

Combined treatment with myo-inositol, alpha-lipoic acid, folic acid and vitamins significantly improves sperm parameters of sub-fertile men: a multi-centric study

P. CANEPA¹, A. DAL LAGO², C. DE LEO³, M. GALLO², C. RIZZO¹,
E. LICATA², P. ANSERINI¹, R. RAGO², P. SCARUFFI¹

¹UOS Physiopathology of Human Reproduction, IRCCS Ospedale Policlinico San Martino, Genoa, Italy

²UOSD Physiopathology of Reproduction and Andrology, Ospedale Sandro Pertini, Rome, Italy

³Laboratory of Andrology, University of Genova, Genova, Italy

Abstract. – **OBJECTIVE:** Reduction in motility and number of spermatozoa and change in their morphology are some of the most relevant causes of male infertility. Production of reactive oxygen species may affect motility, morphology and DNA stability of spermatozoa. This study aimed at evaluating the effect of combined treatment with myo-inositol, alpha-lipoic acid, folic acid, betaine and vitamins (namely, Sinopol®) on semen parameters of sub-fertile men.

PATIENTS AND METHODS: We recruited 143 sub-fertile men, 26-53 years aged, no-smokers, without any testicular pathologies, with a normal endocrinological/metabolic profile, and no concomitant consumption of drugs. Out of them, 25 patients did not meet study inclusion criteria mainly due to the history of genital diseases that came to light after Sinopol® prescription. Among the 118 men that fulfilled inclusion criteria, 10 (8.4%) patients were lost at follow-up and in 8 (6.8%) cases the partner got pregnant spontaneously. Thus, 100 patients completed the study and semen analysis was performed before and after 90 days of treatment.

RESULTS: Semen quality improved after 90 days of treatments, with a statistically significant increase of sperm concentration ($p=0.0009$), of number of spermatozoa ($p=0.0017$), of progressive motility ($p=0.0047$), of total motile sperm count ($p=0.0010$), and of normal sperm morphology ($p<0.0001$).

CONCLUSIONS: For the first time we reported that a combination of nutraceuticals composed of myo-inositol, alpha-lipoic acid, folic acid, betaine and vitamins improves sperm parameters in sub-fertile men. We are aware that to clarify the clinical relevance of the data studies with larger sample sizes and longer durations are needed, as well as evaluation of myo-inositol and alpha-lipoic acid co-treatment effectiveness

in improving the chances to obtain a pregnancy spontaneously or following assisted reproduction.

Key Words:

Myo-inositol, Alpha-lipoic acid, Asthenozoospermia, Semen quality, Pregnancy.

Introduction

Infertility is a disease of reproductive system defined by failure to achieve the clinical pregnancy after 12 months or more of regular unprotected sexual intercourse¹. It affects about 10% of couples of childbearing age. Male partner is the sole cause, or a contributing cause, of infertility in about 40% of infertile couples². It has been estimated that almost 7% of men in their reproductive age are subfertile or infertile, due to testicular, pre-testicular or post-testicular problems. Conditions such as varicocele, cryptorchidism, and hypogonadism are among the many causes of male infertility³. Nevertheless, idiopathic infertility is found in about 25-30% of infertile patients⁴. Males with sperm parameters below the World Health Organization (WHO) reference values are considered to have male factor infertility⁵. The most significant of parameters are low sperm concentration (oligospermia), poor sperm motility (asthenospermia), and abnormal sperm morphology (teratospermia). Other factors less well associated with infertility include semen volume and seminal markers of epididymal, prostatic, and seminal vesicle function⁶. Some 30% to 80% of male factor subfertility cases are believed to be due to the damaging effects of

