Current diagnosis and potential obstacles for post-neurosurgical bacterial meningitis

M.-Y. BAO¹, H.-T. XIE², P. GAO¹, X. MAO¹, Z.-Y. LI¹, W.-H. WANG¹, S. SOPHEAK¹, H.-W. CHENG¹, L. YE¹, X. ZHANG¹

¹Department of Neurosurgery, the First Affiliated Hospital of Anhui Medical University, Hefei, China ²First Clinical Medical College, Anhui Medical University, Hefei, China

Bao and Xie contributed to the study equally

Abstract. - Despite the guidance of aseptic technology applied, bacterial meningitis seems to be an unavoidable obstacle in the process of neurosurgery, with high rates of disability and mortality. The diagnosis of post-neurosurgical bacterial meningitis (PNBM) mainly depends both on clinical symptoms and laboratory outcomes. Due to the excessive neuro-inflammatory reactions which are evoked by the primary brain disease or the craniotomy operation, the symptoms derived from the infection and aseptic may not be easily distinguished. On the other hand, the low positive rate and time-consuming character restrict the clinical practical values of bacterial culture. Therefore, it is always difficult to make a definite diagnosis of post-neurosurgical bacterial meningitis. Here, we reviewed the established literature about the diagnostic biomarkers for the PNBM and analyzed the potential obstacles in both clinical and scientific studies. Given the obstacle which has negative impacts on further investigation about the biology of PNBM, we only find relatively small numbers of study on PNBM. In this review, we summarize the established diagnostic methods and biomarkers for PNBM. Meanwhile, we also propose some potential investigation prospects. This review may help to better understand the character of PNBM in both clinical diagnosis and scientific investigations.

Key Words:

Meningitis, Neurosurgery, Bacterial infection, Aseptic inflammation, Biomarker.

Introduction

Central nervous system (CNS) infection is a severe infectious disease, leading to about 20.4 million disability-adjusted life year globally. Among that, nosocomial bacterial meningitis, also named

post-neurosurgical bacterial meningitis (PNBM), refers to a category of severe complications after the craniotomy, causing inevitable obstacles in the procedures of neurosurgery.

The PNBM types mainly include the meningitis, brain abscess, subdural empyema, and epidural abscess, with a common hallmark event of the blood brain barrier (BBB) breakdown which is attributed to the pathogen derived toxin and the host-related excessive neuro-inflammatory reactions²⁻⁴. In comparison with the community-acquired bacterial meningitis, the excessive neuro-inflammatory reactions by the triplicity of primary CNS disease, neurosurgical process and bacterial infection in the PNBM conspire to make accurate diagnosis difficult. However, as the occurrence of PNBM increases the overall cost of hospital care, postpones the neurological recoveries after effective treatment for the primary CNS disease and, the most importantly, results in disabilities^{5,6}, it is still urgently necessary to seek for the optimal management of these often critically ill patients.

The incidence of PNBM varies a lot among studies, ranging from 0.3% to 10%^{7,8}, and is usually influenced by numerous factors, such as indication for surgery, underlying medical condition, longer operation time, indwelling drainage tube, implantation of artificial materials, cerebrospinal fluid leakage and underlying diseases of the patients⁹. The epidemiological study showed that <45 aged patients had higher risks for infection¹⁰. In addition, among all the brain tumors, infratentorial and intraventricular tumors have higher risks for infection¹⁰. Importantly, the application of artificial material significantly increases the incidence rate of PNBM. It has been reported¹¹ that the patients who used external drainage have a 9.4-fold higher risks than the patient who did not use it. Meanwhile, we also noticed that the mortality of PNBM was 20%-50%, if no proper treatments were timely applied¹². In some specific type of meningitis, such as carbapenem-resistant gram-negative postoperative meningitis (CR-GNPOM), the mortality can reach as high as 60%-70%¹³.

Clinically, the etiology of PNBM has a wide spectrum of microorganisms, which are from gram-positive cocci to gram-negative bacilli. Meanwhile, during the last decade, the infectious bacterial spectrum for the PNBM has altered from staphylococcus aureus, coagulase negative staphylococcus, and enterococcus gram-positive bacteria to gram-negative bacilli, especially enterobacteriaceae, probably due to the extensive usage of antibiotics nosocomially or in community^{14,15}. Another important epidemiological issue is the ever-increasing situation for drug-resistance bacteria. World Health Organization (WHO) reported in 2017 that the carbapenem-resistant enterobacteriaceae and extended spectrum beta-lactamase (ESBL) producing enterobacteriaceae were the most urgent antibiotic-resistant bacteria worldwide¹⁶. In China, the Chinese Anti-microbial Surveillance Network reported that the proportion of methicillin-resistant S. aureus is almost 100% and the resistance rates for both erythromycin and clindamycin are also 100%¹⁷. In clinical practice, the clinicians usually use vancomycin as an empiric choice for management of multi-drug resistance (MDR) S. aureus, but the low rates for BBB penetration and bioavailability in CNS restrict its application in treating PNBM¹⁷. As the empirical usage of antibiotics increases the risk for the bacteria spectrum alteration and causes more chances for the MDR bacteria, the timely diagnosis of infection and the accurate recognition of bacteria species are two issues which are urgently to be coped with.

