

Chi311 regulates APAP-induced liver injury by promoting macrophage infiltration

Y. WANG¹, M. ZHONG², W. WANG³, Y.-H. LI¹

¹Department of General Surgery, China-Japan Union Hospital of Jilin University, Changchun, China

²Department of Respiratory, China-Japan Union Hospital of Jilin University, Changchun, China

³Department of Breast Surgery, Jilin Province Tumor Hospital, Changchun, China

Abstract. – **OBJECTIVE:** This study aims to investigate the role of Chi311 in Acetaminophen (APAP)-induced liver injury.

MATERIALS AND METHODS: *In vivo* model of liver injury was established in mice administered with APAP (250 mg/kg) or equivalent phosphate-buffered saline (PBS). Mouse liver tissues were collected at 1 h, 3 h, 6 h, 12 h, and 24 h after treatment, respectively. ALT levels and apoptosis were evaluated. Additionally, we established APAP-induced acute liver injury model in wild-type (WT) mice and Chi311-deficient (Chi311^{-/-}) mice. Pathological changes of liver tissue were observed by hematoxylin and eosin (HE) staining. Mononuclear cells (MNCs) were isolated from mouse liver, and amounts of infiltrating macrophages and neutrophils were then counted by flow cytometry. Serum levels of cytokines were detected by enzyme-linked immunosorbent assay (ELISA). Bone marrow-derived macrophages (BMDMs) were extracted from each mouse.

RESULTS: After APAP treatment, Chi311^{-/-} mice showed more severe liver injury than that of WT mice, which was manifested as higher ALT levels and more necrotic and apoptotic cells. Compared with WT mice, Chi311^{-/-} mice expressed higher levels of inflammatory cytokines (MCP-1 and IL-6), macrophage-associated molecules (CD68 and CD86), as well as the amounts of infiltrating macrophages and neutrophils. In addition, higher expression of inflammatory cytokines were found in BMDMs extracted from WT mice treated with those APAP compared with those APAP-treated cells. APAP-treated Chi311^{-/-} mice showed more severe liver injury than that of WT mice.

CONCLUSIONS: Our study confirmed that Chi311 protects the liver function from APAP-induced liver injury by inhibiting the secretion of inflammatory factors and macrophage infiltration.

Keywords:

Acetaminophen, Drug-induced liver injury, Chi311.

Introduction

Drug-induced liver injury (DILI), as a common side effect of drugs, is one of the leading causes of acute hepatic failure and liver transplantation in Western countries¹. DILI is the major reason for the failure of many new drugs development, which is also being concerned by the regulatory department^{2,3}. Acetaminophen (APAP) is an acetanilide antipyretic analgesic, which is one of the most popular non-prescription drugs in the pharmaceutical market due to its safety and reliable dose. In China, APAP has been used as an adjunct to acute and chronic hepatitis for several decades⁴. Overdose of APAP is one of the leading causes of DILI in the world⁵, accounting for more than 50% of reported DILI. Severities of symptoms caused by APAP are varied because of individual differences, which makes it difficult to determine its pathogenesis⁶. After a small amount of APAP enters the liver, it produces biologically active N-acetyl-to-benzenequinoneimine (NAPQI), which further exerts its detoxification function by combining with reduced glutathione (GSH). However, overdose of APAP will cause NAPQI accumulation in the liver and excessive consumption of GSH⁷. Glutathione peroxidase is the main inactivating enzyme of peroxides. GSH deficiency significantly inhibits glutathione peroxidase, thereby leading to accumulation of peroxides in the body⁸. It has been reported that macrophages and their related inflammatory responses exert an important role in APAP-induced liver damage, while their roles in promoting the occurrence and progression of DILI require further explorations⁹. Chi311 (chitinase 3 like 1) is a chitinase-like soluble secretory protein without any enzymatic activity¹⁰. It can be produced by a variety of cells, including neutro-

