.

### Cell Intervention

NSCLC cells A549 and PC9 were inoculated into a 96-well plate ( $1 \times 10^4$ /mL) and treated with SbW9 in different concentrations (0.1, 0.5, 10, 20, 50 and 100  $\mu$ M) after 48 h. After treatment for different time, 50  $\mu$ L 2,3-bis-(4-methoxy-5-nitrophenyl)-5-[(phenylamino)carbonyl]-2H-tetrazolium hydroxide (XTT) was added for incubation for 2 h, and the absorbance was read at 450 nm.

### Western Blotting

After treatment of the two kinds of NSCLC cells with SbW9 for 72 h, the total protein was extracted, and the specific steps are as follows: (1) The culture solution in the medium was discarded and cells were washed with PBS for 3 times. (2) 1000  $\mu$ L lysis buffer was added into every dish and fully vibrated for 20 min. (3) The cells at the bottom of the dish were scraped off using a brush and placed into the Eppendorf (EP) tube. (4) The

cells collected were lysed using an ultrasonic pyrolyser for about 15 s. (5) After standing for 15 min, the cells were centrifuged at 12000 rpm for 0.5 h. (6) The supernatant was taken and placed into the EP tube, the protein concentration was detected via ultraviolet spectrometry, and all the protein samples were quantified to be the same concentration. (7) The protein was sub-packaged and placed in the refrigerator at -80°C. After the total protein was extracted from NSCLC cells, Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) was performed. Then, the protein gel was transferred to a polyvinylidene difluoride (PVDF) membrane (Merck, Basel, Switzerland) and incubated with primary antibody at 4°C overnight. Then, it was incubated again with the goat anti-rabbit secondary antibody in a dark place for 1 h. The protein band was scanned and quantified using the Odyssey scanner, and the level of protein to be detected was corrected using glyceraldehyde-3-phosphate dehydrogenase (GAPDH).

### Colony Formation Assay

NSCLC cells in each group were cultured till logarithmic growth phase, and digested with 0.25% trypsin into single cell suspension (the proportion of viable cells >95%). Then, the suspension was inoculated into a 6-well plate (about 500 cells/well) and added with 2 mL 1640 medium per well, and the medium was replaced once every 48 h. After 10 d, the cells were fixed with formaldehyde and stained with crystal violet, and the number of colonies in each well was counted.

### Detection of Apoptosis Via Flow Cytometry

The cells in the logarithmic growth phase were taken, digested with 0.25% trypsin-EDTA (ethylenediaminetetraacetic acid) into the cell suspension, and inoculated into the 6-well plate. The sample was loaded and the apoptosis rate was detected according to the operation steps of the Annexin V-FITC PI (Propidium Iodide) apoptosis assay kit (Beyotime, Shanghai, China).

### Terminal Deoxynucleotidyl Transferase-Mediated dUTP Nick End Labeling (TUNEL) Staining

The cells on the slide were fixed in fixing solution for 1 h and washed with PBS for 3 times. After transparentization, TUNEL reagent was prepared and two negative controls were set. After staining, the cells were observed, photographed and counted under a fluorescence microscope.

### Statistical Analysis

Statistical Product and Service Solutions (SPSS) 22.0 software (IBM, Armonk, NY, USA) was used for the analysis of all data. Measurement data were expressed as mean  $\pm$  standard deviation, and the *t*-test was used for the comparison of data between the two groups.  $p < 0.05$  suggested that the difference was statistically significant.

## Results

### Polyoxometalate SbW9 Inhibited Proliferation of NSCLC Cells in a Concentration-Dependent Manner

After treatment of two kinds of NSCLC cell lines with SbW9 in different concentrations (0.1, 1, 5, 10, 20, 50, and 100  $\mu\text{M}$ ) for 72 h, the cell proliferation in each group was detected via XTT assay. As shown in Figure 1, SbW9 (5, 10, 20, 50, and 100  $\mu\text{M}$ ) could significantly inhibit the proliferation of A549 and PC9 cells in a concentration-dependent manner ( $p < 0.05$ ). SbW9 in a concentration of 0.1 and 1  $\mu\text{M}$  had no influence on the proliferation of NSCLC cells ( $p > 0.05$ ). Therefore, in the subsequent experiments, SbW9 in a concentration of 50  $\mu\text{M}$  was selected for verification.

### Polyoxometalate SbW9 Inhibited Proliferation of NSCLC Cells in a Time-Dependent Manner

As shown in Figure 2, the proliferation of A549 cells significantly decreased after treatment with SbW9 (50  $\mu\text{M}$ ) at 24, 36, and 72 h compared with that in control group ( $p < 0.05$ ). SbW9 (50  $\mu\text{M}$ ) could significantly inhibit the proliferation of

PC9 cells at 12, 24, 36, 48, and 72 h. Therefore, SbW9 in a concentration of 50  $\mu\text{M}$  was selected in the subsequent experiments, and NSCLC cells were stimulated for 72 h.