In this review, we mainly focus on the current diagnostic methods and biomarkers of PNBM. Given the limited numbers of the research on PNBM, we meanwhile discuss the existing obstacles in both clinical diagnosis and laboratory investigations, and at the end we are seeking for several potential breakthroughs in the PNBM study. Ultimately, we want the review to provide some clinical or laboratory clues in helping the precision medicine for PNBM.

Diagnostic Methods and Biomarkers

Diagnostic Criteria

The clinical diagnosis of PNBM usually depends on both symptoms and laboratory outcomes, but it is still dubious sometimes regardless

of the applied methods. The clinical symptoms include neck stiffness, fever, headache and vomiting. However, since the patients receive a neurosurgery before symptoms' occurrence, the signs of both chemical meningitis and aseptic inflammation, which are attributed to the primary neurosurgical disease and the craniotomy itself, are performed similarly¹⁸. Thus, clinical symptoms could only provide preliminary clues, rather than a specific indication in the diagnosis.

On the other hand, the clinicians would like to use the laboratory tests to make a definite diagnosis, especially for the outcomes in the cerebrospinal fluid (CSF). Although the PNBM infects the tissues in CNS in a localized manner, systemic immunological reactions also have significant effects against the pathogenic bacteria¹⁹. In normal conditions, the micro-environment of the CSF keeps steady since the segregation function of BBB. However, BBB dysfunction usually can be found due to the primary neurosurgical disease or the secondary infection before the PNBM onset^{20,21}. Subsequently, the CSF biochemical alterations, including the proteins, glucose and chloride ions, are observed in laboratory tests. In another aspect, immunocytes are another indication for the infection. It has been reported that massive peripheral immunocytes infiltrate across the highly permeable BBB into the subarachnoid space at the early stage of PNBM, especially for leukocytes and neutrophils. However, numerous factors could mimic the infection-related CSF alterations, such as blood, surgical operation, artificial materials and bone dust, triggering the inflammation processes, so the index are not specific enough in the differential diagnosis between infection and aseptic inflammations²². We summarized potential biomarkers in diagnosis of PNBM in currently established studies (Table I).

Even so, the diagnosis of PNBM still predominantly depends on the biochemical results of CSF. Currently, four criteria are widely used for diagnosing PNBM which are proposed by the Centers for Disease Control and Prevention (CDC)²³, the Massachusetts General Hospital (MGH)²⁴, the Infectious Diseases Society of America (IDSA)²⁵ and the Committee of Neurocritical Specialists of China²⁶.

Neuroimage

In most hospitals, neuroimaging has the characteristic of easy operating to be performed routinely to have a preliminary screening for the suspected PNBM and other neurosurgery-related infections. The neuroimage has outstanding per-

Current diagnosis and potential obstacles for post-neurosurgical bacterial meningitis

Table I. Clinical indications for the biomarkers for the potential diagnostic of post-neurosurgical bacterial meningitis.

Clinical indication		Sensitivity	Specificity	Potentially influencing factor	Reported cutoff value for diagnosis	Ref.
Bacterial culture		Low	High	Antibiotic application prior to the sampling	-	25
Gram staining		Low	High	Antibiotic application prior to the sampling	-	
CSF lactate		Moderate to high (76.36% - 97.0%)	Moderate to high (78.0% - 91.6%)	Hemolysis	>3.45 mmol/L to >4 mmol/L	8, 22, 69
CSF procalcitonin		Moderate to high (68.0% - 100.0%)	Moderate to high (66.0% - 100.0%)	-	>0.075 ng/L to >2 ng/L	5, 26, 52, 55
CSF cytokines	IL-6	High (100.0%)	High (100.0%)	-		34
	IL-8	Moderate (67.6%)	Low (46.8%)	-	≥85.5 ng/L	35
	IL-12	High (94.6%) High (83.3%)	Moderate (64.6%) High (85.7%)	-	≥3.2 ng/L	34, 35
	IL-13	Moderate (67.6%)	Low (40.0%)	-	≤42.3 ng/L	35
	IL-17	High (100.0%)	High (100.0%)	-	-	34
	IL-23	High (83.3%)	High (100.0%)	-	-	34
	IFN-γ	High (100.0%)	High (99.9%)	-	≤200 ng/L	35

formance in some infection types, such as an abscess, subdural and epidural empyemas, or other surgical site infection²⁷. Meanwhile, a high-resolution thin section CT with bone window clearly identifies a port-of-entry of the infection. However, the neuroimage has its inherent defect that cannot distinguish the bacterial species, and therefore, it provides poor information about the precision antibiotics treatments.

Gram Staining and Bacterial Culture

Both gram staining and bacterial culture are commonly used in clinical practice towards the suspected PNBM patients. Gram staining is rapid and highly specific. However, it has poor sensitivity in clinical practice, applying only about 20% positivity. Bacterial culture is considered as a gold standard with a high specificity for distinguish exact bacterial species and drug sensitivity types for the PNBM patients. However, probably due to the influences of antibiotics pre- and post-surgery, the positive rate of CSF culture is extremely low, accounting for ~10% in the clinical test and 10-20% in the majority of investigations^{28,29}. Furthermore, the time-consuming character of the bacterial culture, which usually takes more than 24 hours for the results, is always inconducive to the precision antibiotics treatment. The false-positive results which are caused by the contamination also restrict its clinical efficacy in usage. Therefore, according to the standard proposed by the Centers for Disease Control (CDC), patients exhibiting certain symptoms and signs would be categorized as having meningitis even if no organism is detected in the CSF³⁰.