phils, macrophages, chondrocytes, synoviocytes, smooth muscle cells, endothelial cells and tumor cells^{10,11}. Accumulating evidences have suggested that elevated level of Chi311 is associated with poor prognosis of liver diseases, such as liver fibrosis, non-alcoholic fatty liver, alcoholic liver disease and hepatocellular carcinoma^{12,13}. We speculated that Chi311 exerts a crucial role in promoting macrophage infiltration and secretion of inflammatory factors, thereby aggravating liver damage. In this study, we first confirmed that Chi311 participated in the development of APAP-induced DILI by detecting the mRNA expression of Chi311. Second, we verified that APAP could induce hepatocyte apoptosis, macrophage infiltration and inflammatory response. Therefore, our results suggested that Chi311 participated in APAP-induced liver injury and might be served as a potential therapeutic target for drug-induced hepatitis.

Materials and Methods

Animal Rearing

Chi311^{-/-} mice used in this study were imported from the Jackson Laboratory in the United States. 8-week old wild-type (WT) and Chi311^{-/-} mice were fed in our SPF (specific-pathogen-free) Experimental Animal Center. All mice were supplied with free access to food and drinking water and kept in a single cage. This study was approved by the Institutional Ethics Committee of Jilin University Animal Center.

Animal Model Establishment

WT and Chi311^{-/-} mice were intraperitoneally injected with 250 mg/kg APAP to induce acute liver injury. Mice tissues were collected after administration for 8 h and 24 h, respectively.

Biochemical and Pathological Analysis

Serum samples collected from WT and Chi311^{-/-} mice treated with APAP for 8 h and 24 h were used for detecting activities of ALT and AST. GSH level in liver tissues was measured according to the instructions of GSH assay kit (Jiancheng, Nanjing, China). Liver tissues were observed under a microscope after hematoxylin and eosin (HE) staining.

Real-Time Fluorescence Quantitative Polymerase Chain Reaction (qRT-PCR)

Total RNA was extracted from liver tissues by TRIzol method (Invitrogen, Carlsbad, CA, USA) and reversely transcribed into complementary Deoxyribonucleic Acid (cDNA) as previously described. RNA concentration was detected using a spectrometer and those samples with A260/A280 ratio of 1.8-2.0 were selected for the following qRT-PCR reaction. QRT-PCR was then performed based on the instructions of SYBR Premix Ex Taq II-M (TaKaRa, Otsu, Shiga, Japan). The experiment was repeated 3 times in parallel. QRT-PCR reaction parameters were as follows: denaturalization at 95°C for 60 s, extension at 55°C for 30 s and annealing at 60°C for 40 s, for a total of 40 cycles. Primer sequences were shown in Table I.

Extraction of Liver Mononuclear Cells (MNCs)

Liver tissue of mouse was completely digested with collagenase and then ground to prepare for the single cell suspension. After centrifugation at a gradient density, the red blood cell lysate was used to lyse the pellet. The lysate was washed with the buffer containing 0.5% fetal bovine serum (FBS) (Gibco, Rockville, MD, USA), and finally the cells were resuspended using 100 µL of 2% fetal bovine serum (FBS) containing buffer.

Table I. qRT-PCR primer sequences

	Forward	Reverse
mGSH	AGGTCGGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA
mAPAP	TGGACCTTCCAGGATGAGGACA	GTTCATCTCGGAGCCTGTAGTG
mTNF-α	TACCACTTCAAGTCGGAGGC	CTGCAAGTGCATCATCGTTGTTC
mChi311	GGTGCCATGTCTCAGCCTCTT	GCCATAGAAGTATGAGAGGGAG
	TCCTTACGGAGAGACACCT	GGCTGGGAACCATTAGTC
β-actin	GGGGATCCATGGGCAATCCTTAT	TCGGGTGACCTTGCTTAGACGTGCAGG

