

Effect of exosomes derived from mir-126-modified mesenchymal stem cells on the repair process of spinal cord injury in rats

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Abstract. – OBJECTIVE: To investigate the effect of the micro ribonucleic acid (miR)-126-modified mesenchymal stem cell (MSC)-derived exosomes (MSC-exos) on the repair process of spinal cord injury (SCI) in rats.

MATERIALS AND METHODS: MiR-126-modified MSCs were cultured, the exosomes were extracted, and a rat model of SCI was established. The Quantitative Polymerase Chain Reaction (qPCR) was carried out to detect the expression of miR-126 in the injured spinal cord. The Western blotting (WB) was adopted to detect the expressions of the exosome-related molecules. Subsequently, the motor function recovery of rats was examined via the Basso, Beattie and Bresnahan (BBB) locomotor rat scale. The effects of miR-126 exosomes on SCI injury volume and NeuN retention in spinal cord were evaluated via immunohistochemistry.

RESULTS: MSCs were able to package miR-126 into exosomes. After SCI, the recovery of the hind limb function of rats was remarkably improved by miR-126-modified MSC-exos relative to the control group. Treatment with miR-126-modified MSC-exos remarkably decreased SCI injury volume, retained the neuronal cells, and triggered the axon regeneration following SCI. Besides, the expression of Ras homolog gene family member A (RhoA), identified as the downstream gene of miR-126, was downregulated in the group with miR-126-modified MSC-exos. Moreover, miR-126-modified MSC-exos activated the extracellular regulated protein kinases 1/2 (ERK1/2) pathway.

CONCLUSIONS: MiR-126-modified MSC-exos protect neurons of rats with SCI, stimulates axon regeneration, and improves the recovery of limb motor function after SCI.

Key Words:

MiR-126, Mesenchymal stem cell, Exosome, Spinal cord injury repair.

Introduction

Traumatic Spinal Cord Injury (SCI) as severe damage to the central nervous system (CNS), often leads to irreversible neurological deficits. SCI involves complex pathophysiology as its direct primary mechanical injury is accompanied by a series of secondary processes including neuroinflammation and ischemia that aggravate SCI¹. In spite of profound breakthroughs in preclinical studies on the promotion and regeneration of neurons so far, the specific pathophysiological mechanism is still unclear. Therefore, basic science and clinical researchers are still facing a challenge to find an effective treatment method for SCI.

As an endogenous non-coding ribonucleic acid (RNA), a micro RNA (miRNA) can bind to the 3' untranslated region (3'-UTR) of its target gene messenger RNA (mRNA), so as to exert the post-transcriptional regulation². MiRNAs have been revealed in recent studies to exert crucial effects in synaptic activity, regeneration, and neurogenesis of the CNS. Besides, it has been reported that several miRNAs are regarded as the underlying novel targets for treating SCI^{3,4}. Yu et al⁵ reported that miR-133b is critical for the function recovery after SCI in zebrafishes. Moreover, Gordon et al⁶ have also indicated that miR-126 targets Ras homolog gene family member A (RhoA) to accelerate the neurite outgrowth *in vitro*.

Exosomes are a class of small membrane vesicles derived from the lumen of a multivesicular body (30-100 nm in diameter). Exosomes are considered as mediators of intercellular communication by transporting various RNAs, DNAs, and proteins between cells⁷. Many studies⁸ have shown that, exosomes, as intercellular communicators, function both locally and systematically.

Additionally, some investigations have evidenced that exosomes contribute to the transportation of therapeutic miRNAs to cells producing exosomes. There are numerous cell types that could produce exosomes, and MSCs are one of the most common types. MSCs have been confirmed in multiple investigations to be the ideal candidate cells for cell therapy of SCI. It is reported that MSCs could promote tissue repair by inhibiting the inflammatory response and activating endogenous repair mechanisms. It has been verified in a previous study that MSCs decreased the secretion of multiple inflammatory cytokines [tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1), and IL-6] and blocked the activation of the nuclear factor kappa-B (NF- κ B)⁹. Furthermore, MSCs are able to guide the transformation of the stimulated macrophages from a proinflammatory M1 phenotype to an anti-inflammatory M2 phenotype. It has been revealed in a recent study that the therapeutic effect of MSCs may be mediated by exosomes¹⁰. According to reports, MSC-derived exosomes (MSC-exos) can mimic most of the biological functions of MSCs, and also attenuate inflammatory response and apoptosis, reduce the level of proinflammatory cytokines, and inhibit the activation of astrocytes and macrophages¹¹. Similar to that of MSC, the infusion of MSC-exos is capable of suppressing the activation of NF- κ B. Besides, the therapeutic effect of the cell-free exosomes is equivalent to that of the intact MSCs after brain injury. According to a previous study, the application of MSCs in the SCI model has manifested that a systemic administration can lower the level of the pro-inflammatory cytokines and improve the recovery of function. Nevertheless, the exact beneficial mechanism to SCI is still unclear.