Peripheral Indications

As PNBM also stimulates systemic immunological reactions, some studies31,32 attempted to investigate whether biochemical index in the peripheral blood could indicate the occurrence of meningitis. Zhang et al⁸ compared the index in the blood routine test between 554 patients with PNBM and 868 patients with aseptic inflammation. Unlike the sensitive index in the patients of sepsis, the diagnostic values of the white blood cell (WBC) proportion and neutrophil proportions were inferior to the platelets counts and the Na concentration. However, the area under the curve (AUC) values of the index, which reflect diagnostic accuracies, in blood routine tests were all lower than 0.7, indicating poor diagnostic accuracies for these peripheral biomarkers in PNBM.

CSF Cytokines

Increasing evidence has found that the cytokines in the CSF have potential diagnostic roles in PNBM. Ye et al³³ has found an association of interleukins (ILs) with the bacterial gram staining types in both sepsis and PNBM. The results indicated the IL-6 and IL-10 levels increased in patients with Gram-negative bacteria infection, while the IL-2 level significantly decreased when patients suffered from Gram-negative bacteria infection. However, the results were controversial. Cuff et al34 measured proteomics that included 182 immunological and neurological biomarkers in 14 PNBM patients using the Olink platform. The results indicated that although increased IL-6 level could be observed in the inflammatory response, it could not distinguish between bacterial infection and aseptic inflammation. Within the patient cohort with neurological inflammation, a pattern of raised IL-17, IL-12p40/p70 and IL-23 levels delineated nosocomial bacteriological infection from background neuro-inflammation. Furthermore, Kul et al³⁵ found that the CSF concentrations of IL-8 and IL-12 were upregulated, while IL-13 and IFN-γ were downregulated in the PNBM patients.

Lactate

As a metabolite production, the CSF accumulation of lactate is predominantly derived from the increased productions of anaerobic glucose metabolism, bacterial metabolism and neutrophil glycolysis³⁶. Meanwhile, as the penetration speed of lactate from the peripheral circulation is rather slow, it could be considered that CSF lactate is rarely affected by serum lactate at the early stage of PNBM^{26,37}. Therefore, CSF lactic acid or lactate could directly reflect the brain metabolism in the presence of bacterial infections. Meanwhile, it has also reported that CSF lactate was used to distinguish bacterial from viral meningitis in children^{22,38,39}. Some meta-analyses^{40,41} demonstrated that the CSF lactate level is served as a better marker for bacterial meningitis than conventional markers, such as CSF glucose, CSF protein, and CSF cell count. Zhang et al⁸ found that the receiver operator characteristic (ROC) curve value of CSF lactate on postoperative days >7 was lower than that on postoperative days <7. However, some reports^{42,43} have indicated that the CSF lactate level was influenced by the intrinsic metabolism of CSF red blood cell (RBC). Therefore, the tissue damages in the processes of neurosurgery or lumbar puncture might inevitably release RBC, and subsequently influence the lactate level, leading to unwittingly mock CSF solutions and false-positive results. So, its diagnostic efficacies should be carefully taken into account in some neurosurgical diseases, especially in the hemorrhagic diseases.

Procalcitonin (PCT)

PCT, the propeptide of calcitonin, is commonly considered as an endogenous nonsteroidal substance without a hormone activity^{44,45}. It is reported that PCT is an indicator for the bacterial infection because the bacterial endotoxins and cytokines in the circumstance of infections could blunt the final step of synthesis of calcitonin^{46,47}. thus causing an abundance of the precursor PCT. However, the originality of PCT has been in a matter of debate among scientists. Some researchers^{48,49} believed that the CSF PCT could not be synthesized and released from the brain tissue and was derived from the serum as a result of dysfunction of BBB. Recently, Karzai et al⁵⁰ reported that PCT was released by parenchymal cells when stimulator was present, such as bacterial endotoxins, IL-6 and TNF-α. Muller et al⁵¹ also demonstrated that the mRNA of calcitonin was isolated in hamster brain tissues, providing the possibility and rationality of PCT in the bacterial meningitis. Numerous studies^{52,53} have found the excellently diagnostic values of PCT in PNMB. Viallon et al⁵² and Tomio et al⁵³ drew the same conclusions, demonstrating that the specificity and sensitivity of CSF PCT in diagnosing PNBM could be as high as 100% and 95%, respectively.

However, there are some disagreements for its diagnostic efficacies. It has been reported that PCT concentration usually remained within the normal range even in the presence of a positive bacterial culture in patients with ventriculitis and ventricular catheters. Wang et al⁵ and Laifer et al⁵⁴ found that no significant differences in serum PCT levels were found between PNBM patients with the non-infection patients, but Alkholi et al⁵⁵ reported that serum PCT with cutoff values higher than 2 ng/mL showed sensitivity, specificity, positive predictive value, and negative predictive value of 100%, 66%, 68%, and 100%, respectively, for the diagnosis of bacterial meningitis. Moreover, there has no uniform cutoff value of CSF PCT concentration for the diagnosis. Current studies^{52,55} usually define the positivity of PNBM according to the PCT value are mostly ranging from 0.28 ng/mL to 2 ng/mL. In addition, as Tomio et al⁵³ suggested that PCT had higher levels in the specific typical meningitis agents (pneumococci and meningococci), we assumed that the PCT value, both in CSF and serum, might probably depend on the bacterial species and virulence.