Flow Cytometry

Fc receptor blocker and anti-CD16/3 antibody were added to the MNCs cell suspension and incubated for 10 min to block non-specific binding sites. Subsequently, the corresponding flow cytometry (Partec AG, Arlesheim, Switzerland) antibody was incubated in dark for 15 min. Next, 1 mL of 2% FBS was used to wash the cells. After centrifugation at 400 g for 5 min, 400 μ L of 2% FBS were added for resuspension. The antibodies used in this assay were as follows: PE-Cyanine7-labeled anti-mouse CD11b, APC-labeled anti-mouse F4/80, fluorescein isothiocyanate (FITC)-labeled anti-mouse Ly6C, PE-labeled anti-mouse Ly6G, APC Vio770-tag anti-mouse CD45. All antibodies were purchased from eBioscience (San Diego, CA, USA).

Determination of Serum Cytokine Levels

After APAP treatment in mice for 8 h and 24 h, serum samples were collected and subjected to experimental procedures according to the enzyme-linked immunosorbent assay (ELISA) kit (Bio Legend, San Diego, CA, USA). The optical density (OD) values of each well at the wavelength of 562 nm and 450 nm were measured with a microplate reader (Bio-Rad, Hercules, CA, USA). Excel was used to draw a standard curve of cytokine concentration and absorbance value.

Cell Extraction and Culture

After APAP injection, WT mice were sacrificed, the muscle tissue was blunted, and the femur and tibia tissue were dissected and soaked in sterile phosphate buffered saline (PBS). Bone marrow cells were flushed and cultured in RPMI-1640 (Rochester Park Memorial Institute-1640) medium (HyClone, South Logan, UT, USA) containing 10 ng/mL M-CSF (macrophage colony-stimulating factor). After culturing for 6 days, liver lysate derived from Chi311^{-/-} mouse (1:2) was added to BMDMs extracted from WT mice and incubated for 17 h. Expression levels of inflammatory factors in BMDMs extracted from mice were examined. Cells in the control group did not receive any treatment. The liver tissue of Chi311^{-/-} mice received APAP treatment was homogenized in PBS. After frozen in liquid nitrogen, liver tissue was subjected to a 37°C water bath. The liver lysate was then washed with a 70- μ m sieve and the supernatant was harvested.

Statistical Analysis

All experiments were repeated at least three times. Data were expressed as mean \pm SD (Standard Deviation). The independent-sample *t*-test was used to compare the intergroup differences. Multiple-group comparisons were performed using one-way analysis of variance (ANOVA) followed by Post-Hoc Test (Least Significant Difference) or Bonferroni was performed to detect significant differences between groups. GraphPad Prism 6.0 (La Jolla, CA, USA) was used for statistical analysis. $p < 0.05$ was considered statistically significant.

Results

Chi311 Was Involved in the Pathological Process of APAP-Induced DILI

To investigate whether Chi311 participates in the pathogenesis of APAP-related drug-induced liver injury, we randomly assigned WT mice into two groups. Mice received intraperitoneal injections of APAP or PBS, respectively. After injection for 1 h, 3 h, 6 h, 12 h, and 24 h, respectively, mice were sacrificed by carbon dioxide asphyxiation for collecting liver tissues. We detected the mRNA level of Chi311 by qRT-PCR (Figure 1). The results showed that compared with the PBS group, mRNA level of Chi311 remarkably increased after injection of APAP in a time-dependent manner, which lasted for 24 h. These results suggested that Chi311 might be involved in the pathogenesis of APAP-induced liver injury.