oxidative stress that occurs when reactive oxygen species (ROS) overcome the semen's natural antioxidant defenses and cause cellular damage to the sperm⁷. ROS include a broad spectrum of molecules such as oxygen free radicals (superoxide anion, hydroxyl radical, hydroperoxyl radical), non-radical species (hypochlorous acid and hydrogen peroxide), reactive nitrogen species and free nitrogen radicals (nitroxyl ion, nitrous oxide, peroxynitrite). Physiologically, in the seminal plasma homeostasis between free radicals and antioxidant substances is guaranteed by a very complex system involving superoxide dismutase, catalase and glutathione peroxidase dependent activity, associated with significant concentrations of antioxidants such as ascorbic acid (vitamin C), tocopherol (vitamin E) and glutathione. Several conditions may alter this antioxidant system and potentially lead to a harmful accumulation of ROS in seminal fluid. These include pathological conditions involving the reproductive tract (varicocele, prostatitis), infections, autoimmunity and chronic disease, environmental factors (high temperatures, electromagnetic radiation, pesticides and air pollution), some lifestyles (smoking, alcohol abuse, drug addiction), and nutritional errors (unbalanced hyperlipidic diet, obesity, poor diet)⁸. Recently, metabolic syndrome and its individual symptoms as obesity, dyslipidemia, hypertension, and impaired glucose metabolism with insulin resistance, have been associated with detrimental effects on sperm fertility potential and DNA integrity⁹. In fact, research over the past decade has demonstrated that the pro-inflammatory state seen in metabolic dysfunction and the subsequent development of oxidative stress may have direct detrimental effects on normal spermatogenesis and sperm function¹⁰. Spermatozoa are particularly susceptible to oxidative damage because their plasma membranes are rich in polyunsaturated fatty acids and have low concentrations of scavenging enzymes¹¹. Under physiologic states, spermatozoa themselves produce little amounts of ROS that are needed for capacitation and acrosomal reaction¹². Excessive generation of ROS in semen by leukocytes as well as by abnormal spermatozoa may affect motility, morphology, mitochondrial membranes, chromatin condensation and DNA stability, thus compromising sperm function¹³. Oral supplementation with antioxidants may improve sperm quality by reducing oxidative stress¹⁴, thus dietary supplementation with antioxidants has gained much attention in recent years^{13,15-17}. One potent antioxidant is alpha-lipoic acid (ALA), a natural short-chain fatty acid containing

sulfhydryl groups¹⁸. It is active both in the aqueous phase (cytoplasm) and in the lipid phase (membrane) of cells. ALA is the coenzyme for pyruvate dehydrogenase and α -ketoglutarate dehydrogenase mitochondrial respiratory enzymes and thus it improves mitochondrial function^{19,20}. Inside cells and tissues, ALA is reduced to dihydrolipoic acid (DHLA), which has more antioxidant properties than does lipoic acid. Both DHLA and ALA have metal-chelating capacity and scavenge ROS, and DHLA is able to regenerate endogenous antioxidants vitamin E, vitamin C and glutathione, and to repair oxidative damage. Alpha-lipoic acid is also a main factor in the Krebs cycle and contributes to ATP biosynthesis, which is crucial for the sperm viability²¹. Results of animal studies showed that the percentage of motile sperm increased dramatically after ALA administration²¹⁻²³. Until now, only one study has been conducted in humans²⁴ and, although further investigation is suggested in this regard, medical therapy of asthenoteratospermia with ALA supplement could improve quality of semen parameters. Moreover, ALA was shown to be effective in reducing symptoms of diabetic polyneuropathy²⁵ and body weight in obese subjects²⁶.

Few studies²⁷⁻³¹ have investigated the role of another possible antioxidant agent, namely myo-inositol (MYO), both for systemic treatment of male infertility and for improvement of sperm quality used in *in vitro* fertilization techniques. Inositol is a polyalcohol, naturally occurring as nine stereoisomers, including D-chiro-inositol and MYO, that belongs to the vitamin B complex group. In male reproductive organs, MYO is mainly produced by Sertoli cells in response to follicle-stimulating hormone (FSH) and is involved in processes that include the regulation of motility, capacitation and acrosome reaction of sperm cells^{28,32-34}. Moreover, since concentrations of MYO increase from the epididymis to the deferent duct, this compound is considered important for sperm maturation and for sperm chemotaxis through production of inositol triphosphate and calcium channels opening³⁵. Low concentration of MYO within epididymis has been associated with reduced fertility in transgenic mice³⁶. MYO also seems to play a role in osmoregulation in seminal vesicular fluid, contributing to reduce viscosity and increase sperm motility³¹. In relation to this, when added to seminal fluid of patients with oligoasthenoteratospermia, MYO reduces the presence of amorphous material²⁷. At a functional level, MYO acts directly on mitochondria increasing the membrane potential and thereby improving sperm motility in patients with altered sperm parameters³⁷.

In fact, *in vitro* MYO increased the number of spermatozoa with high mitochondrial membrane potential (MMP) and it decreased the number of those with low MMP in OAT patients³⁷.

