

MiR-1269a acts as an onco-miRNA in non-small cell lung cancer via down-regulating *SOX6*

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Abstract. – **OBJECTIVE:** Lung cancer, especially non-small cell lung cancer (NSCLC), remains one of the leading death-causing malignant tumors worldwide. MicroRNAs (miRNAs) have been identified to participate in the development and progression of NSCLC. However, the role of miR-1269a in NSCLC still needs to be elucidated. The objective of this study was to investigate the function of miR-1269a in NSCLC and its underlying mechanism.

PATIENTS AND METHODS: Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR) was utilized to measure the expression level of miR-1269a in NSCLC tissues and cell lines. After transfection with miR-1269a mimics or inhibitors, the expression level of miR-1269a in NSCLC was up- or down-regulated. Cell Counting Kit-8 (CCK-8) assay and colony formation assay were used to measure cell proliferation ability. Flow cytometry assay was applied to verify the cell cycle distributions of established cell lines. The potential target of miR-1269a was determined by using dual-luciferase and Western blot.

RESULTS: miR-1269a was significantly over-expressed in NSCLC tissues than that in adjacent normal tissues. The expression of miR-1269a was also up-regulated in NSCLC cell lines. Up-regulation of miR-1269a inhibited the abilities of cell proliferation, colony formation, and induced cell cycle arrest. Meanwhile, down-regulation of miR-1269a increased the capacities of cell proliferation, colony formation, and arrested the cell cycle. It was further indicated that *SOX6* was verified as a target of miR-1269a in NSCLC and over-expressed *SOX6* could rescue the effect of miR-1269a up-regulation.

CONCLUSIONS: Our study demonstrated that miR-1269a could function as an onco-miRNA in NSCLC and promote NSCLC growth via down-regulating the expression of *SOX6*.

Key words:

miR-1269a, NSCLC, Proliferation, *SOX6*.

Introduction

Non-small cell lung cancer (NSCLC), which accounts for 85% of lung cancer, is one of the

most common malignant tumor. NSCLC is also the most common reason for tumor mortality in the world^{1, 2}. Although the technologies of diagnosis and treatment have improved rapidly, the prognosis of NSCLC is still very unoptimistic³. Therefore, it is important to find more specific and conclusive evidence on NSCLC progression, which may serve as a novel target for the biological treatment of NSCLC.

MicroRNAs (miRNAs) are a subtype of non-coding RNA with 20-22 nucleotides in length. miRNAs are involved in many biological processes of cancers, including NSCLC^{4, 5}. In a general way, miRNAs can bind to the 3'-UTR of target genes and reduce protein expression, eventually influencing the proliferation, metastasis, or oncogenesis of cancers^{6, 7}. For example, miR-216a may suppress the growth of renal cell carcinoma via suppressing TLR4⁸. In colorectal cancer, miR-218 is indicated as a good prognostic ending and may enhance cell apoptosis through targeting BIRC5⁹. Moreover, in lung cancer, miR-182 inhibits EMT and the metastasis of cells by targeting the Met gene, while miR-106b-5p may promote cell growth and inhibit cell apoptosis via regulating BTG3^{10, 11}.

miR-1269a is an onco-miRNA firstly identified in colorectal cancer, which may promote the development and metastasis of prostate cancer via a feedback loop related to TGF- β ¹². Meanwhile, miR-1269a can affect the occurrence and progression of hepatocellular cancer through targeting LRP6 and SPATS2L¹³. However, the expression of miR-1269a in NSCLC and the underlying mechanism has not been fully elucidated. In this study, we detected the expression of miR-1269a in 49 paired NSCLC and para-tumor tissues as well as in NSCLC cell lines. Subsequently, we constructed several functional experiments to identify the effect of miR-1269a on NSCLC proliferation and cell cycle. *SOX6* was also verified as a potential target for miR-1269a in NSCLC. Our study demonstrated miR-1269a as an onco-miR-

NA in NSCLC via regulating *SOX6*, which might provide a novel target for the biological therapy of NSCLC.

Patients and Methods

NSCLC and Adjacent Normal Tissues

All the 49 pairs of NSCLC tissues and adjacent normal tissues were obtained from surgical resection on NSCLC patients in the China-Japan Union Hospital of Jilin University from 2009 to 2011. All the samples were conserved in liquid nitrogen for subsequent experiments after removal. Informed consent was obtained from all patients and/or their families. Our study was approved by the Ethics Committee of the China-Japan Union Hospital of Jilin University.

Cell Lines and Cell Culture

All the 6 NSCLC cell lines (H1975, SPCA1, PC-9, H1299, H460, A549) and the human normal bronchial epithelium cell line (BEAS-2B) were obtained from American Type Culture Collection (ATCC) (Manassas, VA, USA). All the cells were cultured in DMEM (Dulbecco's modified eagle medium) (Invitrogen, Carlsbad, CA, USA) containing 10% v/v fetal bovine serum (FBS) (Corning, Rockville, MD, USA), 100 UI/mL penicillin (Gibco, USA), 100 µg/mL streptomycin (Gibco, Rockville, MD, USA), respectively, and incubated at 37°C, 5% CO₂ cell incubator.

Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Total RNA of NSCLC tissues, adjacent normal tissues, and NSCLC cell lines were extracted using TRIzol reagent (Invitrogen, Carlsbad, CA, USA). SYBR Green PCR Master Mix Kit (TaKaRa, Dalian, China) was used to detect the expression level of miR-1269a. Housekeeping U6 was applied as an internal control. The relative expression level of *SOX6* to glyceraldehyde 3-phosphate dehydrogenase (*GAPDH*) which was used as an internal control was measured by using the SYBR Green PCR Master Mix Kit (TaKaRa, Dalian, China). The relative expression level was calculated by the 2^{-ΔΔCT} method. Each experiment was repeated at least for three times. Primers for miR-1269a, U6, *SOX6*, and *GAPDH* were synthesized by Genewiz Co. (Shanghai, China). Primer sequences used were as follows: miR-1269a, F: 5'-ACGTTGGATGAAATCTCATGATAGGCCATC-3', R: 5'-ATTGCGTGTCGTGGAGTCGGCAATGC-3'; *SOX6*,

F: 5'-GCTGGAGGAGATGACTCGGAC-3', R: 5'-GTTCGGTCCACTATTGATGGGG-3'.

Cell Transfection

miR-1269a mimics, negative control (NC), inhibitors, and inhibitors negative control (INC) were bought from Genewiz Co. (Shanghai, China). Cells were transfected with these compounds using lipofectamine3000 (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. pcDNAs for *SOX6* overexpression were obtained from Ribobio Co. (Guangzhou, China) and were transfected with lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA). The efficiency of transfection was confirmed by qRT-PCR.

Cell Counting Kit-8 (CCK-8) Assay

Cells were treated with miR-1269a mimics, NC, inhibitors, or INC, respectively, and were seeded into 96-well plates. Subsequently, after culturing for 0 h, 24 h, 48 h, 72 h, a total of 10 µL CCK8 reagent (Dojindo, Kumamoto, Japan) was added to the cells for incubation for 1 h at 37°C. The data were measured by a 550 Microplate Reader (Bio-Rad, Hercules, CA, USA) was utilized to measure the absorbance at 470 nm. The experiments were repeated for three times.

Colony Formation Assay

H1975 and A549 cells were seeded into 6-well plates at the density of 400 per well, and were cultured for 2 weeks. After dyeing with crystal violet, the numbers of colonies with a diameter greater than 50 µm were measured. The experiments were repeated at least for three times.

Cell Cycle Analysis

Flow cytometry was employed to detect cell cycle distributions. H1975 and A549 cells were washed with cooling phosphate-buffered saline (PBS) after miR-1269a mimics or inhibitors transfection. Then, the cells were re-suspended in 1000 µL binding buffer mixing with 10 µL Propidium Iodide (PI) (Thermo Fisher, Waltham, MA, USA). Cell cycle was detected by flow cytometry (FACS, Arlesheim, Switzerland). The percentage of cells on G0/G1, S, and G2/M phase was recorded and measured.

Dual-Luciferase Assay

Dual-Luciferase Reporter Assay System Kit (Promega, Madison, WI, USA) was used to detect the luciferase activity. Vectors of the wild-type group or mutant group were constructed by

inserting a luciferase reporter vector with the region of the SOX6-3'-UTR containing the specific wild-type or mutant miR-1269a binding site, respectively. H1975 cells were co-transfected with the luciferase reporter vector and miR-1269a mimics according to the instruction of Lipofectamine3000 (Invitrogen, Carlsbad, CA, USA). Then, the luciferase activity was measured. The experiments were repeated for three times.

Western Blot Analysis

Total protein of the cells was extracted and quantified according to the Protein Extraction Kit Instructions (Beyotime, Nanjing, China). Loading Buffer Mix (Beyotime, Nanjing, China) was added, and the proteins were denatured at 100°C for 5 min. A total of 10 µL protein was used for sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), followed by transferring to the polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). 5% non-fat milk suspended in Tris-buffered saline-Tween (TBS-T) (Beyotime, Shanghai, China) was used to block non-specific protein loci for 2 h. The membranes were incubated with anti-SOX6 at 4°C overnight. After washing with TBST for 3 times, the membranes were incubated with HRP-labeled secondary antibody for 2 h at room temperature. After washing with phosphate-buffered saline-tween (PBS-T) for 3 times, the membranes were imaged on a gel imager. Finally, gray values of relative bands were analyzed. The expression of GAPDH was used as a reference, and the ratio of SOX6 to GAPDH was calculated and statistically analyzed.

Statistical Analysis

All quantitative data were expressed as mean ± standard deviation. The difference between groups was compared using One-way ANOVA, followed by the Post-hoc Test (Least Significant Difference). Statistical Product and Service Solutions (SPSS) 16.0 Version Software (SPSS Inc., Chicago, IL, USA) and GraphPad Prism Version 5.0 Software (San Jolla, CA, USA) were employed for data analysis. $p < 0.05$ was considered statistically significant.