Polymerase Chain Reaction (PCR) and Next-Generation Sequencing (NGS)

In recent decades, PCR and NGS tests for bacterial infections have been widely applied among investigations. Under the principle of reference of 16S rRNA sequence of bacteria, both PCR and NGS could generally detect the bacterial species and drug resistant types with high positive rate^{56,57}. Meanwhile, in comparison with traditional bacterial culture method, PCR and NGS are timesaving which only take several hours to read the consequences. Nonetheless, the technologies have their defects that are still not widely applied in clinical laboratory tests to support a final diagnosis. PCR test has a high false positive rate due to the sample contamination during the lumbar puncture and the interaction between the gene probes and other unrelated germ, resulting in a low specificity of the technique^{58,59}. According to another study 60 , NGS has a high-throughput characters, allowing thousands of samples tested simultaneously. However, the expensive device and the related reagents, complex and time-consuming operation restrict its application for clinical routine tests. Furthermore, what the clinicians or laboratorians mostly care about is that it could not distinguish the bacterial types which have predominantly infectious property when multiple bacterial infections occur by NGS test.

Key Barriers in Clinical and Scientific Research

We roughly screened the established clinical and mechanical investigations through the PUBMED database using the keyword of "post-neurosurgical bacterial meningitis", but there are just hundreds of related studies. In comparison with other investigations on a certain disease, the numbers of the study on PNBM are extremely limited. Owing to the low diagnostic rate and the indefinite information about the infectious bacterial species, it is difficult to collect enough size of positive samples and also to make a definite research target and strategy. Additionally, due to the prophylactic antibiotic treatments which could uncover the phenotypes of infections, it could hardly eliminate the influences for the bacteria from the antibiotics. Given that these potential obstacles restrain the scientists on performing relative studies on this field, the most important issue on PNBM is to develop an effective method on recognizing the infection occurrence and the bacterial species.

Hypotheses in Scientific Studies Of PNBM

Bacterial Translocation from Gut Causes the Infection

Due to the lack of sufficient studies, the originality of infectious bacteria is unclear. Previous studies^{14,61} indicated that the infection might be derived from colonized cutaneous organisms, even though the surgery was free of pathogens. The environment of CNS was considered to be privileged for pathogens and the immunocytes in the peripheral circulations, but current attitudes have changed a lot. BBB prohibits the various materials from the brain⁶². Whereas, in most of CNS diseases, the physiological structures of cerebral vessels are changed, leading to the destruction of BBB. As a consequence, various materials could penetrate from the permeable BBB. Kigerl et al⁶³ reported that in the animal model of spinal cord injury (SCI), the gut bacteria translocated to multiple abdominal organs through the destructed gastrointestinal mucosa, such as lung, liver, spleen and mesenteric lymph nodes. Therefore, we might hypothesize a probability that gut bacteria can also translocate to the CNS environment through the dysfunctional BBB, leading to the bacterial meningitis. However, the correlation study between the bacterial meningitis and gut bacterial translocation after brain injury or craniotomy are still blank.

In Situ Infection by the Intra-Tumoral Bacteria

In 2020, Nejman et al⁶⁴ tested the existence of bacteria in seven cancers, including breast, lung, ovary, pancreas, melanoma, bone, and brain tumors. Interestingly, the results found that the intra-tumoral bacteria existed mostly in intracellular region, and the intra-tumoral bacteria might exert certain effects on influencing the phenotypes of tumors and the response to therapies. Meanwhile, in spite of the pathogen free characters and aseptic technique applied in the neurosurgery, especially for the neurosurgery on the brain tumors, the rate of PNBM on the brain tumor-related neurosurgery is supposed to be lower than the other type of the neurosurgical diseases⁸. However, the epidemiological studies on the PNBM of brain tumors have

discovered that the incidence rate of PNBM on brain tumors was generally ranging from 6.8% to 7.9%, in spite of tumor types, indicating a slightly higher rate^{8,10}. Based on the situation, we hypothesize a potential relationship between the releases from the tumor-colonized bacteria and the PNBM occurrence. Therefore, studying on the originality of infected bacteria is essential for guidance of surgical methods in clinical practice.

Genetic Susceptibility in PNBM

The bacterial genetic variations for the drug resistance have been widely investigated, indicating that the bacteria have the genetic evolution with drugs, but there is no report investigating the potentially genetic risks for the patients to be affected with bacteria. Previously established evidence^{65,66} supported the attitude that the risk of PNBM predominantly influenced by both the bacterial virulence and the immunological status of individuals. It seems that the infectious disease has little relationship with the genetics factors. Interestingly, we found that a study group by Zhang et al⁶⁷ has intensively screened the genetic risks for the leprosy infection, a disease caused by mycobacterium leprae. Multiple genes and related gene pathways were discovered to be associated with susceptibility in the infection of M. leprae⁶⁸. It provides a clue that all the patients who underwent neurosurgical processes have potential exposure risks to be infected by bacteria. Whether the genetic factors influence the susceptibility of PNBM may be targeted for further investigation.