On the basis of related studies reported, it is speculated that miR-126b-modified MSC-exos administration could improve the functional recovery of the injured spinal cord by transferring miR-126b to the lesion site.

Materials and Methods

Animals

Male adult Sprague-Dawley rats (80-100 g) purchased from the Animal Center of the Chinese Academy of Sciences in Shanghai, China, were used in this study. All of the experimental procedures were approved by the Animal Care and Use Committee of our hospital and the rats were raised in line with the guidelines for the care and

use of laboratory animals approved by the National Institutes of Health.

Preparation of MiR-126-Modified MSC-Exos

The primary MSCs were isolated from the bone marrow of male rats. Briefly, femur and tibia bone marrow were separated and rinsed with Phosphate-Buffered Saline (PBS). Centrifuged precipitates were suspended and subjected to thermal inactivation in Dulbecco's Modified Eagle's Medium (DMEM; Life Technologies, Gaithersburg, MD, USA) containing 10% fetal bovine serum (FBS; Life Technologies, Gaithersburg, MD, USA) and 1% penicillin-streptomycin. Then, they were maintained in a incubator (5% CO₂, 37°C).

MSCs were then transfected with miR-126 mimics or negative control using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA). The exosomes were harvested from the supernatant using the Quick-TC kit (System Biosciences, Palo Alto, CA, USA) at 72 h. Subsequently, the exosome precipitates were resuspended in PBS at 10 μ g/ μ L.

Construction of the SCI Model

All rats were anesthetized with 400 mg/kg ketamine hydrochloride before the operation. After the laminectomy, the paraspinal muscle was dissected, T9-T11 laminectomy was conducted. After that, the muscle was kept with the aneurysm clamp at T10 level with a closing force of 35 g for 60 s, and finally the incision was sutured with the silk thread. The rats in the sham operation group only received laminectomy. All rats were administrated with penicillin and analgesics for 3 days after the operation and urinated manually three times a day. At 24 h after trauma, the rats were injected miR-126 exosomes (100 μ g exosomes in 0.5 mL PBS) and miR-con exosomes were treated with PBS (0.5 mL) in the tail vein.

Animal Grouping

The rats were randomly allocated into 4 groups by a blind method: the sham operation group (sham operation); the control group (SCI and PBS treatment); the miR-con group (SCI and miR-con exosomes treatment); and the miR-126 group (SCI and miR-126 exosomes treatment).

Immunohistochemistry

The rats were finally anesthetized with isoflurane on the 4th day after injury. The spinal cord was embedded into compounds with the optimal cutting temperature. Then, T9-T11 segments of the

spinal cord near the lesion center were collected for histological evaluation. Hematoxylin and eosin (H&E) staining was conducted to quantify the lesion degree by Imagepro-Plus software. T9-T11 segments of the spinal cord were also dissected in 6 rats in each group, and 8 sections of each rat were selected to calculate cystic cavity size.

Subsequently, the sections were deparaffinized in xylene, dehydrated in gradient ethanol, and then boiled twice in citrate buffer (pH 6.0) for 5 min. Thereafter, the cooled sections were incubated with 3% H₂O₂ at room temperature for 15 min to inactivate the endogenous peroxidase. Then, the glass slides were sealed with 10% FBS for 10 min before incubation with the corresponding primary antibody at 4°C overnight. After TBS washing, the slides were incubated with the fluorescent-labeled secondary antibody (Abcam, Cambridge, MA, USA). Ultimately, the sections were stained with 4',6-diamidino-2-phenylindole (DAPI) and visualized under a confocal laser scanning microscope (Olympus LSM-GB200, Tokyo, Japan).

RNA Extraction and q-PCR

The tissues and cells were lysed using the TRIzol kit (Invitrogen, Carlsbad, CA, USA) followed by the measurement of RNA concentration using an ultraviolet spectrophotometer (Hitachi, Tokyo, Japan). The complementary Deoxyribonucleic Acid (cDNA) was synthesized following the protocols of the PrimeScript RT MasterMix kit (Invitrogen, Carlsbad, CA, USA). The thermal cycle protocols were as follows: 4°C for 30 s, 55°C for 30 s, and 72°C for 90 s for a total of 40 cycles. The target gene level was calculated by the 2^{-ΔΔCt} method. The primer sequences used in this study were as follows: miR-126, F: 5'-GCTTCGCTGCACATAACTGCCTG-3', R: 5'-AGGTTCTGCTGGAGACAG-3'; U6: F: 5'-GCTTCGCTGCACATAACTAAAAT-3', R: 5'-GCTTCGCTGCACATAACTGCGTGTCAT-3'.