Results

miR-1269a Was Over-Expressed in NSCLC Tissues and Cell Lines

To validate the expression level of miR-1269a in NSCLC, we detected miR-1269a expression

in 49 pairs of NSCLC tissues and adjacent normal tissues. Results showed that the expression of miR-1269a in NSCLC tissues was markedly higher than that of adjacent normal tissues (Figure 1A). The relative expression was shown in Figure 1B. Meanwhile, we measured the expression level of miR-1269a in NSCLC-derived cell lines and 1 normal bronchial epithelial cell line BEAS-2B. Results demonstrated that the expression of miR-1269a was markedly higher in NSCLC cell lines (Figure 1C). All these data above indicated that miR-1269a might act as an oncogene in NSCLC.

Up-Regulation of miR-1269a Accelerated the Cell Proliferation of NSCLC Cells

Next, to explore the effect of miR-1269a on NSCLC cells, we over-expressed miR-1269a in H1975 cells by transfecting miR-1269a mimics. The expression of miR-1269a was up-regulated about 8.03-fold in the miR-1269a mimics group compared to the negative control group (Figure 2A). The CCK8 assay demonstrated that transfection with miR-1269a mimics significantly promoted the proliferation of H1975 cells when compared with the NC group (Figure 2B). Moreover, we found that more colonies were formed in H1975 cells of the miR-1269a up-regulation group compared with the control group (Figure 2C, 2D). Furthermore, we detected the cell cycle of established H1975 cells and discovered that miR-1269 over-expression induced the transition of G0/G1 to S phase (Figure 2E). These data showed that up-regulation of miR-1269a accelerated cell growth and induced cell cycle transition of NSCLC cells.

Down-regulation of miR-1269a Inhibited the Cell Proliferation of NSCLC Cells

Similarly, we then knocked down the expression of miR-1269a in A549 cells. The expression level of miR-1269a in A549 cells was markedly decreased after miR-1269a inhibitors transfection (Figure 3A). The CCK8 experiment demonstrated that the knockdown of miR-1269a inhibited the cell proliferation of A549 cells (Figure 3B). Colony formation assay illustrated the same result with the CCK8 assay (Figure 3C, D). Moreover, miR-1269a inhibitors significantly blocked A549 cells in the G0/G1 phase when compared with the INC group (Figure 3E). These results indicated that down-regulation of miR-1269a inhibited cell proliferation and induced G0/G1 arrest of NSCLC cells.