### Polyoxometalate SbW9 Inhibited Formation of NSCLC Cells

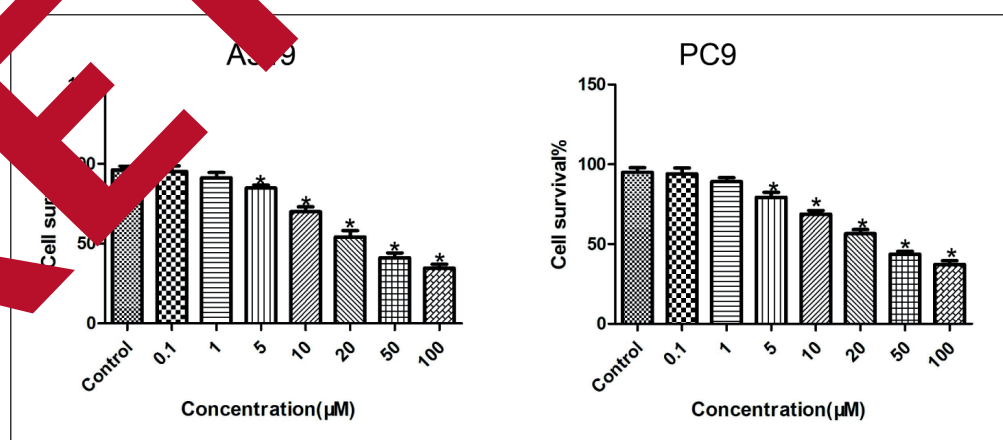
Furthermore, the proliferation of two kinds of NSCLC cell lines was detected via colony formation assay. The results revealed that in A549 cell lines, the number of colonies in SbW9 group ( $44.82 \pm 6.32$ ) was significantly smaller than that in control group ( $220.11 \pm 11.46$ ) ( $p < 0.05$ ). The same was true in PC9 cells ( $40.05 \pm 8.77$ ) vs. ( $199.83 \pm 10.32$ ) ( $p < 0.05$ ) (Figure 3).

### Polyoxometalate SbW9 Induced Apoptosis of NSCLC Cells

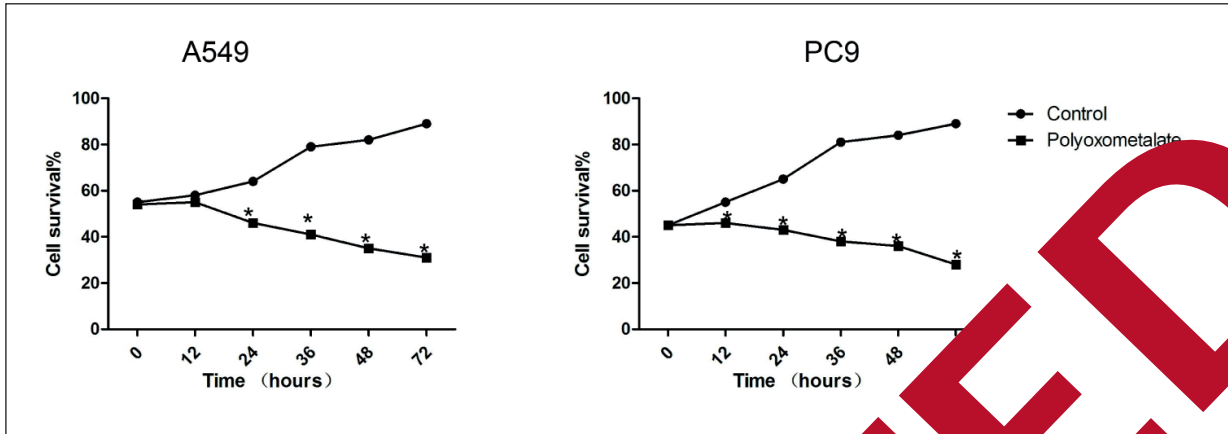
The results of flow cytometry showed that the polyoxometalate SbW9 exerted a significant pro-apoptotic effect on both A549 and PC9 cells. The apoptosis rate in control group and SbW9 group in two kinds of cell lines was ( $1.69 \pm 0.34\%$  vs.  $4.78 \pm 2.11\%$ ) and ( $1.49 \pm 0.34\%$  vs.  $4.78 \pm 2.11\%$ ), respectively ( $p < 0.05$ ) (Figure 4), indicating that the polyoxometalate SbW9 can induce the apoptosis of NSCLC cells.

### Influence of Polyoxometalate SbW9 on Apoptosis-Related Proteins in NSCLC Cells

The results of Western blotting showed that after treatment with polyoxometalate SbW9, the expression of pro-apoptotic gene Bax was up-regulated, while the expression of anti-apoptotic gene Bcl-2 was significantly inhibited in the two



**Figure 1.** Polyoxometalate SbW9 inhibits proliferation of NSCLC cells in a concentration-dependent manner. Control: control group, \* $p < 0.05$ : There is a statistically significant difference compared with control group.



**Figure 2.** Polyoxometalate SbW9 inhibits proliferation of NSCLC cells in a time-dependent manner. Control: control group, Polyoxometalate: polyoxometalate SbW9 group, \* $p < 0.05$ : There is a statistically significant difference compared with control group.

kinds of cell lines ( $p < 0.05$ ) (Figure 5), suggesting that the inducible effect of polyoxometalate SbW9 on apoptosis of NSCLC cells is dependent on the Bcl-2 pathway.

