

Tissue engineering as innovative chance for organ replacement in radical tumor surgery

C. ALBERTI

L.D. of Surgical Semeiotics, Parma, Italy

Abstract. – BACKGROUND: Different pathological conditions such as congenital organ absence, severe organ injuries, end-stage organ failure and malignancy-related organ removal, have few effective therapeutic options a part from a whole organ transplant, that, however, often meets with a serious shortage of suitable donor organs.

AIM: The purpose of this paper consists in highlighting what the novel tissue engineering approaches might help to solve such problems.

EMERGING CONCEPTS: A recent approach in tissue/organ engineering, particularly to build bioartificial airways, is the procedure of decellularizing a whole donor organ to obtain a complex 3D-biomatrix-scaffold maintaining the intrinsic vascular network, that is subsequently recellularized with recipient's autologous organ-specific differentiated cells or/and stem cells, to build a potentially functional biological substitute. Such strategy has been clinically used to replace organ in trachea/bronchus tumor patients. In another approach, mainly used to construct a bioartificial urinary bladder tissue, different types of either biodegradable synthetic polymers or naturally-derived matrices or even polymer/biomatrix-composite materials are used as scaffold for either cell-free or autologous cell-seeded tissue engineering procedures. So far, such technique has been mainly used to make an augmentation cystoplasty in patients with end-stage poorly compliant neuropathic bladder or in exstrophic bladder subjects.

FUTURE PROSPECTS: Intriguing developments in biomaterial science, nanotechnologies, stem cell biology, and further improvements in bioreactor manufacturing will allow to generate, in the near future, tissue engineered organs that, as for structure/function so the native one-like, might represent the optimum solution to replace organs in tumor surgery.

Key Words:

Trachea, Bladder, Biomatrix, Biomaterials, Nanotechnology, Stem-cells.

Introduction

A quite recent intriguing work points out that a rat subtotal cystectomy can elicit an early strong multilayer bladder wall regeneration – resulting in

fully morpho-functional bladder reconstitution – that is unique among mammalian organ systems as it seems to peculiarly mimic salamander limb regeneration via *blastema formation* (cell dedifferentiation, epithelial-mesenchymal transdifferentiation, tissue-specific progenitor cell expansion) rather than the quick mammals liver *compensatory hyperplasia* following a large loss/surgical removal of hepatic tissue¹. The last occurrence recalls the mythologic Greek story of Prometheus – god of fire – whose liver, because of punishment inflicted on him by Zeus, every day was torn by an eagle and every night re-grew.

Unfortunately, such rodent bladder wall strong regeneration modality is little natively embodied in the humans where the ensuing result of a large partial cystectomy, without an appropriate patching cystoplasty, is a severely limited capacity-characterized uncompliant bladder. What also occurs for either damaged or surgically excised other hollow organs – among whose the complex tissue structured trachea – that's why different tissue engineering technologies may be helpful to construct native organ-like morpho-functional integrity endowed substitutes particularly to replace organs in surgical oncology.

As for the tissue engineering strategies mainly accomplished to build replacement organs, quite paradigmatic are those respectively concerning either trachea or bladder tissue fabrication.

The design of variously shaped tissue constructs – such as flat (skin), both nonviscus tubular structures (male urethra) and viscus hollow organs (bladder, trachea, vagina) or complex solid organ architectures (kidney, liver) – needs, anyhow, biomaterial-made/ECM (extracellular matrix) scaffold, autologous either mature or stem cells, specific growth/differentiation factors.

Tissue Engineering Main Procedure to Build Airways

The main approach to construct an airway tissue/organ consists in resorting to natural extracellular matrix, obtained by decellularizing a

donor trachea, because synthetic biodegradable polymer-based scaffolds – such polyglycolic and (PGA)-, polylactic acid (PLA)-, coPGA/PLA – proved inadequate to clinical applications². Indeed, at first, a decellularized human donor wind-pipe, suitably repopulated via an appropriate *ex vivo* bioreactor, with cultured autologous respiratory epithelial cells (REC) and bone marrow-mesenchymal stromal cell-derived chondrocytes, was successfully used as replacement graft of critically diseased main bronchus⁴ (Figure 1).

Intriguing improvements of such technique have been promptly carried-out by shortening the decellularization time of the human donor explanted trachea, hence by intraoperatively seeding that with autologous REC and bone marrow-derived

monocytes and then resorting to recipient's own body as *in vivo* bioreactor⁵. Such innovative quick procedure – mainly *in vivo* trachea tissue regeneration on implanted recellularized donor biomatrix-scaffold – was subsequently adopted in nine patients with serious-either congenital or acquired, particularly malignant diseases of airways. Among these patients, a partial collapse of the scaffold occurred in three cases, thus it entailing proper refinements in different phases of this approach^{6,7}. Particularly, it has been shown that the chemical-enzymatic decellularization modalities of donor trachea can induce a biomatrix critically decline in soluble type II collagen and glycosaminoglycans (GAG) content, thus compromising the mechanical integrity of the tracheal scaffold⁸.