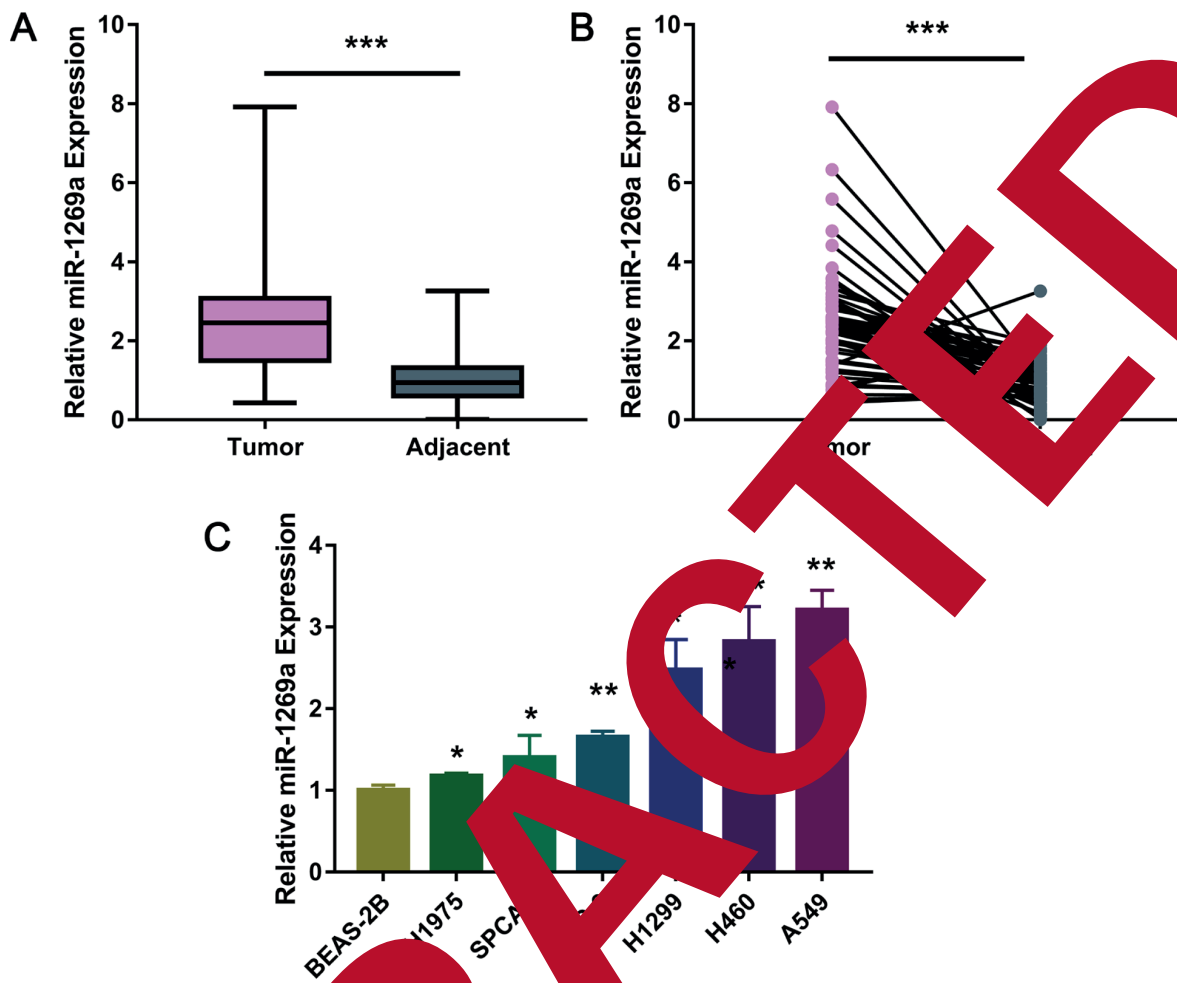


Figure 1. MiR-1269a expression in NSCLC tissues and cell lines. *A-B*, The expression levels of miR-1269a in NSCLC tissues and paired adjacent lung tissues were analyzed by RT-PCR. *C*, MiR-144-5p expression in BEAS-2B cells and NSCLC cell lines (H1975, SPCA1, PC-9, H1299, H460, A549) were shown as mean \pm SD. The experiments were repeated for three times. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

SOX6 Was a Direct Target of miR-1269a in NSCLC

Furthermore, to investigate the underlying mechanism of miR-1269a in NSCLC, we searched several databases, including miRBase, TargetScan and miRWalk. We found that *SOX6* might be a potential target for miR-1269a. We next confirmed this assumption by constructing the miR-REPORT vector carrying mutant or wild-type miR-1269a binding *SOX6*-3'-UTR region (Figure 4A). Dual-luciferase assay indicated that the activity of luciferase was significantly decreased in the wild-type group, but no difference was found in the mutant group (Figure 4B). We detected the mRNA expression of *SOX6* in NSCLC tissues and adjacent

normal tissues, respectively, and found an evident decrease of *SOX6* mRNA expression in the NSCLC group (Figure 4C). Meanwhile, Western blot was used to measure the protein expression of *SOX6* protein in NSCLC cells. Over-expression of miR-1269a inhibited the protein expression of *SOX6* in H1975, whereas down-regulation of miR-1269a increased the protein expression of *SOX6* protein in A549 (Figure 4D, 4E). In addition, we analyzed the relationship between miR-1269a and *SOX6* in 49 pairs of NSCLC tissues and adjacent normal tissues, and found that there was a significant negative correlation ($R^2 = 0.5928$, $p < 0.0001$) (Figure 4F). All the results illustrated that *SOX6* was a target gene of miR-1269a in NSCLC.