Folate is important for the synthesis of DNA, transfer RNA and proteins, and is therefore suggested to be important in spermatogenesis³⁸. Although results are controversial³⁹ and the absence of an effect of folic acid on endocrine parameters⁴⁰ (i.e., FSH, testosterone and inhibin B concentrations), some researchers^{40,41} demonstrated that folic acid intervention improved sperm concentration and motility in sub-fertile males. Betaine is a natural methylamine that acts as a methyl donor to convert the harmful homocysteine into methionine. Several studies suggested that betaine may have an important role in maintaining male fertility^{42,43} and antioxidant proprieties^{44,45}.

As regarding B vitamins, positive effects of vitamin B12 on sperm count, sperm motility, and reducing sperm DNA damage, may be related to increased energy metabolism or to higher oxygenation due to the boost in red blood cell production⁴⁶. In the present study we aimed at evaluating for the first time the effect of combined treatment with MYO, ALA, folic acid and vitamins on semen parameters of sub-fertile men.

Patients and Methods

Patients

A total of 143 no-smokers sub-fertile men were recruited at UOS Physiopathology of Human Reproduction, Ospedale Policlinico San Martino, Genoa and at Physiopathology of Reproduction and Andrology Unit, Sandro Pertini Hospital, Rome from April 2016 to September 2017. The 143 subjects were analyzed for eligibility: 25 did not meet inclusion criteria and, therefore, were excluded from the study. The remaining 118 subjects were enrolled. Eighteen subjects did not complete the study: 10 were lost at follow-up and 8 for spontaneous pregnancy. Therefore, a total of 100 subjects were included in statistical analysis. The patients were 26-53 years aged (average age: 39.6 ± 5.9 years), with one or more altered semen parameters according to WHO 2010 criteria⁵. Specifically, at the baseline 16% of patients presented oligospermia, 43% asthenospermia, 41% both oligospermia and asthenospermia. Moreover, patients had a body mass index (BMI) between 20 kg/m² and 25 kg/m², a normal endocrinological/metabolic profile (homeostatic model assessment

(HOMA) index³⁸ < 2.5; 150 ng/dL < testosterone < 800 ng/dL, 0 ng/ml < prolactin < 20 ng/ml, 1.5 mIU/mL < FSH < 12.4.0 mIU/mL, 6 mIU/mL < LH < 30 mIU/mL serum values), and no concomitant consumption of drugs. Patients with azoospermia or severe oligozoospermia (sperm count less than 5 x 10⁶/ml) or with an identifiable cause of infertility (leukocytospermia and/or positive sperm culture, epididymo-orchitis, prostatitis, inguinoscrotal surgery, cryptorchidism, varicocele, etc.) were excluded from the study.

Study Design

Semen analysis was performed at baseline (T0) and after 90 days (T90) of treatment, and results were compared. The study was approved by the Ethical Committee of Regione Liguria (Approval no. 416REG2017) and each patient signed an informed consent to the enrollment in the study.

Treatment Protocol

Diet of patients was supplemented with 2 tablets/day of Sinopol[®] (Laborest S.r.l., Nerviano, Milan, Italy). Two Sinopol[®] tablets contained ALA (800 mg), MYO (1000 mg), folic acid (400 mg), betaine (100 mg), vitamin B2 (1.7 mg), B6 (1.9 mg), and B12 (2.6 mg).

Sperm Analysis

At T0 and T90 semen samples were obtained after 3 days of sexual abstinence. They were collected into sterile containers and held at 37°C for 30 minutes to liquefy. After liquefaction, a routine sperm analysis was carried out according to the WHO criteria⁵, evaluating sperm number, motility, and morphology. Sperm score for motility evaluation was expressed as grades a to d, and progressive motility rate was calculated as the percentage of a + b.

Statistical Analysis

Data were expressed as mean ± standard deviation (SD). Mann-Whitney test was performed using MedCalc[®] software (Mariakerke, Belgium), in order to investigate differences between semen parameters before and after the treatment. Results were considered statistically significant at $p < 0.05$.

Results

One hundred and forty-three patients were recruited and began Sinopol[®] treatment. Out of them, 25 patients did not meet inclusion criteria in the stu-

Table I. Semen characteristics of patients before and after treatment.