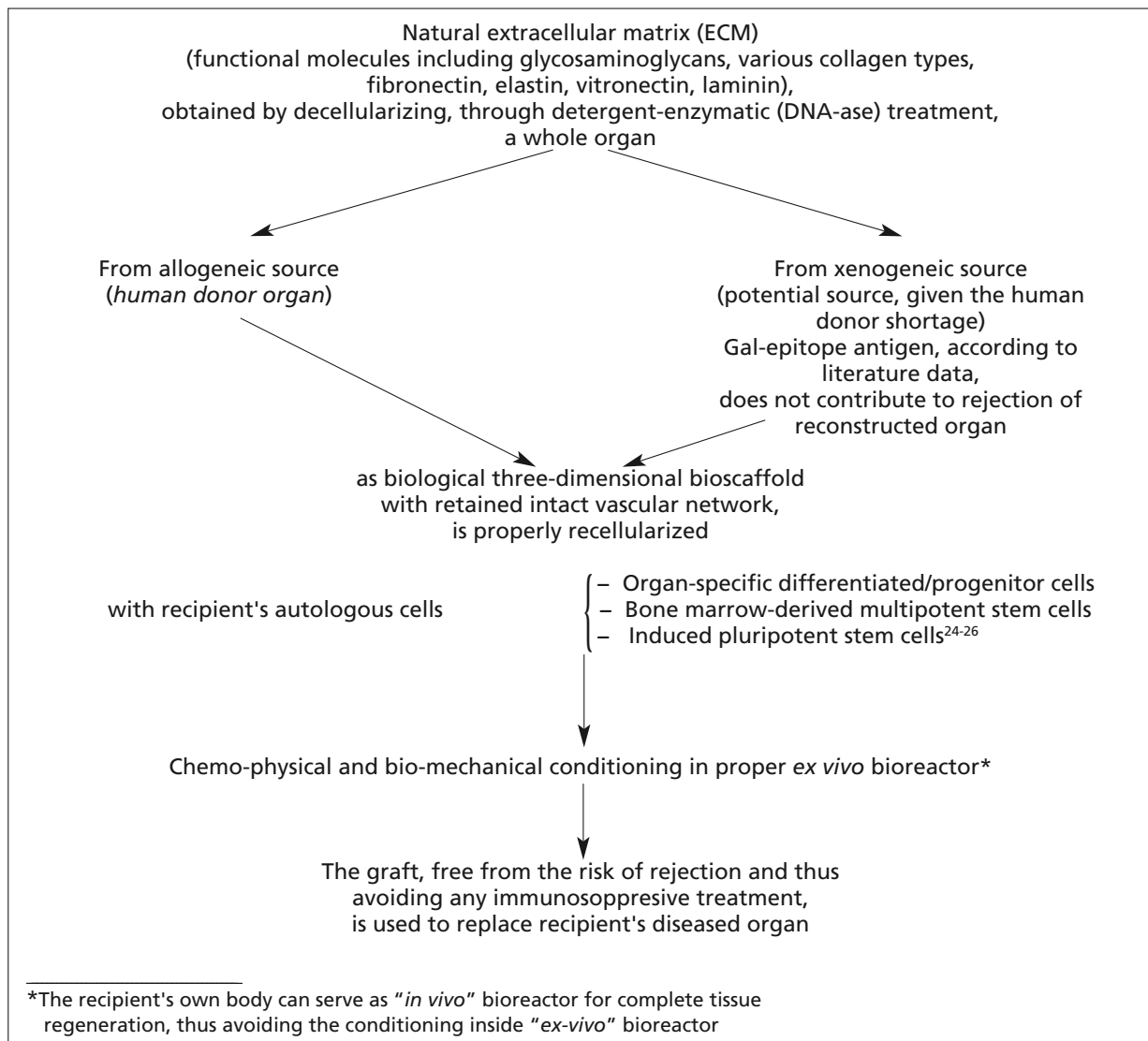


Figure 1. Tissue engineering strategy from naturally-derived extracellular matrix-based scaffold.

Given the above biomatrix-related problems and, in addition, the difficulties in obtaining suitable donor organs, the attention has been turned to fabrication of tailored tracheal/bronchial-shaped nanostructured polymeric scaffolds, endowed with native tissue-like mechanical features, properly *in vitro* seeded with autologous REC and bone marrow-derived stromal cells, via *in vitro* bioreactor, to reach a phenotypic and functional appropriate maturation^{3,9,10}. Such *nanocomposite scaffold*, seeded with autologous stem cells, has been successfully implanted into a subject suffering from recurrent primary tracheo-bronchial tumor, together with enhancing the *in vivo* airway wall regeneration by use of specific bioactive agents and growth factors, directed to attract, within the scaffold, peripheral and local progenitor/stem cells¹⁰.

Even though considering these innovative technological contributions, the already proven procedure of using a decellularized donor tracheal scaffold, seeded with autologous bone-marrow mesenchymal stem cells plus autologous epithelium patches, and properly boosting the angiogenesis and chondrogenesis within the engineered tissue, has been clinically validated in recent 2 year follow-up study¹¹. Thus it is strengthened that to reach clinically successful whole airway replacement by tissue engineering technologies, a 3D-bioscaffold, composed of nonautologous source-derived biomatrix, hence recellularized with autologous either stem cells of differentiated cells, together with using specific bioagents to boost a proper stem cell recruitment/mobilization, represents an effective tissue engineered prosthesis^{3,12}.