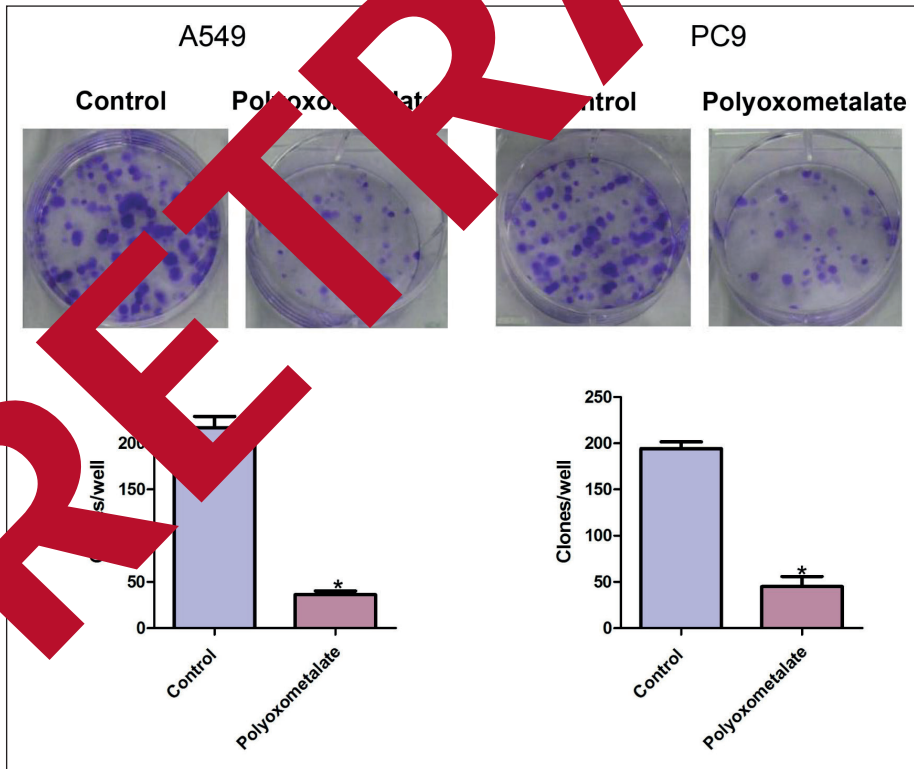
apoptosis rate in control group and SbW9 group in A549 and PC9 cells was  $(2.34 \pm 0.77)\%$  vs.  $(18.21 \pm 1.12)\%$  and  $(3.81 \pm 1.09)\%$  vs.  $(24.77 \pm 1.92)\%$ , respectively ( $p < 0.05$ ).

**TUNEL Staining of NSCLC Cells in Each Group**

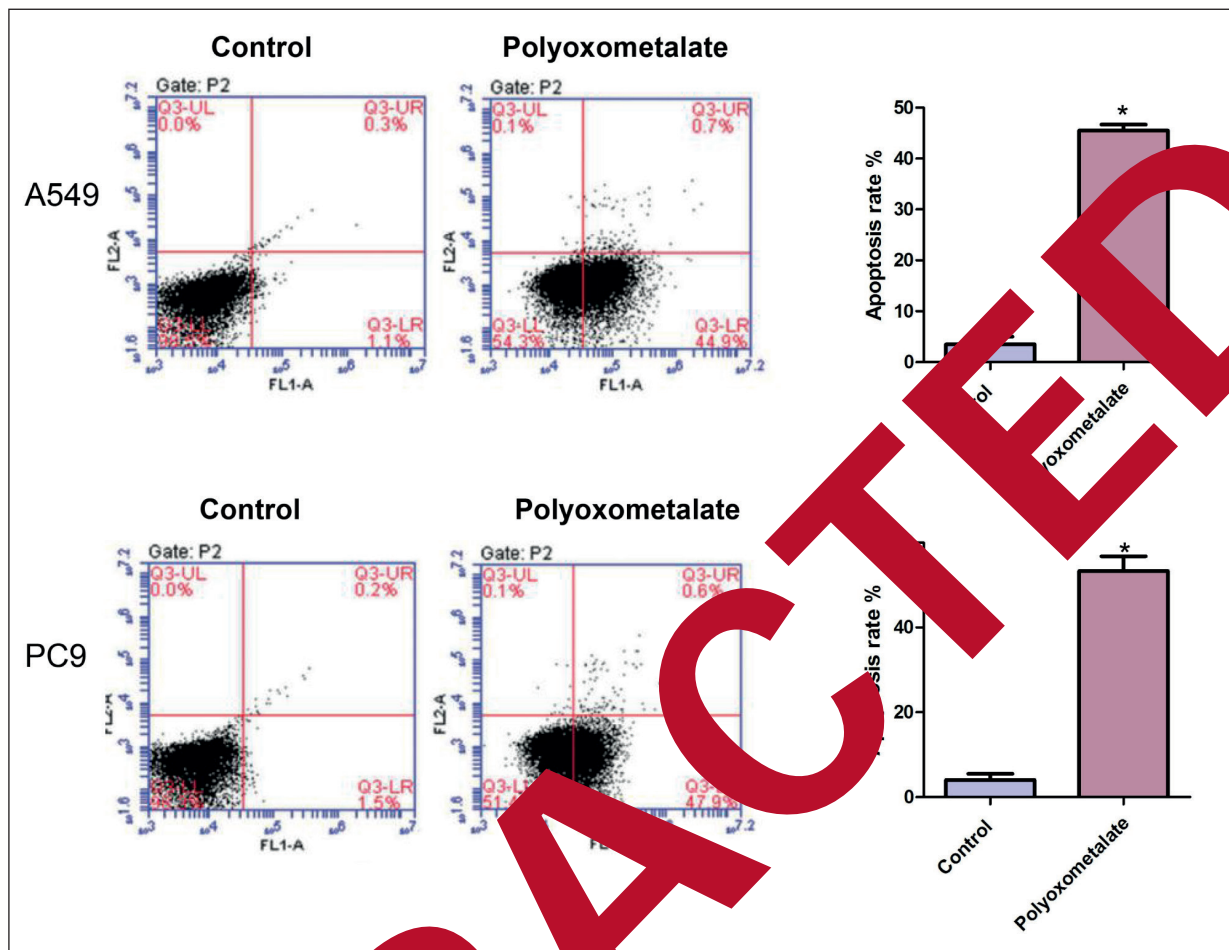
To further detect the apoptosis in each cell line more intuitively, TUNEL staining was performed for cells in each group and the TUNEL-positive cells were counted. As shown in Figure 6, the