Prospect

Although the current incidence rate of PNBM is lower than that in the last century because of the advanced technologies applied in the neurosurgical processes, it seems unchanged during the recent decades. The bottleneck on meningitis is still attributed to the low recognition rate of microbe species and related drug-resistance types. Except for the definite results in bacterial culture. most diagnostic methods and biomarkers could only provide suggestive information about the inflammatory or infectious status of patients. We infer that this problem might be of greatest priority in both clinical therapies and laboratory studies. Meanwhile, two prospects in the PNBM can be taken into account, which might open a new insight for the disease. First, we should explore a novel method with high sensitivity and specificity to make a definite recognition of bacterial species and drug-resistance types. Second, given that we stated in the above context, the originality of microbe should be carefully noticed.

Conclusions

In this review, we summarize the established diagnostic methods and biomarkers for PNBM. Meanwhile we also propose some potential investigation prospects. This review may help to better understand the character of PNBM in both clinical diagnosis and scientific investigations.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors' Contributions

LY and HTX designed the study. PG, XM, ZYL, WHW and BSW collected and interpreted the literature. LY, HTX and SS drafted the manuscript. YQZ and HWC revised the manuscript. All authors read and approved the final manuscript.

Funding

The work was supported by the National Natural Science Foundation of China (No. 81901238), the University Natural Science Research Project of Anhui Province (grant No. KJ2019A0248), and Co-construction Project of Anhui Medical University and Affiliated Hospital (grant No. 2021lcxk017).

Institutional Review Board Statement Not applicable.

Informed Consent Statement Not applicable.

Data Availability Statement Not applicable.

References

 Collaborators GBDN. Global, regional, and national burden of neurological disorders, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet Neurol 2019; 18: 459-480.

- Yu Y, Li HJ. Diagnostic and prognostic value of procalcitonin for early intracranial infection after craniotomy. Braz J Med Biol Res 2017; 50: e6021.
- McClelland S, 3rd. Postoperative intracranial neurosurgery infection rates in North America versus Europe: a systematic analysis. Am J Infect Control 2008; 36: 570-573.
- 4) Yin L, Han Y, Miao G, Jiang L, Xie S, Liu B. CSF leukocyte, polykaryocyte, protein and glucose: Their cut-offs of judging whether post-neurosurgical bacterial meningitis has been cured. Clin Neurol Neurosurg 2018; 174: 198-202.
- Wang H. Higher Procalcitonin Level in Cerebrospinal Fluid than in Serum Is a Feasible Indicator for Diagnosis of Intracranial Infection. Surg Infect 2020; 21: 704-708.
- Chen Y, Zhang L, Qin T, Wang Z, Li Y, Gu B. Evaluation of neurosurgical implant infection rates and associated pathogens: evidence from 1118 postoperative infections. Neurosurg Focus 2019; 47: E6.
- Blomstedt GC. Infections in neurosurgery: a retrospective study of 1143 patients and 1517 operations. Acta Neurochir 1985; 78: 81-90.
- Zhang Y, Xiao X, Zhang J, Gao Z, Ji N, Zhang L. Diagnostic accuracy of routine blood examinations and CSF lactate level for post-neurosurgical bacterial meningitis. Int J Infect Dis 2017; 59: 50-54.
- McCutcheon BA, Ubl DS, Babu M, Maloney P, Murphy M, Kerezoudis P, Bydon M, Habermann EB, Parney I. Predictors of Surgical Site Infection Following Craniotomy for Intracranial Neoplasms: An Analysis of Prospectively Collected Data in the American College of Surgeons National Surgical Quality Improvement Program Database. World Neurosurg 2016; 88: 350-358.
- 10) Shi ZH, Xu M, Wang YZ, Luo XY, Chen GQ, Wang X, Wang T, Tang MZ, Zhou JX. Post-craniotomy intracranial infection in patients with brain tumors: a retrospective analysis of 5723 consecutive patients. Br J Neurosurg 2017; 31: 5-9.
- 11) Richardson D, Duncan C, Sinha A, Hennedige AA. Pseudomeningocele With Orbital Extension as a Complication of Fronto-Orbital Advancement and Remodeling in Craniosynostosis. J Craniofac Surg 2015; 26: 2142-2147.
- Leib SL, Boscacci R, Gratzl O, Zimmerli W. Predictive value of cerebrospinal fluid (CSF) lactate level versus CSF/blood glucose ratio for the diagnosis of bacterial meningitis following neurosurgery. Clin Infect Dis 1999; 29: 69-74.
- 13) Moon C, Kwak YG, Kim BN, Kim ES, Lee CS. Implications of postneurosurgical meningitis caused by carbapenem-resistant Acinetobacter baumannii. J Infect Chemother 2013; 19: 916-919.
- 14) Talibi S, Tarnaris A, Shaw SA. Has the introduction of antibiotic-impregnated external ventricular drain catheters changed the nature of the microorganisms cultured in patients with drain-related infection? A single neurosurgical centre's experience. Br J Neurosurg 2016; 30: 560-566.