Intriguingly, an experimental investigation has been carried out, in a pig model, to study *in vivo* regeneration of decellularized pig trachea – without recellularization before its transplantation – by intraoperatively treating that with specific growth/differentiation factors and mononuclear cells, and then by using the recipient pig's own body as *in vivo* bioreactor. The post-operative controls showed quick trachea *in vivo* reconstruction (just after two weeks), provided with respiratory tissue, thus validating such *in vivo* airway tissue engineering strategy¹³.

Tissue Engineering Main Procedure to Build the Urinary Bladder

To avoid both metabolic and malignant problematic complications of the intestinal neobladder – however it still remaining the gold standard

of the urinary diversion following radical cystectomy in bladder tumor patients – different solutions have been proposed to get an artificial neobladder: from alloplastic nonbiomaterial-made bladder – by using polyurethane, polytetrafluoroethylene, silicon rubber – that are discarded because of nonbiodegradable material-related negative outcomes, to, more recently, tissue-engineered bladder by using autologous urothelial- and smooth muscle-cells seeded onto biocompatible either synthetic (PGA, PLA, coPGA/PLA) or natural ECM scaffolds, the just-mentioned synthetic ones showing termic stability under various body temperature conditions together with both enzymatic-hydrolytic biodegradability and absence of toxicity (Figure 2)¹⁴⁻²³.

Current technologies to obtain an augmentation cystoplasty with a bladder tissue engineered include both *unseeded* (cell free matrix) and *seeded* (cell matrix) modalities. The *unseeded method* consists in anastomosing a naturally-derived acellular matrix – particularly collagen sponge, small intestinal submucosa (SIS), bladder acellular matrix (BAM) – with host bladder to induce, *in vivo*, a natural biomatrix-guided vesical wall cell-repopulation from both urothelial and smooth muscle cells, arising from the neighbouring native bladder tissue or/and even from the ureters when directly implanted into biomatrix¹⁶⁻¹⁹. Porcine urinary bladder matrix (UBM) seems to be provided with significantly higher potential, compared with SIS, to support the growth of human urothelial cells²⁰. The *seeded method* is characterized by seeding cultured autologous urothelial and smooth muscle cells, obtained from the host urinary tract tissue, onto either synthetic biodegradable material (PGA, PLA, co-PGA/PLA)- or natural matrix (collagen, SIS, BAM)-made scaffold to *in vitro* build a morpho-functionally suitable replacement tissue²¹⁻²³ (Figure 2). Anyway made, tissue engineered bladder must have peculiar native bladder-like properties, some of them properly urothelium-related – urine permeability barrier, intravesical pressure-sensitive transducer function, effective ECM-cytoskeleton-nuclear matrix interactions by both various shuttle molecule/growth factor-mediated chemical cell signaling and extra/intracellular microelectric current/bioresonance biophysical connections – while others smooth muscle layer-linked such its specific dynamics^{14,27-30}.

Getting down to tissue engineering clinical implementations, bladder augmentation with *acellular matrix*, in exstrophic bladder patients,