**Influence of Polyoxometalate SbW9 on PTEN/AKT Signaling Pathway in NSCLC**

Considering the important role of the PTEN/AKT signaling pathway in the tumor, whether the influence of polyoxometalate SbW9 on biological



**Figure 3.** Polyoxometalate SbW9 inhibits colony formation of NSCLC cells. Control: control group, Polyoxometalate: polyoxometalate SbW9 group, \* $p < 0.05$ : There is a statistically significant difference compared with control group.



**Figure 4.** Polyoxometalate SbW9 promotes apoptosis of lung cancer cells. Control: control group, Polyoxometalate: polyoxometalate SbW9 group, \* $p < 0.05$ : There is a statistically significant difference compared with control group.

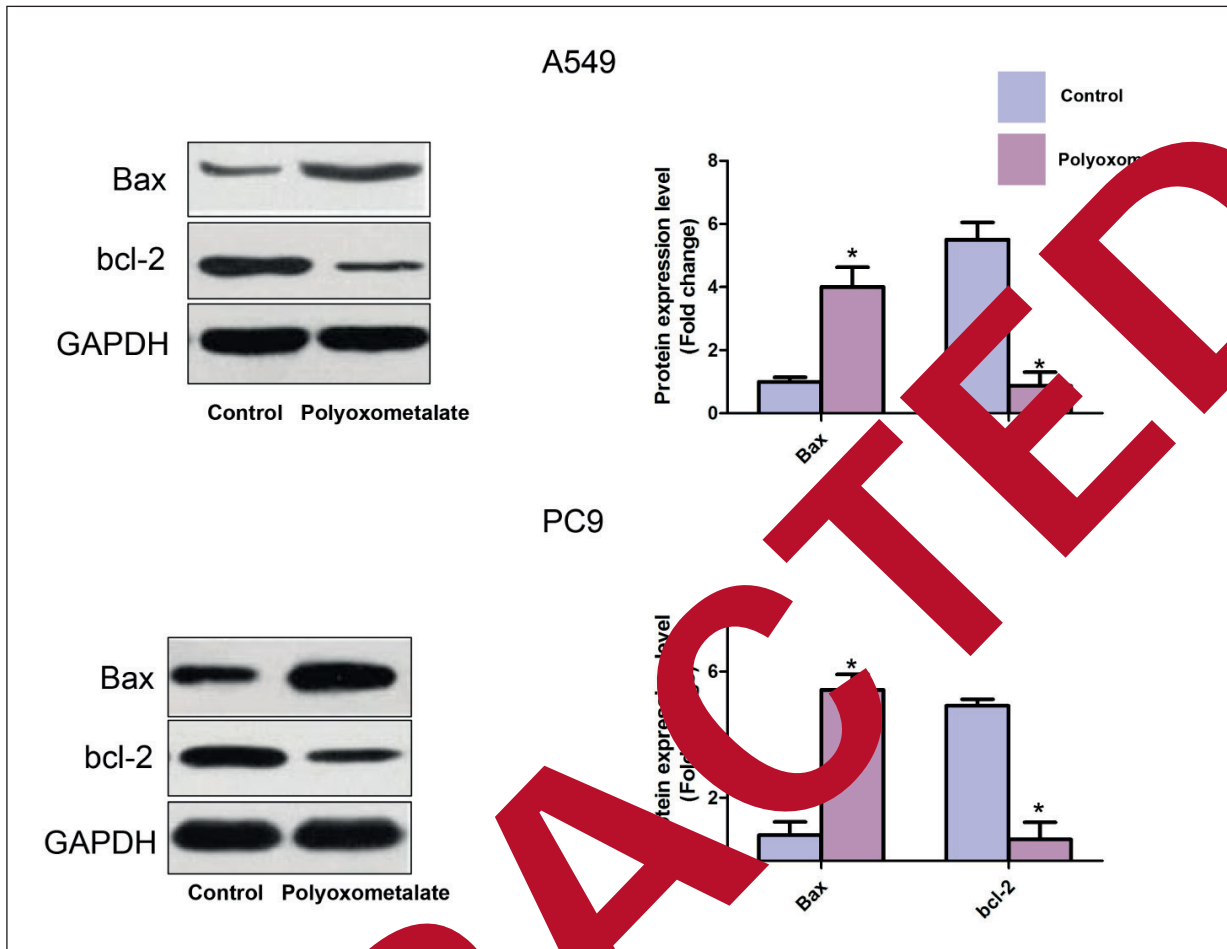
behaviors of NSCLC cells is dependent on the PTEN/AKT signaling pathway was detected. The expression of PTEN and AKT in each group was detected, and it was found that the polyoxometalate SbW9 could activate the PTEN expression and inhibit the phosphorylation level of AKT in the two types of cell lines ( $p < 0.05$ ) (Figure 7), indicating that the PTEN/AKT signaling pathway plays an important role in the anti-proliferative and pro-apoptotic effects of polyoxometalate SbW9.