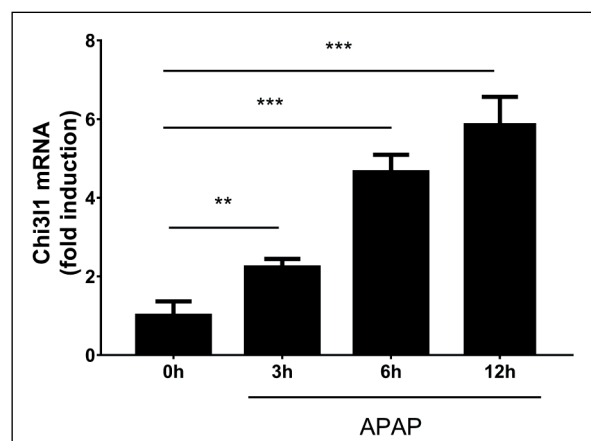


Figure 1. Chi311 was involved in the pathological process of APAP-mediated drug-induced liver injury. The mRNA expression of Chi311 in the liver of mice was detected by qRT-PCR after PBS or APAP injection for 1 h, 3 h, 6 h, 12 h and 24 h (** $p < 0.01$).

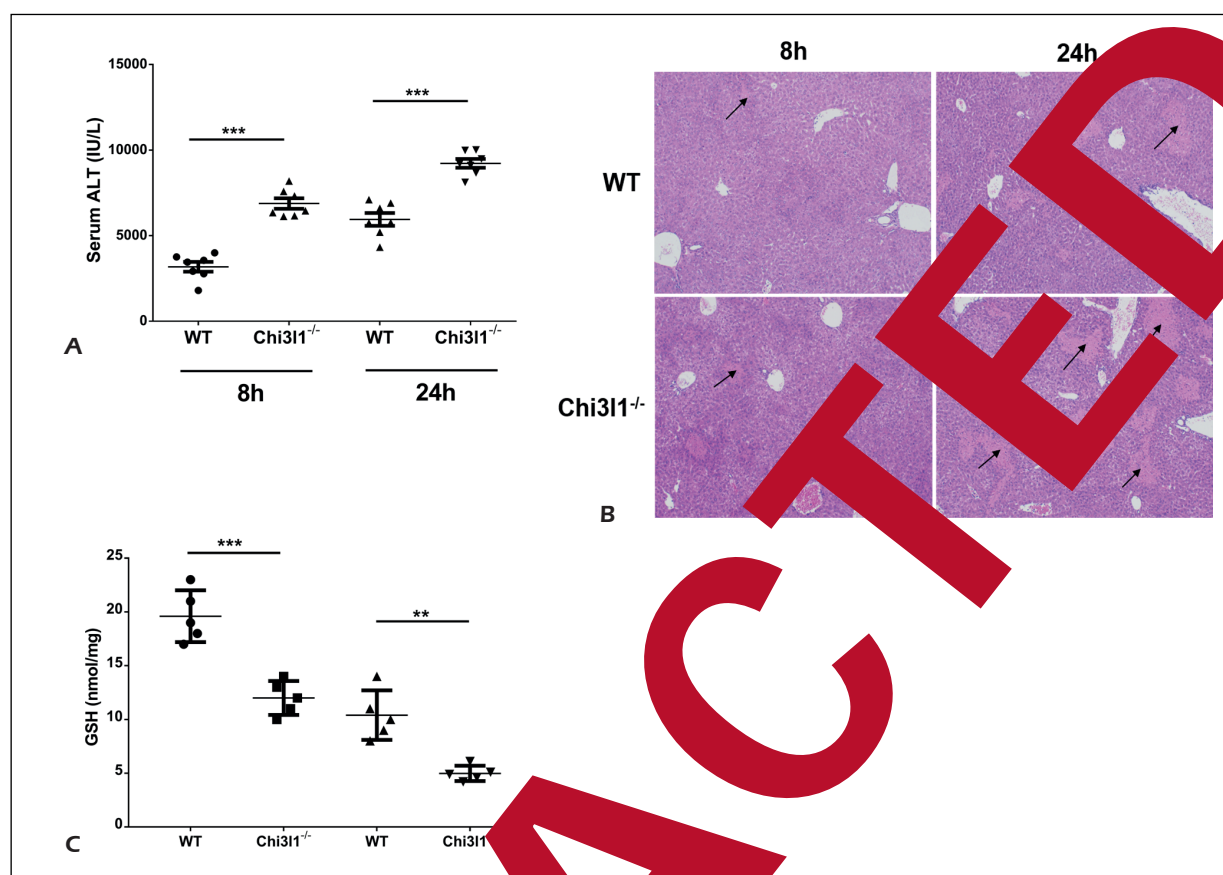


Figure 2. Chi311 knockout exacerbated APAP-induced liver injury in mice. **A**, Serum ALT levels in WT and Chi311^{-/-} mice at 8 and 24 h after APAP injection. **B**, HE staining micrographs (magnification 100 \times , 200 \times) of liver tissues of WT and Chi311^{-/-} mice at 8 h and 24 h; significant hepatocyte necrosis was observed in Chi311^{-/-} mice (black arrows). **C**, GSH levels in livers of WT and Chi311^{-/-} mice 24 h after APAP injection; GSH levels were significantly lower in APAP group than those of the PBS group, and GSH levels in the Chi311^{-/-} group were significantly lower than that of the WT group (** $p < 0.01$).

Knockout of Chi311 Exacerbated APAP-Induced Liver Injury in Mice

To study the role of Chi311 in APAP-mediated drug-induced liver injury, we intraperitoneally injected APAP into WT and Chi311^{-/-} mice. Mice blood and liver tissues were collected after injection at 8 h and 24 h, respectively. Serum levels of ALT was measured after centrifugation