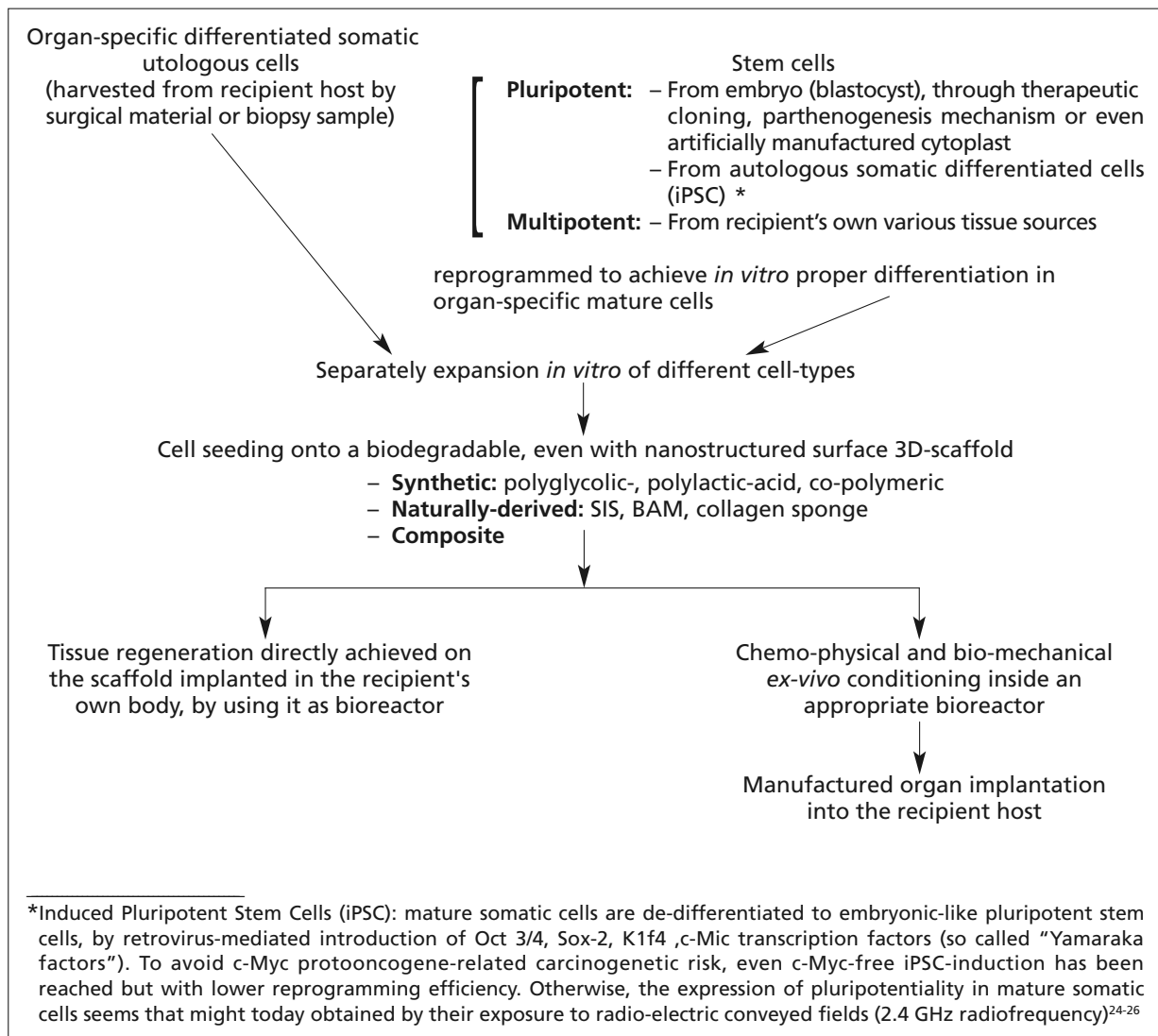


Figure 2. Tissue engineering strategy from biodegradable polymer-based scaffold.

failed to provide long-term effective outcomes as for both bladder capacity and compliance together with urinary continence³¹. Instead, a *cell seeded composite 3D-bladder engineered tissue*, made up of collagen-biomatrix plus polyglycolic acid, has been implanted, with omental drape, in patients with end-stage neuropathic poorly compliant bladder to successfully reach an augmentation cystoplasty^{32,33}.

Because of potential limitations of *in vitro* urinary tract-derived autologous cells – possible complications due to invasive tissue biopsy, precariousness of specimens from a widely unhealthy organs, low *in vitro* proliferation of adult organ-derived cells – the resort has been taken into consideration, for cell-based bladder engineering, to either pluripotent/multipotent *stem*

cells, endowed with self-renewal and differentiation in various tissue-specific cell-lineages, or proper progenitor cells. Among pluripotent stem cells, the interest in the induced pluripotent stem cells (iPSC) is more and more increasing^{24-26,34-37} (Figure 2). On this subject, even amniotic fluid-, placenta-, umbilical cord-derived stem cells and, in addition, those isolated in urine samples collected from upper urinary tract, could be an alternative cell source to build a tissue engineered neobladder for vesical tumor patients undergone total cystectomy^{38,39}.

Recent advances in the field of nanostructured biomaterials – *polymer surface nanofabrication* – that can mimic the nanoscale topography of native tissues with optimum cell/scaffold interactions, together with the innovative design of vari-

ous *smart biomaterials* (materials provided with specific protein domains such as RDG, arginine-glycine-aspartate, a cell-binding domain of fibronectin; gene-engineering-induced mutants of natural proteins; etc), and the resort to *hydrogels for bioprinting* applications, have significantly enhanced the tailored tissue engineering-related challenges^{38,40-51}. As for as *gene-engineering* – gene (nucleic acids) delivery to a variety of cell populations such as nature cells, progenitor cells, stem cells – to specifically direct neotissue formation, physical methods, such as transfection by electroporation, rather than viral vectors, can induce highly effective results⁵².

With reference to above nanotechnology innovations, it has been proven that bladder smooth muscle cell adhesion to *nanostructured polymeric surfaces* is significantly improved compared with that to conventional polymeric materials^{53,54}. Moreover, in animal models, nanostructured polymeric surfaces of bladder wall engineered tissue prove to be refractory to calcium incrustation and calcium stone formations⁵⁵. What's more, in a bladder cancer animal research study, specifically regarding the post-total cystectomy replacement bladder tissue engineering, the dispersion of carbon nanofibers within a polyurethane elastomer-based scaffold can inhibit the tumoral relapse into the bladder prosthetic construct⁵⁶.

Just recently, great promises of tissue engineering strategies, for bladder tumor-related surgical organ substitution, have been highlighted, although underlining a number of still unsolved problems, such as particularly the difficulty in obtaining, from different sources, a population of effective smooth muscle cells that may be functionally assimilated to those of native bladder^{57,58}.