	Before*	After*	p-value
Sperm concentration (x 106/ml)	16.6 + 13.1	24.4 + 23.4	0.0009
Number of spermatozoa (x 106/ml)	46.5 + 38.7	69.1 + 59.0	0.0017
Progressive motility (%)	19.5 + 15.6	24.8 + 16.5	0.0047
Total motile sperm count (x 106)	9.8 + 11.5	22.3 + 30.8	0.0010
Normal sperm morphology (%)	4.9 + 3.1	7.9 + 4.1	< 0.0001

*Data are expressed as mean + SD

dy mainly due to history of genital diseases as epididymo-orchitis, prostatitis, cryptorchidism, current genital inflammation or varicocele that came to light after Sinopol[®] prescription. Among the 118 men that fulfilled inclusion criteria, 10 (8.4%) patients were lost at follow-up and in 8 (6.8%) cases the partner got pregnant spontaneously. A total of 100 patients completed the whole study (Figure 1). The patients were 28-52 years aged (average age: 37.7 + 4.7 years). No side effects due to the oral administration of Sinopol[®] were observed in any participants. Sperm parameters of the 100 participants

at baseline and at the end of the study are presented in Table I. Analyses of data showed a statistically significant increase of sperm concentration (percentage increase=+41.2%, $p=0.0009$), of number of spermatozoa (percentage increase=+50%, $p=0.0017$), of progressive motility (percentage increase=+31.6%, $p=0.0047$), of total motile sperm count (percentage increase=+120%, $p=0.0010$), and of normal sperm morphology (percentage increase=+60%, $p<0.0001$) after the treatment respect on the baseline. We also registered the reproductive follow-up of the 100 patients that completed the study. Within

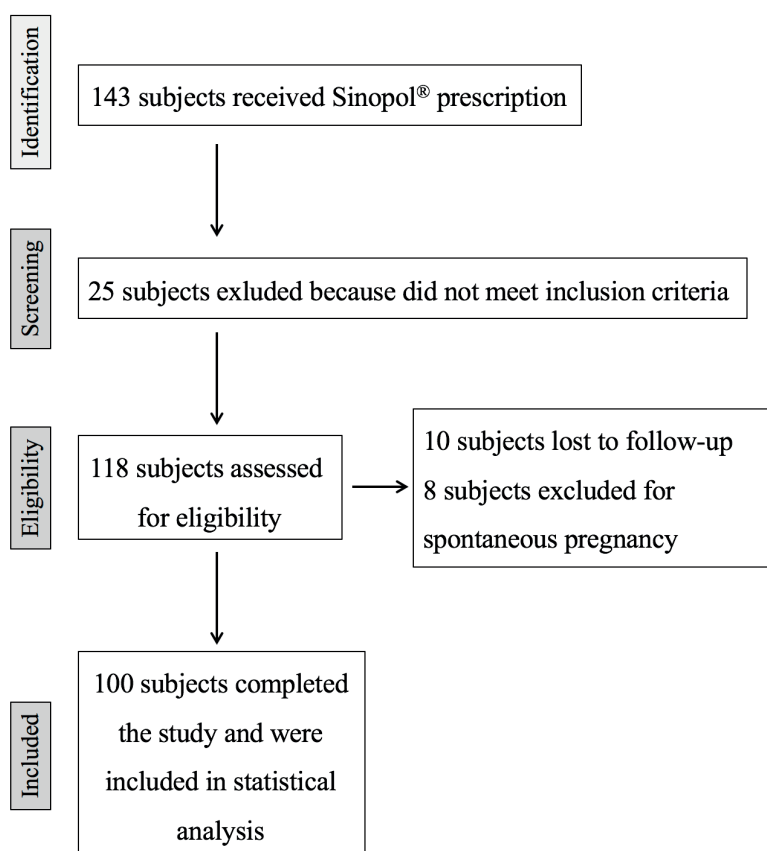


Figure 1. Flowchart of patient recruitment for the study of Sinopol[®] supplementation in sub-fertile men.

6 months following Sinopol[®] discontinuation, the partner got pregnant in 40 cases (6 spontaneously, 4 with intra-uterine insemination, and 30 after ICSI or FIVET treatment).

Discussion

This multi-centric study represents the first evaluation of effects of combined treatment with myo-inositol, alpha-lipoic acid, folic acid, betaine and vitamins of sub-fertile men. After 3 months of co-administration of these dietary supplements, a significant improvement of sperm parameters was observed. Regulation of sperm mobility is largely controlled at the mid-piece: the sperm flagellum handles the activation of motility, and the principal mid-piece controls hyperactivation⁴⁸. The mid-piece is rich in unsaturated lipids and saturated protein channels that are vulnerable to free radical attack. Therefore, the anti-oxidant properties of ALA and MYO may protect these components and, in turn, ensure their structural and functional integrity⁴⁹. The rate of sperm motility is also highly dependent on its energy supply and thus on external and internal structural integrity of mitochondria⁵⁰. At the same time, very active mitochondria of normal function sperms create high levels of free radicals as by-products of the Krebs cycle. Therefore, antioxidant addition would protect these organelles from free radicals attack and ensure a constant yield of ATP²¹. The ALA is thought to assist the metabolism of oxidative decarboxylation by acting as a co-enzyme, thus increasing cytochrome C concentration and directly raising the mitochondria's membrane potential, likewise MYO does^{27,35,51}. Moreover, MYO has been suggested to play a role in sperm chemotaxis through the activation of phospholipase C, resulting in the production of inositol triphosphate and calcium channels opening⁵², and through reduction of the semen amorphous material. Therefore, the synergic effect of ALA and MYO would result in a positive effect on sperm mobility, as observed.