- Laxmi S, Tunkel AR. Healthcare-associated bacterial meningitis. Curr Infect Dis Rep 2011; 13: 367-373.
- Willyard C. The drug-resistant bacteria that pose the greatest health threats. Nature 2017; 543: 15.
- Huang Q, Zhou Q, Ju T, Xu H, Wang W. Meropenem and Amikacin for Management of Post-Neurosurgical Infections from Acinetobacter baumannii. Surg Infect 2019; 20: 292-297.
- Cherian A, Baheti NN, Easwar HV, Nair DS, Iype T. Recurrent meningitis due to epidermoid. J Pediatr Neurosci 2012; 7: 47-48.
- 19) Cyrus SS, Advisory Committee on Immunization Practices of the Centers for Disease C, Prevention. Protecting the world against meningitis: new recommendations from the CDC's advisory committee on immunization practices. J Am Osteopath Assoc 2011; 111: S17-19.
- Sweeney MD, Zhao Z, Montagne A, Nelson AR, Zlokovic BV. Blood-Brain Barrier: From Physiology to Disease and Back. Physiol Rev 2019; 99: 21-78.
- Zhao Z, Nelson AR, Betsholtz C, Zlokovic BV. Establishment and Dysfunction of the Blood-Brain Barrier. Cell 2015; 163: 1064-1078.
- 22) Maskin LP, Capparelli F, Mora A, Hlavnicka A, Orellana N, Díaz MF, Wainsztein N, Del Castillo M. Cerebrospinal fluid lactate in post-neurosurgical bacterial meningitis diagnosis. Clin Neurol Neurosurg 2013; 115: 1820-1825.
- 23) Korinek AM, Baugnon T, Golmard JL, van Effenterre R, Coriat P, Puybasset L. Risk factors for adult nosocomial meningitis after craniotomy: role of antibiotic prophylaxis. Neurosurgery 2008; 62 Suppl 2: 532-539.
- 24) Durand ML, Calderwood SB, Weber DJ, S I Miller, Southwick FS, Caviness VS Jr, Swartz MN. Acute bacterial meningitis in adults. A review of 493 episodes. N Engl J Med 1993; 328: 21-28.
- 25) Tunkel AR, Hasbun R, Bhimraj A, Byers K, Kaplan SL, Scheld WM, van de Beek D, Bleck TP, Garton HJL, Zunt JR. 2017 Infectious Diseases Society of America's Clinical Practice Guidelines for Healthcare-Associated Ventriculitis and Meningitis. Clin Infect Dis 2017; 64: e34-e65.
- 26) Li Y, Zhang G, Ma R, Du Y, Zhang L, Li F, Fang F, Lv H, Wang Q, Zhang Y, Kang X. The diagnostic value of cerebrospinal fluids procalcitonin and lactate for the differential diagnosis of post-neurosurgical bacterial meningitis and aseptic meningitis. Clin Biochem 2015; 48: 50-54.
- 27) Kim YJ, Moon KS, Kim SK, Kang SJ, Lee KH, Jang WY, Jung TY, Kim IY, Jung S. The difference in diffusion-weighted imaging with apparent diffusion coefficient between spontaneous and postoperative intracranial infection. Br J Neurosurg 2014; 28: 765-770.
- Favaro M, Savini V, Favalli C, Fontana C. A multi-target real-time PCR assay for rapid identification of meningitis-associated microorganisms. Mol Biotechnol 2013; 53: 74-79.

- 29) Wu HM, Cordeiro SM, Harcourt BH, Carvalho M, Azevedo J, Oliveira TQ, Leite MC, Salgado K, Reis MG, Plikaytis BD, Clark TA, Mayer LW, Ko AI, Martin SW, Reis JN. Accuracy of real-time PCR, Gram stain and culture for Streptococcus pneumoniae, Neisseria meningitidis and Haemophilus influenzae meningitis diagnosis. BMC Infect Dis 2013; 13: 26.
- Hussein K, Bitterman R, Shofty B, Paul M, Neuberger A. Management of post-neurosurgical meningitis: narrative review. Clin Microbiol Infect 2017; 23: 621-628.
- 31) Lembo RM, Rubin DH, Krowchuk DP, McCarthy PL. Peripheral white blood cell counts and bacterial meningitis: implications regarding diagnostic efficacy in febrile children. Pediatr Emerg Care 1991; 7: 4-11.
- 32) He H, Zou Y, He J, Bu H, Liu Y. A Diagnostic Scoring System for Distinguishing between Tuberculous and Bacterial Meningitis Based on Clinical and Laboratory Findings. Biomed Res Int 2021; 2021: 1220650.
- 33) Ye Q, Shao WX, Xu XJ, Yang YZ. The clinical application value of cytokines in treating infectious diseases. Plos One 2014; 9: e98745.
- 34) Cuff SM, Merola JP, Twohig JP, Eberl M, Gray WP. Toll-like receptor linked cytokine profiles in cerebrospinal fluid discriminate neurological infection from sterile inflammation. Brain Commun 2020; 2: fcaa218.
- 35) Kul G, Sencan I, Kul H, Korkmaz N, Altunay E. The Role of Cerebrospinal Fluid Biomarkers in the Diagnosis of Post-Neurosurgical Meningitis. Turk Neurosurg 2020; 30: 513-519.
- 36) Guerra-Romero L, Tauber MG, Fournier MA, Tureen JH. Lactate and glucose concentrations in brain interstitial fluid, cerebrospinal fluid, and serum during experimental pneumococcal meningitis. J Infect Dis 1992; 166: 546-550.
- 37) Cameron PD, Boyce JM, Ansari BM. Cerebrospinal fluid lactate in meningitis and meningococcaemia. J Infect 1993; 26: 245-252.
- Cunha BA. Distinguishing bacterial from viral meningitis: the critical importance of the CSF lactic acid levels. Intensive Care Med 2006; 32: 1272-1273; author reply 1274.
- 39) de Almeida SM, Furlan SMP, Cretella AMM, Lapinski B, Nogueira K, Cogo LL, Vidal LRR, Nogueira MB. Comparison of Cerebrospinal Fluid Biomarkers for Differential Diagnosis of Acute Bacterial and Viral Meningitis with Atypical Cerebrospinal Fluid Characteristics. Med Princ Prac 2020; 29: 244-254.
- Xiao X, Zhang Y, Zhang L, Kang P, Ji N. The diagnostic value of cerebrospinal fluid lactate for post-neurosurgical bacterial meningitis: a meta-analysis. BMC Infect Dis 2016; 16: 483.
- Sakushima K, Hayashino Y, Kawaguchi T, Jackson JL, Fukuhara S. Diagnostic accuracy of cerebrospinal fluid lactate for differentiating bacterial meningitis from aseptic meningitis: a meta-analysis. J Infect 2011; 62: 255-262.