Further advances to build, by tissue engineering, a neobladder including the trigone-vesical neck, such that might be implanted, after radical cystectomy, in bladder cancer patients, could be likely achieved by resorting to a decellularized donor whole bladder biomatrix – retaining distal ureters and vesical neck together with preserving wall vascular network – then repopulating it with various source-derived autologous cells, to reach a fully functional organ substitute^{30,38,40,62}, particularly with regard to the attainment of a properly contracting so-made smooth muscle layer.

Current Research Focus and Future Prospects

For patients suffering from seriously injured or end-stage diseased organs, it is often necessary to

resort to whole organ transplant. Because of critical shortage of suitable donor organs, the tissue engineering technologies represent intriguing strategies to build biological replacement prostheses, that morpho-functionally can mimic native organs^{3-6,10-13,21-23,28-30,38}.

In the field of organ transplant surgery, clinically validated current tissue engineering approaches essentially consist in two different procedures, one by resorting to a *scaffold composed of naturally occurring extracellular matrix*, obtained by decellularizing an allogeneic (or, in prospect, xenogeneic) whole organ, otherwise retaining intact intrinsic vascular network and appropriately repopulated with autologous stem cells/differentiated cells, the other, instead, by using a synthetic polymeric biomaterial (PGA, PLA, co-PGA/PLA)- or, sometimes, a biomatrix/synthetic polymer composite-based scaffold, seeded with autologous organ-specific differentiated cells/stem cells. The former has been mainly used to achieve replacement tissue engineered airways for patients suffering from tracheal/bronchial different diseases, among whose particularly the malignant ones^{3-6,10-13,59}, while the latter, so far, to accomplish an augmentation cystoplasty in subjects with end-stage neuropathic poorly compliant bladder, thus obtaining an adequate bladder low-pressure capacity to avoid renal damages^{21-23,28-30,38}, nevertheless without any clinical application of wholly tissue engineered neobladder in bladder tumor radical surgery, hence no relevant literature report, unlike what pertaining to airway replacement in trachea tumor patients. The autologous neourinary conduits, that have been also taken into our consideration more than ten years ago⁶⁶ and whose clinical trials are today in course (ClinicalTrials.gov.identifier:NCT01087697), though potentially directed to eliminate the intestinal neobladder-related complications^{15,67} meanwhile simplifying the surgical procedure compared with Bricker's operation, are far from optimizing – as urinary diversion modalities needing an external urinary reservoir impairing the quality of life – the aims of tissue engineering, that should consist of building an orthotopic bladder replacement anastomosed to the urethra or, at least, a continent cutaneous urinary diversion (pouch)⁶².

All the more so, tissue decellularization technology by detergent-enzymatic treatment, as providing a natural ECM-based scaffold retaining own vascular network and native structural cues, besides its use for hollow organ engineering, could be also applied to engineered transplanta-

tion-directed whole solid organs (liver, kidney, lung) for the radical tumor surgery (and just in case of end-stage organ failure⁶⁰. Apart from some above-signified issues¹⁰, the decellularization technology, applied to trachea tissue engineering, allows the complete removal of native organ cellularity/antigenity meanwhile, according to recent studies⁶¹, preserving its histoarchitecture, though with significant loss of glycosaminoglycan, and an adequate mechanical strength with good cell-repopulating compatibility from the decellularized matrix^{11,61}.

As future prospect in the field of surgical oncology, the goal of bladder tissue engineering will be reached by the construction of the whole artificial neobladder, provided with trigone/vesical neck and distal ureters, such that might serve as wholly replacement bio-prosthesis for bladder tumor patients undergone a radical cystectomy⁶²⁻⁶⁵. So-made bioengineered neobladder could efficaciously avoid – as it as been above-mentioned about the tissue engineering main procedure to build the bladder – both metabolic and malignant complications of the intestinal neobladder^{14,15,66,67}. Six years ago, indeed, a careful analysis on perspective feasible modality to replace bladder, after its malignancy-related total removal, just identified it with a tissue engineering-made neobladder⁶⁸.

Facing the future, further discoveries in the field of nanotechnologies, particularly as far as nanostructured polymer biomaterial scaffold surfaces so that better mimic cell/scaffold interactions at the nanoscale topography of the native tissues – thus offering the advantage of «directly speaking the language of cells»⁶⁹ – will allow tissue engineering technological developments for organ replacement applications in radical tumor surgery^{38,40,44,46,47,50,51,53,54,56,60,70-72}.