and the results showed that the ALT levels in Chi311^{-/-} mice were remarkably higher than those of WT mice (Figure 2A). Pathological results of liver tissue sections revealed that Chi311^{-/-} mice were more sensitive to APAP-induced liver injury since they exhibited extensive histiocytic necrosis (Figure 2B). GSH levels in the liver tissues of Chi311^{-/-} mice were also markedly higher than those of WT mice (Figure 2C). The above data indicated that knockout of Chi311 exacerbated liver damage in mice after APAP treatment.

Knockout of Chi311 Aggravated the Inflammatory Response in Liver

After clarifying that Chi311 negatively regulated APAP-induced liver injury, we further explored its mechanisms by examining expressions of inflammatory cytokines in the liver after APAP injection. The mRNA levels of IL-6 and MCP-1 in liver tissues of APAP-treated Chi311^{-/-} mice were remarkably higher than those of WT mice. However, no significant differences in mRNA levels of IL-1 β , TNF- α , and IFN- γ were found between Chi311^{-/-} mice and WT mice (Figure 3A). Protein levels of IL-6 and MCP-1 in Chi311^{-/-} mice were also remarkably higher than those of WT mice (Figure 3B). All these results demonstrated that the Chi311 exacerbated APAP-induced liver injury by promoting the secretion of inflammatory factors.