We also found that administration of ALA and MYO was able to increase sperm number. This finding may be attributed to the normalized oxidative stress which on one hand may modulate proliferation of germ cell, protected against DNA fragmentation and membrane damages, and on the other may maintain signal transduction mechanisms necessary for normal function of hypothalamus-testicular axis. On the whole, the defensive

influence of ALA and MYO against oxidative reproductive damage may lead to normal secretion of testosterone and sperm production. With regards to this issue, it has been demonstrated that ALA sustained sex-hormone-binding globulin and dehydroepiandrosterone sulfate serum concentration, the levels of testicular cholesterol, and glucose-6-phosphatedehydrogenase and 3 β -hydroxysteroid dehydrogenase activities in testis²³. Moreover, with oral administration, MYO has been indicated to improve the hormonal milieu of the testis. Specifically, MYO is able to rebalance serum luteinizing hormone, follicle-stimulating hormone (FSH), and inhibin B concentrations²⁸. Since FSH plays a key role in control of Sertoli cell number and function, that are vital for normal spermatogenesis, it is possible that MYO may enhance the function of these nurse cells. MYO seems to play an important role in the maturation of sperms also during the migration from the epididymis. In fact, MYO shows a gradient of concentration, most abundant in the seminal plasma increasing along epididymis and deferent ducts⁵³. In addition, folic acid and vitamin B12 contained in Sinopol[®] may contribute to the improvement of semen characteristics found at the treatment discontinuation. Folate may be beneficial for spermatogenesis since it is required for the *de novo* synthesis of purines, thymidylate and methionine³⁸. Although the underlying mechanism involved is not clarified and results are controversial³⁹, some authors demonstrated that folic acid improved number and motility of sperm^{40,41}. A recent review demonstrated the positive impact of vitamin B12 on semen quality by increasing sperm count and by enhancing sperm motility and reducing sperm DNA damage⁵⁴. The beneficial effects of vitamin B12 on semen quality may be due to increased functionality of reproductive organs, decreased homocysteine toxicity, reduced amounts of generated nitric oxide, decreased levels of oxidative damage to sperm, reduced amount of energy produced by spermatozoa, decreased inflammation-induced semen impairment, and control of nuclear factor-kB activation⁵⁴. The strength of this study is that it showed for the first time that the co-treatment with MYO and ALA, combined with folic acid and vitamins, can improve semen quality of dispermic men. In fact, up to now it has been demonstrated the positive impact on infertile men of the individual administration of MYO or ALA^{24,28,30,55}, and at present Sinopol[®] is labeled as a dietary supplement to counteract nutritional deficiencies in women affected by PCOS (www.

laborest.com/en/products/sinopol/sinopol-fast-slow-tablets). Moreover, we not only reported no adverse effects of this combined treatment on the male subjects, but we also demonstrated that a lower dosage of MYO and ALA than those previously found to be tolerated by human subjects⁵⁶ or used in infertile men^{24,28,30,56} is properly effective, probably due to a synergic effect between ALA and MYO.

Conclusions

For the first time we reported that a combination of nutraceuticals composed of myo-inositol, alpha-lipoic acid, folic acid, betaine and vitamins improves sperm parameters in sub-fertile men. We are aware that to clarify the clinical relevance of the data more investigations with larger sample sizes and longer durations are needed, as well as evaluation of MYO and ALA co-treatment effectiveness in improving the chances to obtain a pregnancy spontaneously or following assisted reproduction.

Conflict of Interest

The Authors declare that they have no conflict of interest.

Funding

This study was funded in part by Laborest Company. Laborest Company had no role in study design, data collection, data analysis, data interpretation, and writing of the paper.