References

- 1) PEYTON CH, BURMEISTER D, PETERSON B, ANDERSSON K-E, CHRIST G. Characterization of early proliferative response of rodent bladder to subtotal cystectomy: a unique model of mammalian organ regeneration. *PLoS One* 2012; 7: e47414.
- 2) GRILLO HC. Tracheal replacement: a critical review. *Ann Thorac Surg* 2002; 73: 1995-2004.
- 3) BADYLAC S, WEISS D, CAPLAN A, MACCHIARINI P. Engineered whole organs and complex tissues. *Lancet* 2012; 379: 943-952.
- 4) MACCHIARINI P, JUNGEBLUTH P, GO T, ASNAGHI MA, REES LE, COGAN TA, DODSON A, MARTORELL J, BELLINI S, PARNIGOTTI PP, DICKINSON SC, HOLLANDER AF, MANTERO S, CONCONI MT, BIRCHALL MA. Clinical transplantation of a tissue engineered airway. *Lancet* 2008; 372: 2023-2030.
- 5) LAURENCE J. British boy receives tracheal transplant with his own cells. *BMJ* 2010; 340: c1633.
- 6) CURCIO E, MACCHIARINI P, DE BARTOLO L. Oxygen mass transfer in a human tissue-engineered trachea. *Biomaterials* 2010; 31: 5131-5136.
- 7) BAIGUERA S, DEL GAUDIO C, JANS M, POLIZZI L, GONFIOTTI A, COMIN CE, BIANCO A, RIBATTI D, TAYLOR DA, MACCHIARINI P. Long-term changes to in vitro preserved bioengineered human trachea and their implications for decellularized tissues. *Biomaterials* 2012; 33: 3662-3672.
- 8) PARTINGTON L, MORDAN NJ, MASON C, KNOWLES JC, KIM HW, LOWDELL MW, BIRCHALL MA, WALL I. Biochemical changes caused by decellularization may compromise mechanical integrity of tracheal scaffolds. *Acta Biomater* 2013; 9: 5251-5261.
- 9) BAIGUERA S, JUNGEBLUTH P, BURNS A, MAVILLA C, HAAG J, DE COPPI P, MACCHIARINI P. *Biomaterials* 2010; 31: 8931-8938.
- 10) JUNGEBLUTH P, ALICI E, BAIGUERA S, LE BLANC K, BLOMBERG P, BOZÓKY B, CROWLEY C, EINARSSON O, GRINNEMO KH, GUDBJARTSSON T, LE GUYADER S, HENRIKSSON G, HERMANSON O, JUTO JE, LEIDNER B, LILJA T, LISKA J, LUEDDE T, LUNDIN V, MOLL G, NILSSON B, ROBERBURG C, STRÖMBLAD S, SÜTLU T, TEIXEIRA AI, WATZ E, SEIFALIAN A, MACCHIARINI P. Tracheobronchial transplantation with a stem-cell-seeded bioartificial nanocomposite: a proof-of-concept study. *Lancet* 2011; 378: 1997-2004.
- 11) ELLIOT MJ, DE COPPI P, SPEGGIORIN S, ROEBUCK D, BUTLER CR, SAMUEL E, CROWLEY C, MCLAREN C, FIERENS A, VONDREYS D, COCHRANE L, JEPHSON C, JANES S, BEUMONT NJ, COGAN T, BADER A, SEIFALIAN AM, HSUAN JJ, LOWDELL MW, BIRCHALL MA. Stem-cell-based tissue engineered tracheal replacement in a child: a 2-year follow-up study. *Lancet* 2012; 15: 994-1000.
- 12) JUNGEBLUTH P, MOLL G, BAIGUERA S, MACCHIARINI P. Tissue-engineered airway: a regenerative solution. *Clin Pharmacol Ther* 2012; 91: 81-93.
- 13) JUNGEBLUTH P, BADER A, BAIGUERA S, MÖLLER S, JAUS M, LIM ML, FRIED K, KIARTANSDOTTIR KR, GO T, NAVE H, HARRINGER W, LUNDIN V, TEIXEIRA AI, MACCHIARINI P. The concept of in vivo airway tissue engineering. *Biomaterials* 2012; 33: 4319-4326.
- 14) ALBERTI C, SOANA S, ISOLA I. Experimental ileal bladder: urinary release and back-round of radioactive tracers. *Minerva Urol* 1971; 23: 1-3.
- 15) Alberti C. Metabolic and histological complications in ileal urinary diversion. Challenges of tissue engineering technology to avoid them. *Eur Rev Med Pharmacol Sci* 2007; 11: 257-264.
- 16) PIECHOTA HJ, DAHMS SE, NUNES LS, DAHYA R, LUE TF, TANAGHO EA. *In vitro* functional properties of the regenerated rat bladder by the bladder acellular matrix graft. *J Urol* 1998; 159: 1717-1726.
- 17) ZHANG Y, FRIMBERGER D, CHENG EY, LIN H-K, KROPP BP. Challenges in a larger bladder replacement