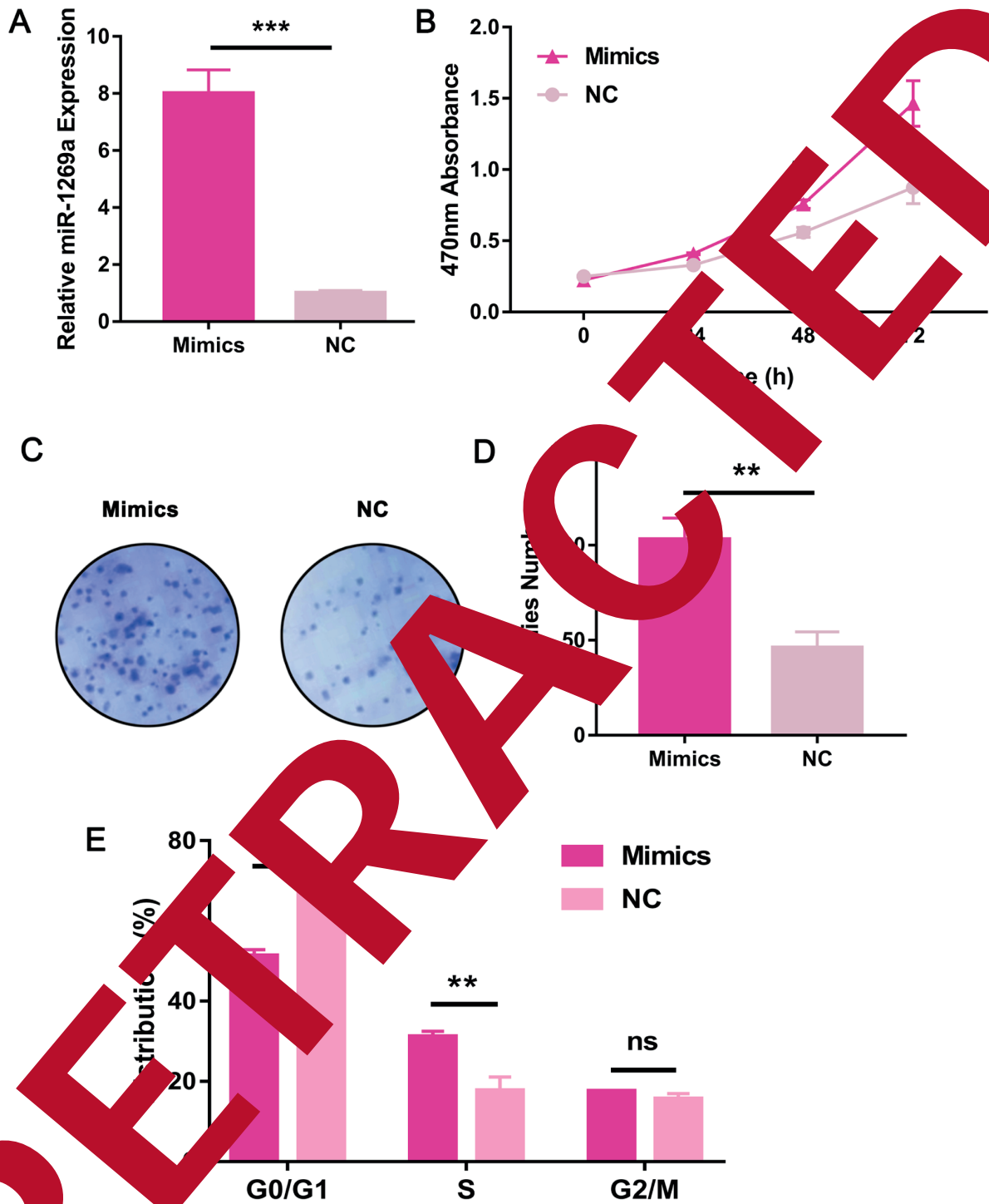


Figure 1. Expression of miR-1269a promoted proliferation and induced cell cycle of H1975 cells. **A**, H1975 cells were transfected with either miR-1269a mimics or negative control (NC). **B**, Cell viability was detected by the CCK8 assay. **C**, Colony formation assay performed in H1975 cells. **D**, Quantitative analysis of the relative colony numbers to the negative control. **E**, After transfection for 48 h, H1975 cells were stained with propidium iodide, and the distribution of cell cycle was detected by flow cytometric analysis. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