### Discussion

Lung cancer is one of the most common and severe cancers, and the number of deaths from lung cancer is larger than the total number of deaths from breast cancer, colon cancer, and prostate cancer every year<sup>11</sup>. SCLC and NSCLC are

two major types of lung cancer. Despite the great progress in the diagnosis and treatment of lung cancer, neither treatment nor prognosis of lung cancer is satisfactory, which is related to the high recurrence and mortality rates<sup>12,13</sup>. There is increasing evidence that the proliferation and apoptosis of lung cancer cells have a great influence on the incidence and prognosis of lung cancer<sup>14</sup>. Therefore, inhibiting the proliferation and promoting the apoptosis of lung cancer cells in time and effectively are of great significance in delaying the progression of lung cancer.

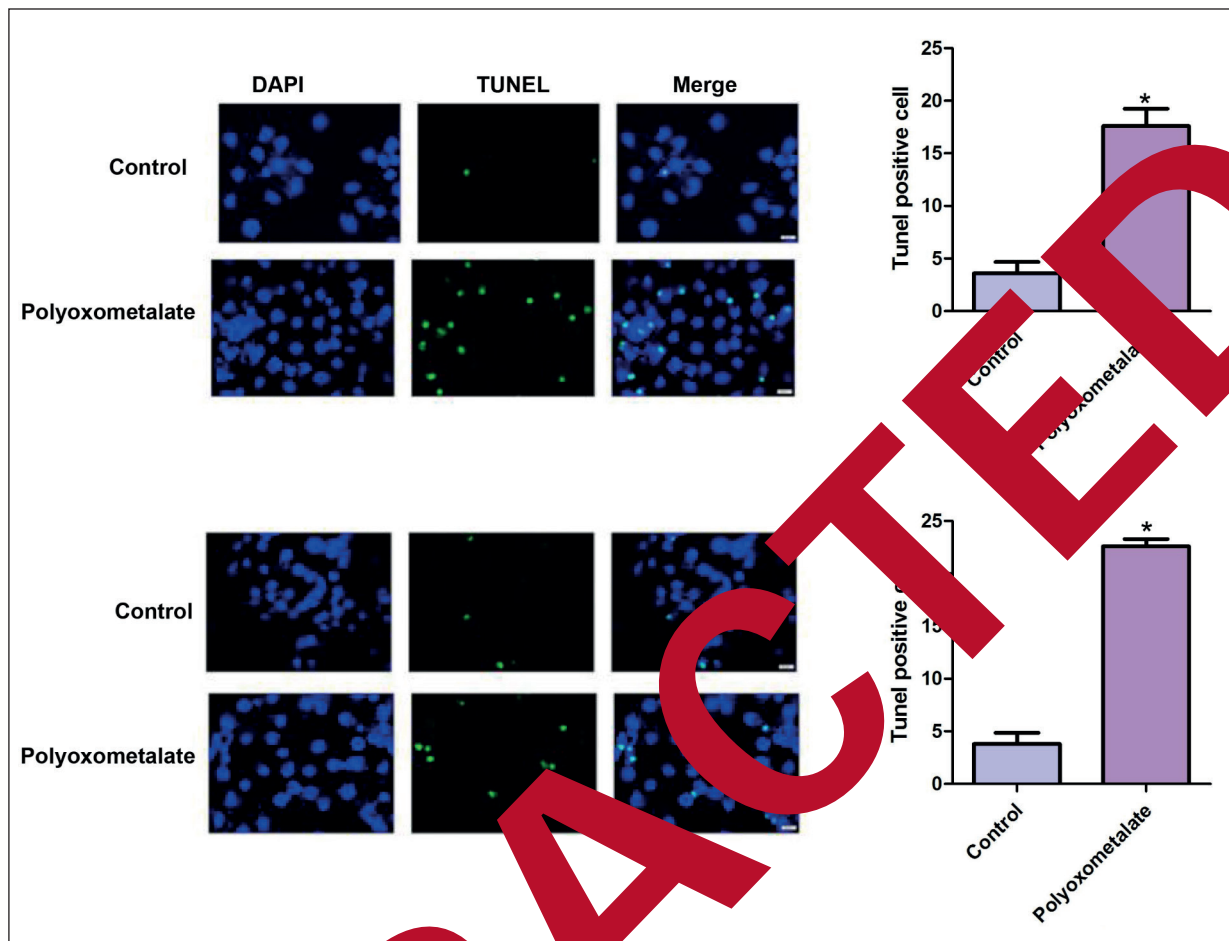
Apoptosis is mediated through the death receptor pathway (exogenous) or the mitochondrial pathway (endogenous), thereby eliminating the damaged cells and keeping the homeostasis<sup>15</sup>. In both pathways, poly ADP-ribose polymerase-1 is cleaved and caspase 3/7 is activated to respond to DNA damage stress in cells<sup>16</sup>. Therefore, target-