- with cell-seeded and un-seeded small intestinal submucosa grafts in subtotal cystectomy model. *BJU Int* 2006; 88: 1100-1105.
- 18) ZHOU L, YABG B, SUN C, QIU X, SUN ZY, CHEN Y, ZHANG Y, DAI YT. Co-administration of PDGF-BB and VEGF with BAM enhance smooth muscle regeneration and vascularization for bladder augmentation in a rabbit model. *Tissue Eng (Part A)* 2013; 19: 264-276.
 - 19) MITSUI Y, SHIINA H, HIRAKA T, ARICHI N, YASUMOTO H, DAHIRA R, TANAGHO EA, IGAWA M. Simultaneous implantation of bilateral ureters into BAM graft after partial cystectomy in a porcine model. *BJU Int* 2012; 110: E1212-E1217.
 - 20) DAVIS NF, CALLANAN A, MCGUIRE BB, FLOOD HD, MCGLOUGHLIN TM. Evaluation of viability and proliferative activity of human urothelial cells cultured onto xenogenic tissue-engineered extracellular matrices. *Urology* 2011; 77: e1-7.
 - 21) ATALA A. Tissue engineering in the genitourinary system. In: Atala A, Mooney D, eds. *Tissue engineering*. Boston: Birkhauser Press 1997, p. 149.
 - 22) OPERPENNING F, MENG J, YOO JJ, ATALA A. De novo reconstitution of a functional mammalian urinary bladder by tissue engineering. *Nat Biotechnol* 1999; 17: 146-155.
 - 23) ATALA A. Recent applications of regenerative medicine to urologic structures and related tissues. *Curr Opin Urol* 2006; 16: 305-309.
 - 24) OKITA K, ICHISAKA T, YAMANAKA S. Generation of germline-competent induced pluripotent stem cells. *Nature* 2007; 448: 313-317.
 - 25) LI W, ZHAO X-Y, WAM H-F, ZHANG Y, LIU L, LV Z, WANG X-J, WANG L, ZHOU Q. iPS cells generated without c-Myc have active DiK1-Dio3 region and are capable of producing full-term mice through tetraploid complementation. *Cell Res* 2011; 21: 550-553.
 - 26) MAIOLI M, RINALDI S, SANTANIELLO S, CASTAGNA A, PIGLIARU G, GUALINI S, CAVALLINI C, FONTANI V, VENTURA C. Radio-electric conveyed fields reprogram human dermal-skin fibroblasts towards cardiac-, neuronal-, skeletal muscle-like lineages. *Cell Transplant* 2012 (in press).
 - 27) FERGUSON DR. Urothelial function. *Br J Urol* 1999; 84: 235-242.
 - 28) ATALA A. Future perspectives in reconstructive surgery using engineering. *Urol Clin North Am* 1999; 26: 157-165.
 - 29) ATALA A, KASPER FK, MICOS AG. Engineering complex tissues. *Sci Transl Med* 2012; 4: 160rv12.
 - 30) ZHANG Y, ATALA A. Urothelial cell culture: stratified urothelial sheet and 3D-growth or urothelial structure. *Methods Mol Biol* 2013; 945: 383-399.
 - 31) CAIONE P, BOLDRINI R, SALERNO A, NAPPO SG. Bladder augmentation using acellular collagen biomatrix: a pilot experience in exstrophic patients. *Pediatr Surg Int* 2012; 28: 421-428.
 - 32) ATALA A, BAUER SB, SOKER S, YOO JJ, RETIK AB. Tissue-engineered autologous bladders for patients needing cystoplasty. *Lancet* 2006; 367: 1241-1246.
 - 33) YOO JJ, OLSON J, ATALA A, KIM B. Regenerative medicine strategies for treating neurogenic bladder. *Int Neurourol J* 2011; 15: 109-119.
 - 34) CROSS WR, THOMAS DFM, SOUTHGATE J. Tissue engineering and stem cell research in urology. *BJU Int* 2003; 92: 165-171.
 - 35) EBERLY D, ATALA A. Tissue engineering using adult stem cells. *Methods Enzymol* 2006; 420: 287-302.
 - 36) AMABILE G, MEISSNER A. Induced pluripotent stem cells: current progress and potential for regenerative medicine. *Trends Molecular Med* 2009; 15: 59-68.
 - 37) ALBERTI C. Tissue engineering technologies: just a quick note about transplantation of bioengineered donor trachea and augmentation cystoplasty by de novo engineered bladder tissue. *G Chir* 2009; 30: 514-519.
 - 38) MURPHY SV, ATALA A. Organ engineering, combining stem cells, biomaterials and bioreactors to produce bioengineered organs for transplantation. *Bioessays* 2013; 35: 163-172.
 - 39) CHUN SY, KIM HT, LEE JS, KIM MJ, KIM BS, KIM BW, KNOW TG. Characterization of urine-derived cells from upper urinary tract in patients with bladder cancer. *Urology* 2012; 79: 1186.e 1-7.
 - 40) FURTH ME, ATALA A, VAN DYKE ME. Smart materials design for tissue engineering and regenerative medicine. *Biomaterials* 2007; 28: 5068-5073.
 - 41) LANGER R. Perspectives and challenges in tissue engineering and regenerative medicine. *Adv Mater* 2009; 21: 3235-3236.
 - 42) KO IK, JU YM, CHEN T, ATALA A, YOO JJ, LEE SJ. Combined systemic and local delivery of stem cell inducing/recruiting factors for in situ tissue regeneration. *FASEB* 2012; 26: 158-168.
 - 43) HERSEL V, DALMEN C, KESSLER H. RDG-modified polymers: biomaterials for stimulated cell adhesion and beyond. *Biomaterials* 2003; 24: 4385-4415.
 - 44) SHASTRI VP. *In vivo* engineering of tissues: biological considerations, challenges, strategies and future directions. *Adv Mater* 2009; 21: 3246-3254.
 - 45) KHADEMHOSEINI A, LANGER R. Microengineered hydrogels for tissue engineering. *Biomaterials* 2007; 28: 5087-5092.
 - 46) YANG Y, LEONG RW. Nanoscale surfacing for regenerative medicine. *Nanomed nanobiotechnol* 2010; 2: 478-495.
 - 47) JOHNSON TD, LIN SY, CHRISTMAN KL. Tailoring material properties of a nanofibrous ECM-derived hydrogel. *Nanotechnology* 2011; 22: 494-515.
 - 48) NOLAN K, MILLET Y, RICORDI C, STABLER CL. Tissue Engineering and biomaterials in regenerative medicine. *Cell Transplant* 2008; 17: 241-243.
 - 49) ALBERTI C. Tissue engineering: technological advances to improve its applications in reconstructive surgery. *G Chir* 2012; 33: 435-443.
 - 50) MURPHY SV, SHARDAL A, ATALA A. Evaluation of hydrogels for bio-printing applications. *J Biomed Mater Res A* 2013; 101: 272-287.