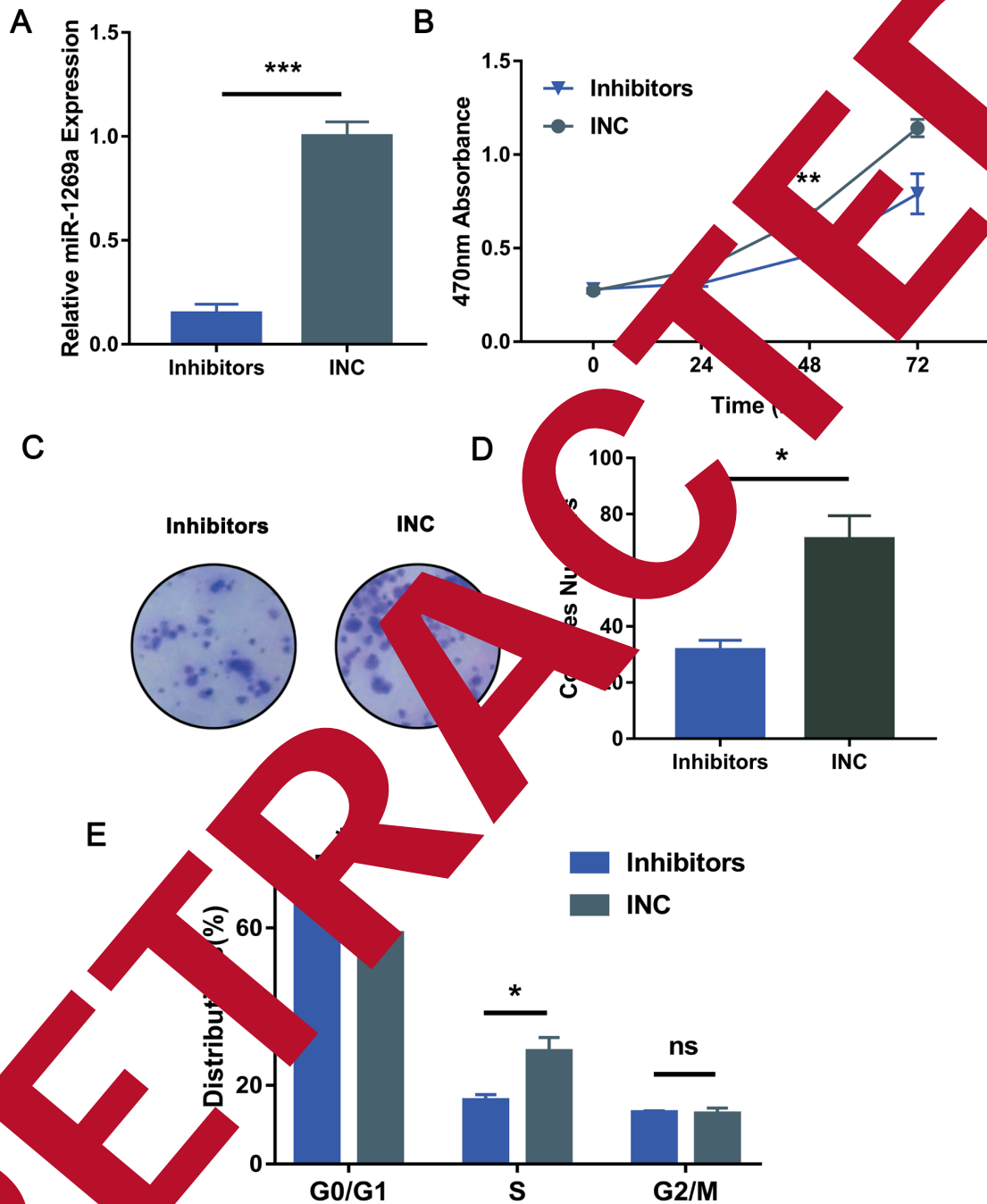


Figure 3. Down-regulation of miR-1269a inhibited proliferation and cell cycle of A549 cells. **A**, A549 cells were transfected with miR-1269a inhibitors or inhibitors negative control (INC). **B**, Cell viability was detected by the CCK8 assay. **C**, Colony formation assay performed in A549 cells. **D**, Quantitative analysis of the relative colony numbers to the INC group. **E**, After transfection for 48 h, A549 cells were stained with propidium iodide, and the distribution of cell cycle was determined by flow cytometric analysis. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