- 42) Venkatesh B, Morgan TJ, Boots RJ, Hall J, Siebert D. Interpreting CSF lactic acidosis: effect of erythrocytes and air exposure. Crit Care Resus 2003; 5: 177-181.
- 43) Froman C, Smith AC. Metabolic acidosis of the cerebrospinal fluid associated with subarachnoid haemorrhage. Lancet 1967; 1: 965-967.
- 44) Riche FC, Cholley BP, Laisne MJ, Vicaut E, Panis YH, Lajeunie EJ, Boudiaf M, Valleur PD. Inflammatory cytokines, C reactive protein, and procalcitonin as early predictors of necrosis infection in acute necrotizing pancreatitis. Surgery 2003; 133: 257-262.
- 45) Casado-Flores J, Blanco-Quiros A, Nieto M, Asensio J, Fernandez C. Prognostic utility of the semi-quantitative procalcitonin test, neutrophil count and C-reactive protein in meningococcal infection in children. Eur J Pediatr 2006; 165: 26-29.
- 46) Gogos CA, Drosou E, Bassaris HP, Skoutelis A. Pro- versus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. J Infect Dis 2000; 181: 176-180.
- Foushee JA, Hope NH, Grace EE. Applying biomarkers to clinical practice: a guide for utilizing procalcitonin assays. J Antimicrob Chemother 2012; 67: 2560-2569.
- 48) Shimetani N, Shimetani K, Mori M. Levels of three inflammation markers, C-reactive protein, serum amyloid A protein and procalcitonin, in the serum and cerebrospinal fluid of patients with meningitis. Scand J Clin Lab Invest 2001; 61: 567-574.
- 49) Jereb M, Muzlovic I, Hojker S, Strle F. Predictive value of serum and cerebrospinal fluid procalcitonin levels for the diagnosis of bacterial meningitis. Infection 2001; 29: 209-212.
- 50) Karzai W, Oberhoffer M, Meier-Hellmann A, Reinhart K. Procalcitonin--a new indicator of the systemic response to severe infections. Infection 1997; 25: 329-334.
- 51) Muller B, White JC, Nylen ES, Snider RH, Becker KL, Habener JF. Ubiquitous expression of the calcitonin-i gene in multiple tissues in response to sepsis. J Clin Endocrinol Metab 2001; 86: 396-404.
- 52) Viallon A, Desseigne N, Marjollet O, Birynczyk A, Belin M, Guyomarch S, Borg J, Pozetto B, Bertrand JC, Zeni F. Meningitis in adult patients with a negative direct cerebrospinal fluid examination: value of cytochemical markers for differential diagnosis. Crit Care 2011; 15: R136.
- 53) Tomio R, Akiyama T, Shibao S, Yoshida K. Procalcitonin as an early diagnostic marker for ventriculoperitoneal shunt infections. Surgical infections. 2013; 14: 433-436.
- 54) Laifer G, Wasner M, Sendi P, P Graber, Gratzl O, Huber P, Fluckiger U, Zimmerli W. Dynamics of serum procalcitonin in patients after major neurosurgery. Clin Microbiol Infect 2005; 11: 679-681.
- Alkholi UM, Abd Al-Monem N, Abd El-Azim AA, Sultan MH. Serum procalcitonin in viral and bacterial meningitis. J Glob Infect Dis 2011; 3: 14-18.