Western Blotting (WB)

Total proteins were isolated with radio-immunoprecipitation assay (RIPA) buffer (Beyotime, Shanghai, China), separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto a polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA). After the proteins were successfully transferred, the membrane was sealed with 5% skim milk at 37°C for 2 h and reacted with the primary antibodies (Cell Signaling Technology, Danvers, MA, USA) at 4°C over-

night. The membranes were then washed with TBST (Tris-buffered Saline with Tween 20) and followed by the incubation of the secondary antibodies conjugated with horseradish peroxidase at 37°C for 1 h. The exposed bands were observed with enhanced chemiluminescence (ECL) detection kit.

Statistical Analysis

The data were analyzed by Statistical Product and Service Solutions (SPSS) 20.0 (IBM Corp., Armonk, NY, USA). All the data collected from at least three independent experiments were expressed as mean ± standard deviation (SD). A significant difference was determined at $p < 0.05$.

Results

MiR-126 Expression Was Reduced in Injured Rats

MiR-126 level in the SCI rats was measured via qRT-PCR at 12 h, 24 h, 2 days, 3 days, 4 days, 5 days, and 7 days following acute SCI. It was found that miR-126 was evidently down-regulated since 24 h following SCI ($p < 0.05$) (Figure 1).

MiR-126 Level in MSC-Exos

MSCs manifested as cluster of differentiation 73⁺ (CD73⁺), CD90⁺, CD105⁺ CD34⁻, and CD45⁻ were cultured as described above. The transfection efficiency of miR-126 mimic reached about 90%. The exosomes were isolated from MSC supernatant 72 h after transfection. Subsequently, WB analysis was carried out and the com-

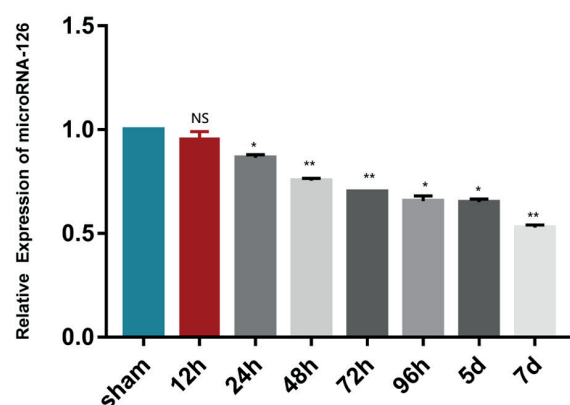


Figure 1. Expression level of miR-126 in the injured spinal cord measured by qRT-PCR compared with those in control group at 12 h, 24 h, 2 day, 3 day, 4 day, 5 day, and 7 day after acute SCI (NS: $p > 0.05$, * $p < 0.05$, ** $p < 0.01$).

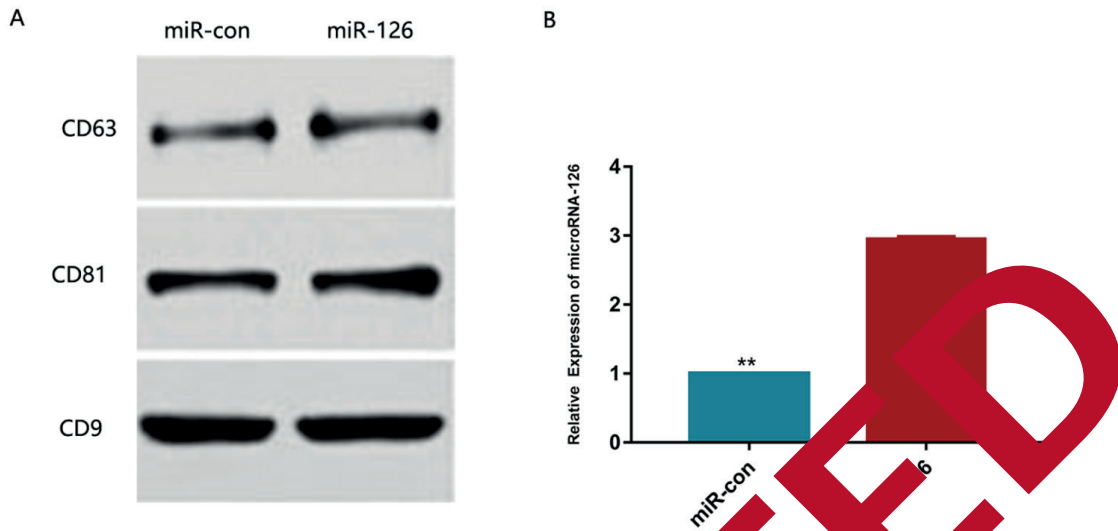


Figure 2. Expressions of CD63, CD81 and CD9 in MSC-exos transfected with miR-126 or miR-con evaluated by WB and qRT-PCR.

monosome markers, including CD9, CD63, and CD81 were detected (Figure 2A). QRT-PCR manifested that miR-126 level was about 3.1 times higher in MSC-exos overexpressing miR-126 than that of the controls ($p < 0.01$) (Figure 2B). The above results proved that miR-126 was effectively packaged into secreted exosomes from MSCs.