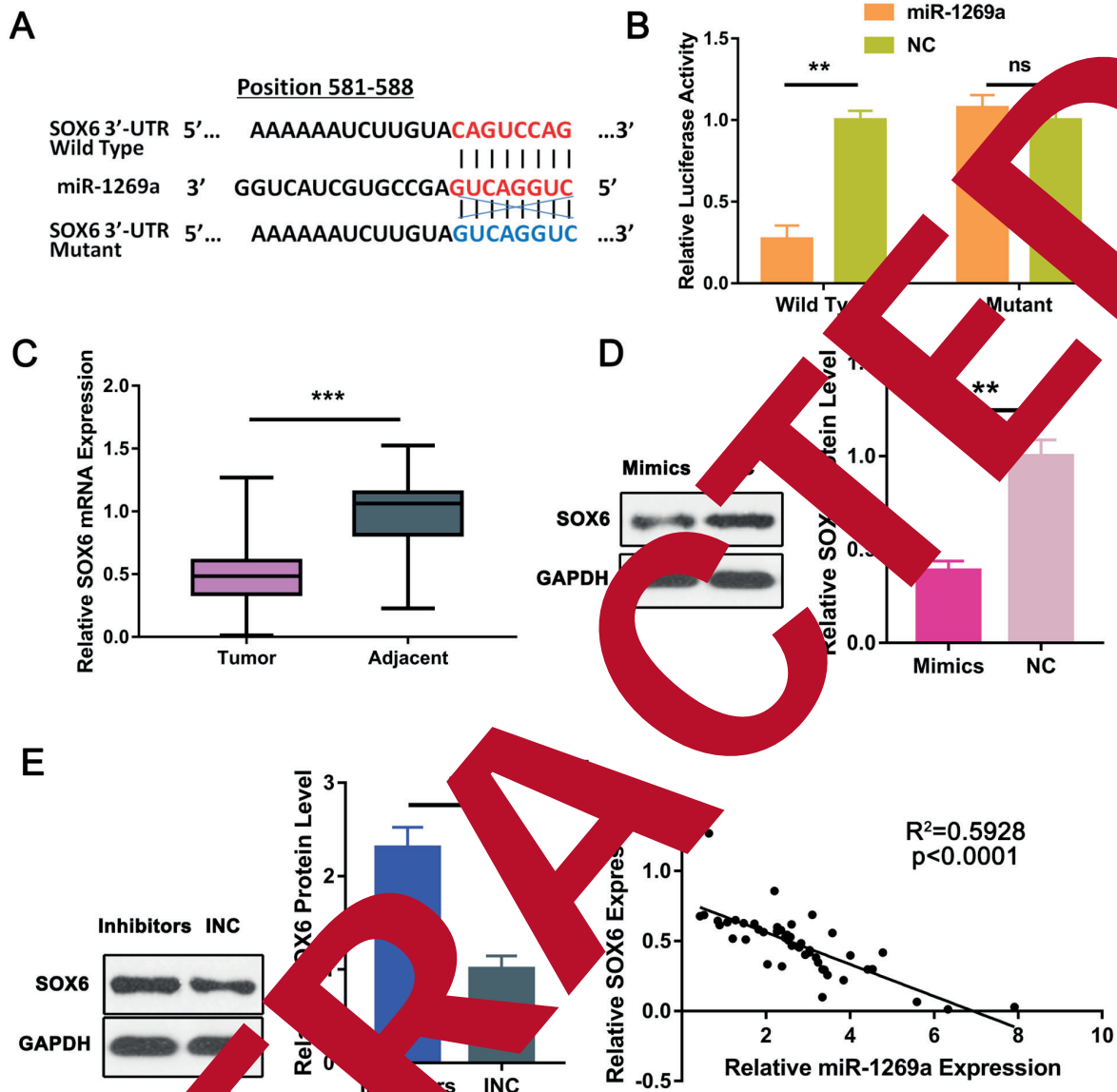


Figure 4. MiR-1269a targets SOX6 in NSCLC. **A**, The predicted binding site of miR-1269a at the 3'-UTR of SOX6 mRNA. **B**, Luciferase reporter assay using pMIR-REPORT (vector), pMIR-REPORT SOX6-3'-UTR-wt, or SOX6-3'-UTR-mut plasmid in H1975 cells transfected with miR-1269a mimics or NC miRNA. **C**, The mRNA expression of SOX6 in NSCLC tissues and paired adjacent normal tissues were analyzed by qRT-PCR. **D-E**, The protein expression of SOX6 in established H1975 and H460/549 cells was detected by Western blot analysis. **F**, Correlation between miR-1269a expression and SOX6 mRNA level in NSCLC tissues ($R^2=0.5928$, $p<0.0001$). * $p<0.05$, ** $p<0.01$, *** $p<0.001$, ns=non-sense.

miR-1269a Promoted NSCLC Growth Via repressing SOX6

To confirm our results, we over-expressed SOX6 in H1975 cells transfected with miR-1269a mimics. The protein expression of SOX6 was significantly increased after SOX6 up-regulation (Figure 5A). Next, CCK8 and colony formation assays were used to explore the changes of

cell proliferation. The enhancement of miR-1269a mimics on H1975 cell growth was significantly offset by SOX6 up-regulation (Figure 5B, 5C, 5D). Meanwhile, SOX6 over-expression restored the decreased distributions of G0/G1 phase caused by miR-1269a up-regulation (Figure 5E). These data demonstrated that miR-1269a could act as onco-miRNA via repressing SOX6 expression.