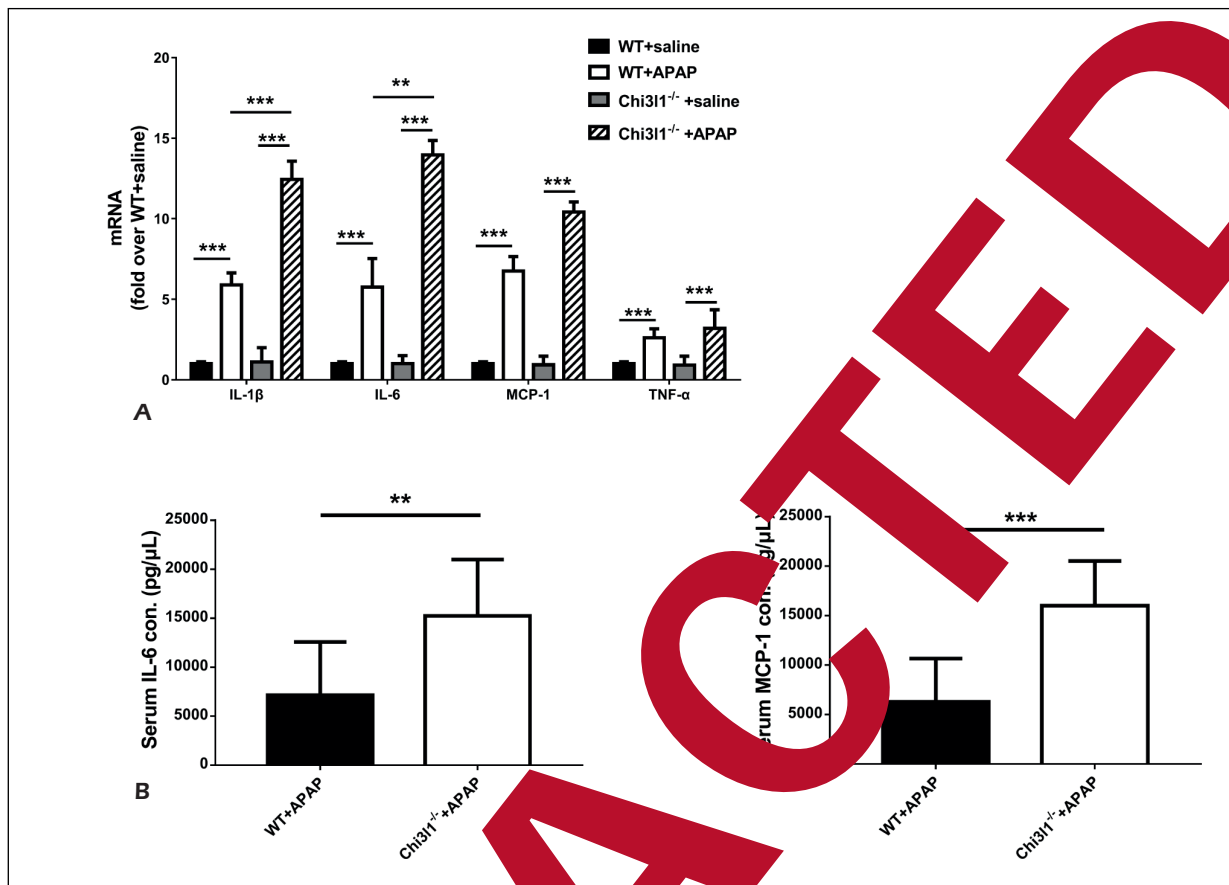


Figure 3. Knockout of Chi311 aggravated the inflammatory response in liver. **A**, The mRNA levels of various inflammatory cytokines in the liver of WT and Chi311^{-/-} mice were measured by qRT-PCR after 24 h of APAP treatment. **B**, Serum IL-6 and MCP-1 levels were measured by ELISA. The expression levels of inflammatory cytokines in tissues and serum of mice treated with APAP were much higher than those in the control group. The expression levels of IL-6 and MCP-1 in Chi311^{-/-} group were much higher than those in the WT group (* $p < 0.05$, ** $p < 0.01$).

Chi311 Regulates the Apoptosis of Hepatocytes and Secretion of Pro-Inflammatory Cytokines Via Macrophage Infiltration

It is reported that Chi311 is involved in the inflammatory reaction of adipose tissues and macrophages. Hence, we investigated whether Chi311 could regulate liver damage and inflammatory responses through macrophages in APAP-induced liver injury. Our results demonstrated that the macrophage marker CD68 and pro-inflammatory macrophage marker CD86 were remarkably higher in liver tissues of Chi311^{-/-} than those of WT mice (Figure 4A). In addition, we examined the macrophage infiltration in liver tissues. The data showed that after APAP treatment for 24 h, the total amounts of infiltrating macrophages (CD11b⁺Ly6C⁺)