- 51) DEMMING A. The nature of progress in nanopatterning. *Nanotechnology* 2012; 23: 460201.
- 52) MELLOTT AJ, FORREST ML, DETAMORE MS. Physical nonviral gene delivery methods for tissue engineering. *Ann Biomed Eng* 2012 (in press).
- 53) THAPA A, MILLER DC, WEBSTER TJ, HABERSTROH KM. Nanostructured polymers enhance smooth muscle cell function. *Biomaterials* 2003; 24: 2915-2926.
- 54) ROTH CC. Urologic tissue engineering in pediatrics: from nanostructures to bladder. *Pediatrics Res* 2010; 67: 509-513.
- 55) SHARMA AK. An examination of regenerative medicine-based strategies for the urinary bladder. *Regen Med* 2011; 6: 583-598.
- 56) TSANG M, CHUN YW, IM YM, KHANG D, WEBSTER TJ. Effects of increasing carbon nanofiber density in polyurethane composites for inhibiting bladder cancer cell function. *Tissue Eng (Part A)* 2011; 13/14: 1879-1889.
- 57) DREWA T, ADAMOWICZ J, SHARMA A. Tissue engineering for the oncologic urinary bladder. *Nat Rev Urol* 2012; 9: 561-572.
- 58) ADAMOWICZ J, JUSZCZAK K, BAJEK A, TWORKIEWICZ J, NOWACKI M, MARSZALEK A, THOR PJ, CHLOSTA P, DREWA T. Morphological and urodynamic evaluation of urinary bladder wall regeneration: muscles guarantee contraction but not proper function in a rat model research study. *Transplant Proc* 2012; 44: 1429-1434.
- 59) RICH JT, GULLAME PJ. Current concepts in tracheal reconstruction. *Curr Opin Otolaryngol Head Neck Surg* 2012; 20: 246-253.
- 60) YAGI H, SOTO-GUTIERREZ A, KITAGAWA Y. Whole organ re-engineering: a regenerative medicine approach in digestive surgery for organ replacement. *Surg Today* 2012 (in press).
- 61) ZANG M, ZHANG Q, CHANG EI, MATHUR AB, YU P. Decellularized tracheal matrix scaffold for tissue engineering. *Plast Reconstr Surg* 2012; 130: 532-540.
- 62) ALBERTI C. Bladder regeneration by tissue engineering (A. Atala's reply). *BJU Int* 2002; 89: 972-973.
- 63) ALBERTI C. Hollow organ tissue engineering: short updating about current approaches and forecast for major research advances. *G Chir* 2011; 32: 345-351.
- 64) YANNAS IV. Emerging rules for inducing organ regeneration. *Biomaterials* 2013; 34: 321-330.
- 65) HE M, CALLANAN A. Comparison of methods for whole organ decellularization in tissue engineering of bioartificial organs. *Tissue Eng Part B Rev* 2012 (in press).
- 66) ALBERTI C, TIZZANI A. Biomaterials in bladder replacement: purposes and prospects. *Urologia* 2002; 69: 296-299.
- 67) HYNDMAN ME, KAYE NC, FIELD NC, LAWSON KA, SMITH ND, STEINBERG GD, SCHOENBERG MP, BIVILACQUA TJ. The use of regenerative medicine in the management of invasive bladder cancer. *Adv Urol* 2012; 2012: 653652.
- 68) McATEER H, COSH E, FREEMAN G, PANDIT A, WOOD P, LIFFORD R. Cost-effectiveness analysis at the development phase of a potential health technology: examples based on tissue engineering of bladder and urethra. *J Tissue Eng Regen Med* 2007; 1: 343-349.
- 69) HARRINGTON DA, SHARMA AK, ERIKSON BA, CHENG EY. Bladder tissue engineering through nanotechnology. *World J Urol* 2008; 26: 315-322.
- 70) PATTISON M, WEBSTER TJ, LESLIE J, KAEFER M, HABERSTROH KM. Evaluating the in vitro and in vivo efficacy of nanostructured polymers for bladder replacement applications. *Macromol Biosci* 2007; 7: 690-700.
- 71) ILIE I, ILIE R, MOCAN T, BARTOS D, MOCAN L. Influence of nanomaterials on stem cell differentiation: designing an appropriate nanobiointerface. *Int J Nanomedicine* 2012; 7: 2211-2225.
- 72) ALBERTI C. Outlines on nanotechnologies applied to bladder tissue engineering. *G Chir* 2012; 33: 234-238.