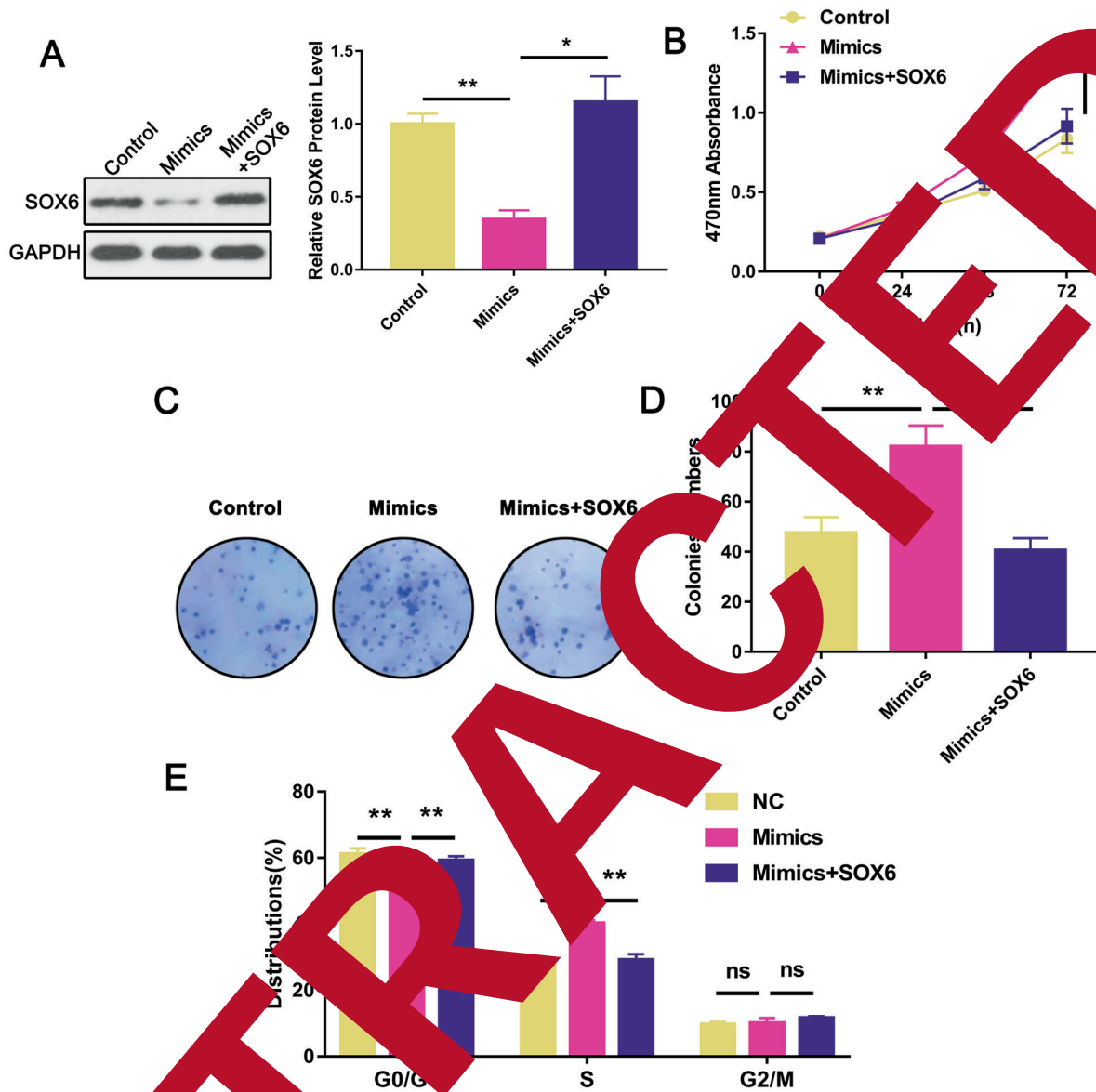


Figure 5 SOX6 over-expression reversed miR-1269a-mediated promotion of the growth of H1975 cells. **A**, The protein expression of SOX6 was detected by Western blot analysis. GAPDH was used as an internal control. **B**, Cell proliferation ability of H1975 cells treated with negative control, mimics, or mimics+SOX6 were measured by the CCK8 assay. **C-D**, Cell growth ability was measured by colony formation assay. **E**, The distribution of cell cycle was determined by flow cytometric analysis. Data represent as mean \pm SD. The experiments were repeated for three times. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Discussion

NSCLC, especially NSCLC, contributes to the highest incidence and mortality of malignancy [14,15]. Despite of the continuous improvement and development of various therapeutic technologies in recent years, the prognosis of NSCLC remains poor [16,17]. MiRNAs, a classic class of regu-

latory factors involved in the biological behavior of many cancers, have been found to be a meaningful target for the diagnosis, prognosis and treatment of tumors [18,19].

miR-1269a has been found to be involved in the biological processes of many tumors. For example, it can participate in the occurrence and development of primary liver cancer, affect the re-

sistance of prostate cancer in castration therapy, and involve in the oncogenesis of gastric cancer. These effects are often achieved by regulating the expression of specific target genes, including MYCBP2, ZNF70, SPATS2L, and LRP6^{12,13,20,21}. However, the role of miR-1269a in NSCLC has not yet been confirmed. Here, we firstly identified that the expression of miR-1269a was increased in NSCLC tissues and cell lines compared to normal controls. Several functional experiments illustrated that over-expression of miR-1269a promoted cell proliferation and induced G0/G1 to S transition, while knockdown of miR-1269a inhibited cell growth and induced G0/G1 arrest of NSCLC cells. These results suggested that ectopic miR-1269a expression could affect the development and progression of NSCLC.

Furthermore, we found SOX6 as a potential target of miR-1269a in NSCLC. SOX6, a member of the Sox transcription-factor family, has been shown to be a tumor suppressor in many researches. The expression of SOX6 is low in multiple tumors. SOX6 usually affects cell proliferation and cell cycle distributions by affecting the expression of its downstream p21 and cyclin D1²²⁻²⁴. For example, in malignant tumors such as liver cancer, esophageal cancer, osteosarcoma, and pancreatic tumor, SOX6 acts as a tumor suppressor gene. Moreover, its decreased expression may promote the occurrence and progression of tumors. In addition, SOX6 can be regulated by many miRNAs. For example, in human epidermal squamous cell carcinoma, miR-208 accelerates cell growth by down-regulating SOX6²⁵. miR-76 promotes human colorectal cancer cell proliferation by targeting SOX6. miR-96 increases proliferation and metastasis of hepatocellular carcinoma through regulating SOX6^{29,30}. In this study, we confirmed that SOX6 could regulate NSCLC growth via acting as a direct target gene of miR-1269a, which was familiar to mutual evidence of many previous studies. Meanwhile, with over-expression of SOX6, the promotion effect of miR-1269a up-regulation was significantly offset. Our results demonstrated miR-1269a could act as an oncogene in NSCLC via up-regulating the expression of SOX6. However, more researches are needed for further illustrating the mechanism of miR-1269a in NSCLC.

Conclusions

This study demonstrated that miR-1269a was significantly over-expressed in NSCLC tissues and cell lines. Moreover, miR-1269a might pro-

mote cell proliferation and cell cycle of NSCLC cells via down-regulating the tumor suppressor SOX6. These findings might provide a new target for NSCLC diagnosis and biological therapy.

Conflict of Interest

The Authors declare that they have no conflict of interest.

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