**Figure 5.** Influence of polyoxometalate SbW9 on apoptosis-related proteins in NSCLC cells. Control: control group, Polyoxometalate: polyoxometalate SbW9 group, \* $p < 0.05$ : There is statistically significant difference compared with control group.

ing the program of cell death through regulating apoptosis and autophagy has become one of the promising methods in cancer therapy. The Bcl-2 family protein is the regulator of apoptosis, and many members can regulate apoptosis, such as Bax and Bak, important pathways for caspase-mediated cell death. The Bcl-2/Bax ratio is an important determinant of apoptosis<sup>17,18</sup>. Currently, the activation of the PI3K/AKT signaling pathway has been observed in various tumors, and the activation of this pathway can promote the survival and proliferation of cancer cells<sup>19</sup>. PTEN negatively regulates the AKT signaling pathway through dephosphorylation of PDK3 to produce PIP2. There is often a genetic mutation in the PTEN gene in cancer tissues, and its protein expression is inhibited. Besides, PTEN can also negatively regulate the cell proliferation<sup>20</sup>. For example, in prostate cancer cells,

PTEN can inhibit the cell proliferation and promote the apoptosis through down-regulating the IGF-IR expression on cell membrane<sup>21</sup>. In addition, in pancreatic cancer cells, IGF-1-mediated inhibition on PTEN can enhance the cell invasion and proliferation through activating the PI3K/AKT signaling pathway<sup>22</sup>. In NSCLC, the low expression of PTEN and excessive phosphorylation of AKT have close correlations with the poorer prognosis of NSCLC patients<sup>23</sup>. In this study, two kinds of NSCLC cell lines were stimulated with polyoxometalate SbW9 in different doses for different time in *in-vitro* experiments. It was found in XTT assay and colony formation assay that the polyoxometalate SbW9 (50  $\mu$ M, 72 h) significantly inhibited the proliferation and colony formation of the two kinds of NSCLC cell lines. At the same time, the apoptosis in each group was further detected *via* flow cytometry and TUNEL



**Figure 6.** TUNEL staining of NSCLC cells in each group. Control: control group, Polyoxometalate: polyoxometalate SbW9 group, \* $p < 0.05$ : There is a statistically significant difference compared with control group.

staining, and the results showed that the polyoxometalate SbW9 could well induce apoptosis of NSCLC cells. Furthermore, the Western blotting showed that the inhibitory effect of polyoxometalate SbW9 on apoptosis of NSCLC cells may be dependent on the Bcl-2 pathway. Finally, the expression of the classical PTEN/AKT signaling pathway was detected, and it was found that the polyoxometalate SbW9 could effectively enhance the protein expression of PTEN and significantly inhibit the phosphorylation of AKT, thus explaining the molecular mechanism of anti-proliferative effect of polyoxometalate SbW9 on lung cancer cells. Moreover, the experiments have revealed that the reason for the low expression of PTEN is closely related to the methylation in its promoter region<sup>24</sup>. Therefore, it is speculated that the regulatory effect of polyoxometalate SbW9 on proliferation and apoptosis of NSCLC cells may be related to its inhibition on PTEN gene methylation, and more

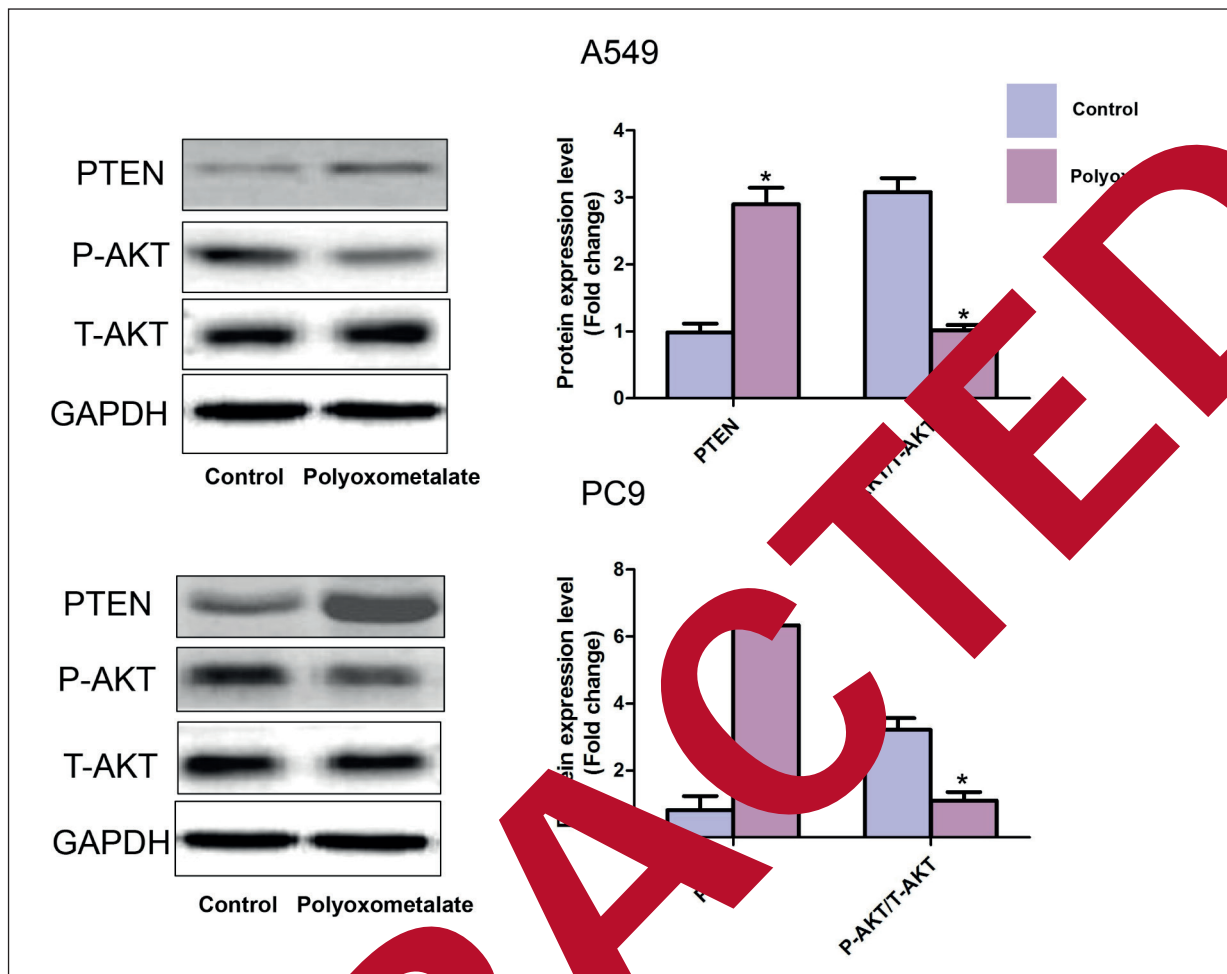
in-depth experiments are needed for verification. However, there were still some deficiencies in this experiment: 1) The animal experiments were not designed, and 2) the anti-cancer effect of the polyoxometalate SbW9 was not verified using PTEN or AKT inhibitors.