References

- ZEGERS-HOCHSCHILD F, ADAMSON GD, DE MOUZON J, ISHIHARA O, MANSOUR R, NYGREN K, SULLIVAN E, VANDERPOEL S; International Committee for Monitoring Assisted Reproductive Technology; World Health Organization. International Committee for Monitoring Assisted Reproductive Technology (ICMART) and the World Health Organization (WHO) revised glossary of ART terminology, 2009. *Fertil Steril* 2009; 92: 1520-1524.
- KUMAR N, SINGH AK. Trends of male factor infertility, an important cause of infertility: a review of literature. *J Hum Reprod Sci* 2015; 8: 191-196.
- LOTTI F, MAGGI M. Ultrasound of the male genital tract in relation to male reproductive health. *Hum Reprod Update* 2015; 21: 56-83.
- QUAAS A, DOKRAS A. Diagnosis and treatment of unexplained infertility. *Rev Obstet Gynecol* 2008; 1: 69-76.
- WORLD HEALTH ORGANIZATION, DEPARTMENT OF REPRODUCTIVE HEALTH AND RESEARCH. WHO laboratory manual for the examination and processing of human semen. Fifth edition. WHO Press, Geneva, Switzerland, 2010.
- COOPER TG, NOONAN E, VON ECKARDSTEIN S, AUGER J, BAKER HW, BEHRE HM, HAUGEN TB, KRUGER T, WANG C, MBIZVO MT, VOGELSONG KM. World Health Organization reference values for human semen characteristics. *Hum Reprod Update* 2010; 16: 231-245.
- TREMELLEN K. Oxidative stress and male infertility—a clinical perspective. *Human Reproduction Update* 2008; 14: 243-258.
- POURMASUMI S, SABETI P, RAHIMINIA T, MANGOLI E, TABIBNEJAD N, TALEBI AR. The etiologies of DNA abnormalities in male infertility: an assessment and review. *Int J Reprod Biomed* 2017; 15: 331-344.
- MORRISON CD, BRANNIGAN RE. Metabolic syndrome and infertility in men. *Best Pract Res Clin Obstet Gynaecol* 2015; 29: 507-515.
- OMU AE. Sperm parameters: paradigmatic index of good health and longevity. *Med Princ Pract* 2013; 22: 30-42.
- OPUWARI CS, HENKEL RR. An update on oxidative damage to spermatozoa and oocytes. *Biomed Res Int* 2016; 2016: 9540142.
- DACHEUX JL, DACHEUX F. New insights into epididymal function in relation to sperm maturation. *Reproduction* 2014; 147: R27-42.
- GHARAGOZLOO P, AITKEN RJ. The role of sperm oxidative stress in male infertility and the significance of oral antioxidant therapy. *Hum Reprod* 2011; 26: 1628-1640.
- ZINI A, SAN GABRIEL M, BAAZEEM A. Antioxidants and sperm DNA damage: a clinical perspective. *J Assist Reprod Genet* 2009; 26: 427-432.
- LANZAFAME FM, LA VIGNERA S, VICARI E, CALOGERO AE. Oxidative stress and medical antioxidant treatment in male infertility. *Reprod Biomed Online* 2009; 19: 638-659.
- ROSS C, MORRIS A, KHAIRY M, KHALAF Y, BRAUDE P, COMARASAMY A, EL-TOUKHY T. A systematic review of the effect of oral antioxidants on male infertility. *Reprod Biomed Online* 2010; 20: 711-723.
- SHOWELL MG, MACKENZIE-PROCTOR R, BROWN J, YAZDANI A, STANKIEWICZ MT, HART RJ. Antioxidants for male subfertility. *Cochrane Database Syst Rev* 2014; 12: CD007411.
- BIEWENGA GP, HAENEN GR, BAST A. The pharmacology of the antioxidant lipoic acid. *Gen Pharmacol* 1997; 29: 315-331.
- PACKER L, WITT EH, TRITSCHLER HJ. alpha-lipoic acid as a biological antioxidant. *Free Radic Biol Med* 1995; 19: 227-250.
- REED LJ. From lipoic acid to multi-enzyme complexes. *Protein Sci* 1998; 7: 220-224.
- IBRAHIM SF, OSMAN K, DAS S, OTHMAN AM, MAJID NA, RAHMAN MP. A study of the antioxidant effect of alpha lipoic acids on sperm quality. *Clinics* 2008; 63: 545-550.