- 56) Grumaz S, Grumaz C, Vainshtein Y, Stevens P, Glanz K, Decker SO, Hofer S, Weigand MA, Brenner T, Sohn K. Enhanced Performance of Next-Generation Sequencing Diagnostics Compared With Standard of Care Microbiological Diagnostics in Patients Suffering From Septic Shock. Crit Care Med 2019; 47: e394-e402.
- 57) Liu CF, Shi XP, Chen Y, Jin Y, Zhang B. Rapid diagnosis of sepsis with TaqMan-Based multiplex real-time PCR. J Clin Lab Anal 2018; 32: e22256.
- 58) Lleo MM, Ghidini V, Tafi MC, Castellani F, Trento I, Boaretti M. Detecting the presence of bacterial DNA by PCR can be useful in diagnosing culture-negative cases of infection, especially in patients with suspected infection and antibiotic therapy. FEMS Microbiol Lett 2014; 354: 153-160.
- 59) Simon TD, Van Yserloo B, Nelson K, Gillespie D, Jensen R, McAllister JP 2nd, Riva-Cambrin J, Stockmann C, Daly JA, Blaschke AJ. Use of quantitative 16S rRNA PCR to determine bacterial load does not augment conventional cerebrospinal fluid (CSF) cultures among children undergoing treatment for CSF shunt infection. Diagn Microbiol Infect Dis 2014; 78: 188-195.
- 60) Zhang G, Zheng G, Zhang Y, Ma R, Kang X. Evaluation of a micro/nanofluidic chip platform for the high-throughput detection of bacteria and their antibiotic resistance genes in post-neurosurgical meningitis. Int J Infect Dis 2018; 70: 115-120.
- 61) Schade RP, Schinkel J, Visser LG, Van Dijk JM, Voormolen JH, Kuijper EJ. Bacterial meningitis caused by the use of ventricular or lumbar cerebrospinal fluid catheters. J Neurosurg 2005; 102: 229-234.
- 62) Engelhardt B, Carare RO, Bechmann I, Flugel A, Laman JD, Weller RO. Vascular, glial, and lymphatic immune gateways of the central nervous system. Acta Neuropathol 2016; 132: 317-338.
- 63) Kigerl KA, Hall JC, Wang L, Mo X, Yu Z, Popovich PG. Gut dysbiosis impairs recovery after spinal cord injury. J Exp Med 2016; 213: 2603-2620.
- Nejman D, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, Rotter-Maskowitz A, Weiser R, Mallel G, Gigi E, Meltser A, Douglas GM, Kamer I, Gopalakrishnan V, Dadosh T, Levin-Zaidman S, Avnet S, Atlan T, Cooper ZA, Arora R, Cogdill AP, Khan MAW, Ologun G, Bussi Y, Weinberger A, Lotan-Pompan M, Golani O, Perry G, Rokah M, Bahar-Shany K, Rozeman EA, Blank CU, Ronai A, Shaoul R, Amit A, Dorfman T, Kremer R, Cohen ZR, Harnof S, Siegal T, Yehuda-Shnaidman E, Gal-Yam EN, Shapira H, Baldini N, Langille MGI, Ben-Nun A, Kaufman B, Nissan A, Golan T, Dadiani M, Levanon K, Bar J, Yust-Katz S, Barshack I, Peeper DS, Raz DJ, Segal E, Wargo JA, Sandbank J, Shental N, Straussman R. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. Science 2020; 368: 973-980.
- 65) Yi HJ, Jeong JH, Jin ES, Shin IY, Hwang HS, Moon SM. Evaluation of vitamin D level in patients from neurosurgical intensive care unit. Neural Regen Res 2013; 8: 1528-1534.

- 66) Gong J, Li D, Yan J, Liu Y, Li D, Dong J, Gao Y, Sun T, Yang G. The accessory gene regulator (agr) controls Staphylococcus aureus virulence in a murine intracranial abscesses model. Braz J Infect Dis 2014; 18: 501-506.
- 67) Zhang FR, Huang W, Chen SM, Sun LD, Liu H, Li Y, Cui Y, Yan XX, Yang HT, Yang RD, Chu TS, Zhang C, Zhang L, Han JW, Yu GQ, Quan C, Yu YX, Zhang Z, Shi BQ, Zhang LH, Cheng H, Wang CY, Lin Y, Zheng HF, Fu XA, Zuo XB, Wang Q, Long H, Sun YP, Cheng YL, Tian HQ, Zhou FS, Liu HX, Lu WS, He SM, Du WL, Shen M, Jin QY, Wang Y, Low HQ, Erwin T, Yang NH, Li JY, Zhao X, Jiao YL, Mao LG, Yin G, Jiang ZX, Wang XD, Yu JP, Hu ZH, Gong CH, Liu YQ, Liu RY, Wang
- DM, Wei D, Liu JX, Cao WK, Cao HZ, Li YP, Yan WG, Wei SY, Wang KJ, Hibberd ML, Yang S, Zhang XJ, Liu JJ. Genomewide association study of leprosy. N Engl J Med 2009; 361: 2609-2618.
- 68) Zhang X, Cheng Y, Zhang Q, Wang X, Lin Y, Yang C, Sun J, Huang H, Li Y, Sheng Y, Fan X, Sun Y, Zhang X, Zheng X, Zhang B, Yang S. Meta-Analysis Identifies Major Histocompatibility Complex Loci in or Near HLA-DRB1, HLA-DQA1, HLA-C as Associated with Leprosy in Chinese Han Population. J Invest Dermatol 2019; 139: 957-960.
- Lotfi R, Ines B, Aziz DM, Mohamed B. Cerebrospinal Fluid Lactate as an Indicator for Post-neurosurgical Bacterial Meningitis. Indian J Crit Care Med 2019; 23: 127-130.