## Conclusions

This investigation revealed the anti-proliferative and pro-apoptotic effects of polyoxometalate SbW9 on NSCLC cells for the first time, whose mechanism may be mediated by the PTEN/AKT signaling pathway.

## Conflict of Interests

The Authors declare that they have no conflict of interests.



**Figure 7.** Influence of polyoxometalate SbW12 on PTEN/Akt signaling pathway in NSCLC. Control: control group, Polyoxometalate: polyoxometalate SbW12 group, \* $p < 0.05$ . There is a statistically significant difference compared with control group.

### References

- 1) AKHURST T. Imaging of non-small-cell lung cancer. *PET Clin* 2018; 13: 1-10.
- 2) ZHANG H, LI XY, WANG ZH, LIAN ZF, ZHAO YH. MiR-221 inhibits lung cancer cell EMT and invasion through targeting Snail. *Eur Rev Med Pharmacol* 2017; 83: 3598-3604.
- 3) MATHIAS SS, YANG JC, LEE CK, KURATA T, KIM DW, JOHNSON GAMI N, CHOI Y, MANN H, RUKAZENKOV Y, GHORGHIAN M, PATSONI M, MARKOVETS A, BARRETT JC, THRESS S, JANNE PA. Erlotinib as first-line treatment of EGFR mutation-positive advanced non-small-cell lung cancer. *J Clin Oncol* 2018; 36: 841-849.
- 4) HORN L, GOVINDAN R, ANDERS RA, ANTONIA SJ, SAGOLSKY S, DAVIES MJ, DUBINETT SM, FERRIS A, GANDHI L, GARON EB, HELLMANN MD, HIRSCH FR, MALIK S, NELSON JW, PAPADIMITRAKOPOULOU VA, RIMM DL, SCHWARTZ LH, SEPELI B, YEAP BY, RIZVI NA, HERBST RS. The Society for Immunotherapy of Cancer consensus statement on immunotherapy for the treatment of non-small cell lung cancer (NSCLC). *J Immunother Cancer* 2018; 6: 75.
- 5) YIN H, MA J, CHEN L, PIAO S, ZHANG Y, ZHANG S, MA H, LI Y, QU Y, WANG X, XU Q. MiR-99a enhances the radiation sensitivity of non-small cell lung cancer by targeting mTOR. *Cell Physiol Biochem* 2018; 46: 471-481.
- 6) DIRICAN E, AKKIPRIK M. Phosphatidylinositol 3-kinase regulatory subunit 1 and phosphatase and tensin homolog as therapeutic targets in breast cancer. *Tumour Biol* 2017; 39: 1393394135.
- 7) PERREAULT S, CHANDRASEKHAR J, CUI ZH, EVARTS J, HAO J, KAPLAN JA, KASHISHIAN A, KEEGAN KS, KENNEY T, KODITEK D, LAD L, LEPIST EI, McGRATH ME, PATEL L, PHILLIPS B, THERRIEN J, TREIBERG J, YAHIAOUI A, PHILLIPS G. Discovery of a phosphoinositide 3-kinase (PI3K) beta/delta inhibitor for the treatment of phosphatase and tensin homolog (PTEN) deficient tumors: building PI3Kbeta potency in a PI3Kdelta-selective template by targeting non-conserved Asp856. *J Med Chem* 2017; 60: 1555-1567.
- 8) GOLOB-SCHWARZL N, KRASSNIG S, TOEGLHOFER AM, PARK YN, GOGG-KAMERER M, VIERLINGER K, SCHRODER F, RHEE H, SCHICHO R, FICKERT P, HAYBAECK J. New liver cancer