- 22) YENI D, FIDAN AF, CIĞERCİ IH, KONUK M, AVDATEK F, GÜNDOĞAN M. Effect of α -lipoic acid on sperm quality, reproductive tract measures in thinner exposed rats. *Andrologia* 2012; 44: 74-80.
- 23) OTHMAN AI, EL-MISSIRY MA, KORIEM KM, EL-SAYED AA. Alfa-lipoic acid protects testosterone secretion pathway and sperm quality against 4-tert-octylphenol induced reproductive toxicity. *Ecotoxicol Environ Saf* 2012; 81: 76-83.
- 24) HAGHIGHIAN HK, HAIDARI F, MOHAMMADI-ASL J, DADFAR M. Randomized, triple-blind, placebo-controlled clinical trial examining the effects of alpha-lipoic acid supplement on the spermatogram and seminal oxidative stress in infertile men. *Fertil Steril* 2015; 104: 318-324.
- 25) ZIEGLER D, AMETOV A, BARINOV A, DYCK PJ, GURIEVA I, LOW PA, MUNZEL U, YAKHNO N, RAZ I, NOVOSADOVA M, MAUS J, SAMIGULLIN R. Oral treatment with alphalipoic acid improves symptomatic diabetic polyneuropathy: the SYDNEY 2 trial. *Diabetes Care* 2006; 29: 2365-2370.
- 26) KOH EH, LEE WJ, LEE SA, KIM EH, CHO EH, JEONG E, KIM DW, KIM MS, PARK JY, PARK KG, LEE HJ, LEE IK, LIM S, JANG HC, LEE KH, LEE KU. Effects of alpha-lipoic acid on body weight in obese subjects. *Am J Med* 2011; 124: 85.e1-8.
- 27) COLONE M, MARELLI G, UNFER V, BOZZUTO G, MOLINARI A, STRINGARO A. Inositol activity in oligoasthenoteratospermia-an in vitro study. *Eur Rev Med Pharmacol Sci* 2010; 14: 891-896.
- 28) CALOGERO AE, GULLO G, LA VIGNERA S, CONDORELLI RA, VAIARELLI A. Myo-inositol improves sperm parameters and serum reproductive hormones in patients with idiopathic infertility: a prospective double-blind randomized placebo-controlled study. *Andrology* 2015; 3: 491-495.
- 29) BEVILACQUA A, CARLOMAGNO G, GERLI S, MONTANINO OLIVA M, DEVROEY P, LANZONE A, SOULANGE C, FACCHINETTI F, CARLO DI RENZO G, BIZZARRI M, HOD M, CAVALLI P, D'ANNA R, BENVENGA S, CHIU TT, KAMENOV ZA. Results from the International Consensus Conference on myo-inositol and D-chiro-inositol in obstetrics and gynecology-assisted reproduction technology. *Gynecol Endocrinol* 2015; 31: 441-446.
- 30) GULINO FA, LEONARDI E, MARILLI I, MUSMECI G, VITALE SG, LEANZA V, PALUMBO MA. Effect of treatment with myo-inositol on semen parameters of patients undergoing an IVF cycle: in vivo study. *Gynecol Endocrinol* 2016; 32: 65-68.
- 31) ARTINI PG, CASAROSA E, CARLETTI E, MONTELEONE P, DI NOIA A, DI BERARDINO OM. *In vitro* effect of myo-inositol on sperm motility in normal and oligoasthenospermia patients undergoing in vitro fertilization. *Gynecol Endocrinol* 2017; 33: 109-112.
- 32) CONDORELLI RA, LA VIGNERA S, BELLANCA S, VICARI E, CALOGERO AE. Myo-inositol: does it improve sperm mitochondrial function and sperm motility? *Urology* 2012; 79: 1290-1295.
- 33) MONTANINO OLIVA M, MINUTOLO E, LIPPA A. Effect of myo-inositol and antioxidants on sperm quality in men with metabolic syndrome. *Int J Endocrinol* 2016; 2016: 1674950.
- 34) CONDORELLI RA, LA VIGNERA S, MONGIOI LM, VITALE SG, LAGANÀ AS, CIMINO L, CALOGERO AE. Myo-inositol as a male fertility molecule: speed them up! *Eur Rev Med Pharmacol Sci* 2017; 21: 30-35.
- 35) HINTON BT, WHITE RW, SETCHELL BP. Concentrations of myo-inositol in the luminal fluid of the mammalian testis and epididymis. *J Reprod Fertil* 1980; 58: 395-399.
- 36) YEUNG CH, ANAPOLSKI M, SETIAWAN I, LANG F, COOPER TG. Effects of putative epididymal osmolytes on sperm volume regulation of fertile and infertile c-ros transgenic Mice. *J Androl* 2004; 25: 216-223.
- 37) CONDORELLI RA, LA VIGNERA S, DI BARI F, UNFER V, CALOGERO AE. Effects of myo-inositol on sperm mitochondrial function in-vitro. *Eur Rev Med Pharmacol Sci* 2011; 15: 129-134.
- 38) FOX JT, STOVER PJ. Folate-mediated one-carbon metabolism. *Vitam Horm* 2008; 79: 1-44.
- 39) RAIGANI M, YAGHMAEI B, AMIRJANNI N, LAKPOUR N, AKHONDI MM, ZERAATI H, HAJIHOSEINAL M, SADEGHI MR. The micronutrient supplements, zinc sulphate and folic acid, did not ameliorate sperm functional parameters in oligoasthenoteratozoospermic men. *Andrologia* 2014; 46: 956-962.
- 40) EBISCH IM, PIERIK FH, DE JONG FH. Does folic acid and zinc sulphate intervention affect endocrine parameters and sperm characteristics in men? *Int J Androl* 2006; 29: 339-345.
- 41) WONG WY, MERKUS HM, THOMAS CM, MENKVELD R, ZIELHUIS GA, STEEGERS-THEUNISSEN RP. Effects of folic acid and zinc sulfate on male factor subfertility: a double-blind, randomized, placebo-controlled trial. *Fertil Steril* 2002; 77: 491-498.
- 42) KELLY TL, NEAGA OR, SCHWAHN BC, ROZEN R, TRASLER JM. Infertility in 5, 10-methylenetetrahydrofolate reductase (MTHFR)-deficient male mice is partially alleviated by lifetime dietary betaine supplementation. *Biol Reprod* 2005; 72: 667-677.
- 43) SLOW S, LEVER M, CHAMBERS ST, GEORGE PM. Plasma dependent and independent accumulation of betaine in male and female rat tissues. *Physiol Res* 2009; 58: 403e10.
- 44) SHADMEHR S, FATEMI TABATABAEI SR, HOSSEINIFAR S, TABANDEH MR, AMIRI A. Attenuation of heat stress-induced spermatogenesis complications by betaine in mice. *Theriogenology* 2018; 106: 117-126.
- 45) ALIREZAEI M, JELODAR G, GHAYEMI Z. Antioxidant defense of betaine against oxidative stress induced by ethanol in the rat testes. *Int J Pept Res Ther* 2012; 18: 239e47.
- 46) MATTHEWS DR, HOSKER JP, RUDENSKI AS, NAYLOR BA, TREACHER DF, TURNER RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; 28: 412-419.
- 47) SUAREZ SS, MARQUEZ B, HARRIS TP, SCHIMENTI JC. Different regulatory systems operate in the midpie-