and neutrophils (CD11b⁺Ly6G⁺) in the liver of Chi311^{-/-} mice were higher than those of WT mice (Figure 4B). Subsequently, we further observed the role of inflammatory macrophages in liver tissues of APAP-induced inflammation *in vitro*. BMDMs in WT and Chi311^{-/-} mice were extracted, respectively. Apoptosis results elucidated that positive staining of Annexin V and 7-AAD in macrophages extracted from Chi311^{-/-} mice was much more pronounced than those of WT mice (Figure 4C). Besides, levels of IL-6 and MCP-1 in the BMDMs incubated with lysates extracted from Chi311^{-/-} mice were higher than those of the untreated group (Figure 4D). All above data confirmed that increased secretion of inflammatory factors by macrophages was the major cause of severe liver damage in Chi311^{-/-} mice.

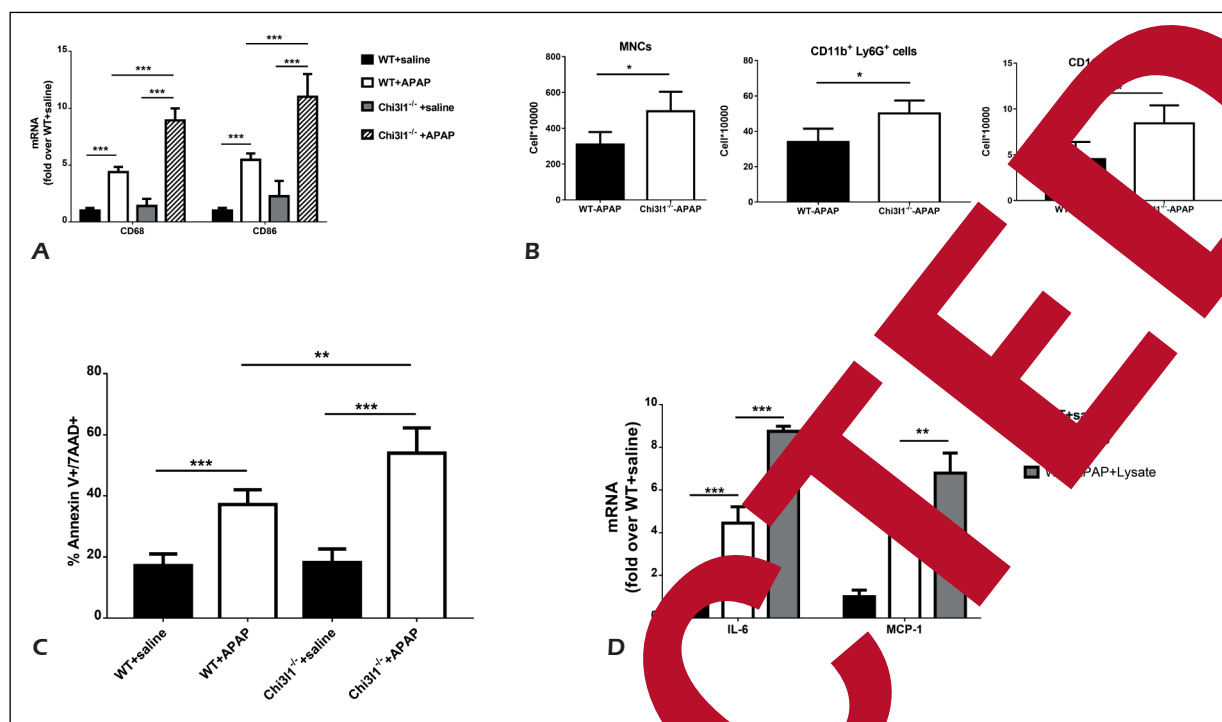


Figure 4. Chi311 regulated the apoptosis of hepatocytes and the secretion of inflammatory cytokines via macrophage infiltration. **A**, After 24 h of APAP treatment, macrophage-related marker CD68 and CD86 mRNA levels in livers of WT and Chi311^{-/-} mice were measured by qRT-PCR. **B**, After 24 h of APAP treatment, liver MNCs were isolated. Significantly increased Ly6G⁺CD11b⁺ macrophages and Ly6G⁻CD11b⁺ neutrophils were observed in the liver of Chi311^{-/-} mice compared to WT mice. **C**, After 24 h of APAP treatment, BMDMs were extracted from WT and Chi311^{-/-} mouse. Positive macrophage was detected by Annexin V and 7-AAD. **D**, After incubation of liver lysates of Chi311^{-/-} mice, expressions of inflammatory factors in BMDMs of wild-type mice were measured. (**p*<0.05, ***p*<0.01, ****p*<0.001).

Discussion

In this study, we demonstrated that APAP treatment induced the secretion of Chi311 in mice, which in turn mediated liver damage by regulating macrophage activation, infiltration and secretion of IL-6 and MCP-1. APAP-induced DILI involves several major steps. Firstly, the drug itself or its metabolites directly damage the hepatocyte stress response (intrinsic pathway) or activate the immune response (extrinsic pathway). APAP also directly leads to mitochondrial dysfunction. Secondly, mitochondrial permeability transition pore (MPT) opens. Thirdly, MPT eventually leads to apoptosis and necrosis of hepatocytes. Previous studies^{4,12,15,16} have reported that Chi311 is involved in the pathogenesis of various liver diseases, especially in immune-mediated liver diseases induced by inflammation and toxins. Our results revealed that IL-6 and MCP-1 levels in liver tissues and serum samples of Chi311^{-/-} mice were remarkably higher than those of wild-type mice. The role of TNF- α in APAP-in-