- biomarkers: PI3K/AKT/mTOR pathway members and eukaryotic translation initiation factors. *Eur J Cancer* 2017; 83: 56-70.
- 9) HADDADI N, LIN Y, TRAVIS G, SIMPSON AM, NASSIF NT, MCGOWAN EM. PTEN/PTENP1: 'regulating the regulator of RTK-dependent PI3K/Akt signalling', new targets for cancer therapy. *Mol Cancer* 2018; 17: 37.
  - 10) NI D, JIANG D, VALDOVINOS HF, EHLERDING EB, YU B, BARNHART TE, HUANG P, CAI W. Bioresponsive polyoxometalate cluster for redox-activated photoacoustic imaging-guided photothermal cancer therapy. *Nano Lett* 2017; 17: 3282-3289.
  - 11) COSTELLO S, MACBETH F. Management of lung cancer. *BMJ* 1991; 302: 293.
  - 12) SORIA JC, OHE Y, VANSTEENKISTE J, REUNGWETWATTANA T, CHEWASKULYONG B, LEE KH, DECHAPHUNKUL A, IMAMURA F, NOGAMI N, KURATA T, OKAMOTO I, ZHOU C, CHO BC, CHENG Y, CHO EK, VOON PJ, PLANCHARD D, SU WC, GRAY JE, LEE SM, HODGE R, MAROTTI M, RUKAZENKOV Y, RAMALINGAM SS. Osimertinib in untreated EGFR-mutated advanced non-small-cell lung cancer. *N Engl J Med* 2018; 378: 113-125.
  - 13) PARK J, HWANG M, CHOI B, JEONG H, JUNG JH, KIM HK, HONG S, PARK JH, CHOI Y. Exosome classification by pattern analysis of surface-enhanced Raman spectroscopy data for lung cancer diagnosis. *Anal Chem* 2017; 89: 6695-6701.
  - 14) LIU G, PEI F, YANG F, LI L, AMIN AD, LIU S, BUCHAN JR, CHO WC. Role of autophagy and apoptosis in non-small-cell lung cancer. *Int J Mol Sci* 2017; 18: 367.
  - 15) ASHKENAZI A, FAIRBROTHER WJ, LEVERSON JD, SOUZA J. From basic apoptosis discoveries to advanced selective BCL-2 family inhibitors. *Nat Rev Drug Discov* 2017; 16: 273-284.
  - 16) ASKEW K, LI K, OLMOS-ALCALA A, DE LA MORENA F, LIANG Y, RICHARDSON P, BERTON T, CHAMMAN MA, RIECKEN K, BECCARI S, SIEMER M, MOLNAR P, CRAGG MS, GARASCHUK O, PERRY MH, GONZALEZ-LEON J. Proliferation and apoptosis maintain the rapid turnover of microglia in the adult brain. *Cell Rep* 2017; 18: 391-405.
  - 17) ZAMZAMI N, EL HC, MAISE C, BRENNER C, MUNOZ-PINEDO C, BELZACO AS, COSTANTINI P, VIEIRA H, LOEFFLER M, MOLLE G, KROEMER G. Bid acts on the permeability transition pore complex to induce apoptosis. *Oncogene* 2000; 19: 6342-6350.
  - 18) OPFERMAN JT, KOTHARI A. Anti-apoptotic BCL-2 family members in development. *Cell Death Differ* 2018; 25: 37-45.
  - 19) SLATTERY ML, MULLANY LE, HODDA LC, WATSON RK, STEVENS JR, SAMOWITZ WS, MERRICK JS. PTEN/PI3K/AKT signaling pathway associations of microRNAs with dysregulated gene expression in colorectal cancer. *Mol Carcinog* 2012; 23: 243-261.
  - 20) ZHANG X, CHEN Y, ZHAO Y, LI L, ZHANG W, WANG X. MicroRNA-143 functions as an oncogene by regulating PI3K/AKT/pAKT pathway in myeloma. *Leuk Lymphoma* 2017; 58: 932-937.
  - 21) ZHAO Y, DUPONT S, YAKAR S, KARAS M, LEROITH D. PTEN inhibits cell proliferation and induces cell apoptosis by downregulating cell surface IGF-1R expression in prostate cancer cells. *Oncogene* 2004; 23: 786-794.
  - 22) MA J, SAWAI H, MATSUYO Y, OCHI N, YASUDA A, TAKAHASHI H, WAKASUGI H, MUNAHASHI H, SATO M, TAKEYAMA H. Akt1/MEK1/ERK1/2 signaling pathway mediates PTEN suppression and enhances cell invasion and proliferation via activation of the IGF-1R/Akt signaling pathway in pancreatic cancer cells. *J Surg Res* 2010; 160: 90-101.
  - 23) TANG JM, HE QY, GUO RX, CHANG XJ. Phosphorylation of Akt overexpression and loss of PTEN expression in non-small cell lung cancer confers poor prognosis. *Lung Cancer* 2006; 51: 181-191.
  - 24) SORIA JC, LEE HY, LEE JI, WANG L, ISSA JP, KEMP BL, LIU DD, KURIE JM, MAO L, KHURI FR. Lack of PTEN expression in non-small cell lung cancer could be related to promoter methylation. *Clin Cancer Res* 2002; 8: 1178-1184.