- ce and principal piece of the mammalian sperm flagellum. *Soc Reprod Fert Suppl* 2007; 65: 331-334.
- 48) HAMANO Y. Continuous infusion of lipoic acid rapidly reduces plasma beta-hydroxybutyrate with elevation of non-esterified fatty acids in broiler chickens. *Br J Nutr* 2007; 97: 495-501.
- 49) RUIZ-PESINI E, DIEZ C, LAPEÑA AC, PÉREZ-MARTOS A, MONTOYA J, ALVAREZ E, ARENAS J, LÓPEZ-PÉREZ MJ. Correlation of sperm motility with mitochondrial enzymatic activities. *Clin Chem* 1998; 44: 1616-1620.
- 50) PALANIAPPAN AR, DAI A. Mitochondrial ageing and the beneficial role of alpha-lipoic Acid. *Neurochem Res* 2007; 32: 1552-1558.
- 51) BANSAL AK, BILASPURI GS. Impacts of oxidative stress and antioxidants on semen functions. *Vet Med Int* 2011; 2011: 686137.
- 52) HINTON BT, WHITE RW AND SETCHELL BP. Concentrations of myo-inositol in the luminal fluid of the mammalian testis and epididymis. *J Reprod Fert* 1980; 58: 396-399.
- 53) BANIHANI SA. Vitamin B12 and semen quality. *Bio-molecules* 2017; 7: E42.
- 54) DINKOVA A, MARTINOV D, KONOVA E. Efficacy of myo-inositol in the clinical management of patients with asthenozoospermia. *Eur Rev Med Pharmacol Sci* 2017; 21: 62-65.
- 55) WOLLIN SD, JONES PJ. Alpha-lipoic acid and cardiovascular disease. *J Nutr* 2003; 133: 3327-3330.