duced liver injury is controversial. Studies have reported that TNF- α receptor knockout could inhibit APAP hepatotoxicity in mice¹⁷. However, some researchers believed that APAP-induced damage is similar in TNF- α knockout mice and WT mice. In our study, no significant difference in mRNA level of TNF- α was observed between WT and Chi311^{-/-} mice. After initially confirming the involvement of Chi311 in APAP-mediated DILI, we hypothesized that Chi311 could directly induce the secretion of these inflammatory factors. A great number of studies have shown that infiltrating macrophages and neutrophils participate in inducing acute liver inflammation in the mouse hepatitis model¹⁸⁻²¹. Therefore, we also considered the role of macrophage infiltration in regulating hepatic inflammatory cytokine secretion by Chi311. We found that both the infiltration of CD11b⁺Ly6G⁺ macrophages and CD11b⁺Ly6G⁻ neutrophils in the liver of Chi311^{-/-} mice were upregulated, further indicating the key role of Chi311 in the macrophage infiltration of liver. The source of

macrophages in liver inflammation has been well studied. Whether this process is a result induced by liver intrinsic macrophages or bone marrow-derived macrophages deserves further investigation. We extracted bone marrow-derived macrophages to observe the direct effect of hepatic lysates on secretion of inflammatory factors by myeloid-derived macrophages. Our data confirmed that Chi311 was involved in the activation of myeloid macrophages and their subsequent cascade of inflammatory responses. It is of great significance to clarify the pathogenesis of DILI to improve the preventive and therapeutic efficacy of DILI. Current researches have suggested that various aspects are involved in DILI pathogenesis, such as drug metabolism, mitochondrial function impairment, immune response, signal transduction, genetics and environmental factors. Therefore, DILI is a result of the combined effects of multiple factors. Chi311 is greatly involved in the macrophage-mediated hepatic inflammation response in the APAP-mediated drug-induced hepatitis model. Our results demonstrated that the Chi311 might be a potential therapeutic target for acute hepatitis and liver failure.

Conclusions

APAP-treated Chi311^{-/-} mice showed more severe liver injury than that of WT mice. Chi311 protects the liver function from APAP-induced injury by inhibiting the secretion of inflammatory factors and macrophage activation, which provides a new direction for the prevention and treatment of clinical drug-induced hepatitis.

Conflict of Interests

The authors declare no competing interests.

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