MiR-126-Modified Exosomes Contribute to Function Recovery and Injury Volume Decline

First, qRT-PCR was adopted to measure the miR-126 level in spinal cord compared with the control group. miR-126-modified exosome markedly increased the miR-126 level on the 4th day after SCI (Figure 3A).

In order to investigate whether miR-126-modified exosomes enhanced the motor function recovery following acute SCI, the BBB locomotor scale was performed at different time points to assess our SCI model. After SCI, 0-1 point of BBB score indicated the successful modeling of SCI. As mentioned earlier (after SCI), we observed a spontaneous functional recovery in all groups. After 5 days, a significant difference was observed in the functional recovery between the rats injected with miR-126 exosomes and those injected with miR-con exosomes ($p < 0.01$, on the 9th day: $p < 0.001$, and on the 14th day: $p < 0.01$), indicating that miR-126-mediated triggered the recovery of the hind limb motor function after SCI (Figure 3B).

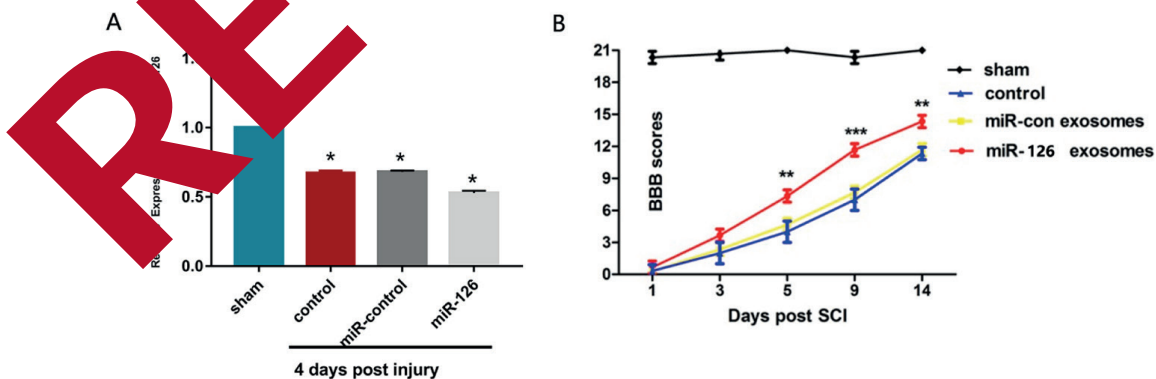


Figure 3. Injection of miR-126 exosomes improved the recovery after SCI. **A**, After SCI, the relative expression of miR-126 in the lesion site compared with that in control group is detected by qRT-PCR, and the expression of miR-126 is notably increased by tail vein injection of miR-126 exosomes ($*p < 0.05$). **B**, The recovery of function (BBB score) is monitored on the 1st, 3rd, 5th, 9th, and 14th day after lower limb SCI ($***p < 0.001$, $**p < 0.01$, $*p < 0.05$, $n = 4$).

Effects of MiR-126 Exosomes on the Injury Volume and Neuron Retention After SCI

Next, the effects of miR-126-mediated exosomes on the injury volume and NeuN retention after SCI were detected via immunohistochemistry. HE staining demonstrated that, compared with that in miR-con or control group, miR-126 exosomes markedly reduced the lesion cavity area (Figures 4A and 4B). On the 4th day after SCI, NeuN, a specific marker of mature neurons, was stained to visualize the number of neurons in the injured spinal cord. Relative to rats receiving miR-con exosomes, the number of mature neurons in the injured rats receiving miR-126 exosomes was significantly increased ($p<0.01$) (Figures 5A and 5B).

MiR-126 Exosomes Promoted Axon Growth Following SCI

To study the underlying mechanisms of miR-126 exosomes on axon growth, WB was carried out for growth-associated protein 43 (GAP43) and NF. GAP43 level in miR-126 exosome group was raised compared with that in miR-con or control group (Figure 6A). Besides, the treatment with miR-126 exosomes also elevated NF expression on the 4th day after SCI (Figure 6B).

Effects of MiR-126 Exosomes on the Phosphorylation of cAMP Response Element-Binding Protein (CREB) and the Activation of Signal Transducer and Activator of Transcription 3 (STAT3) Pathways

RhoA is the direct target of miR-126 as previously indicated. In addition, RhoA has been

reported to participate in the neuronal cell death in the spinal cord. Compared with the miR-con exosomes group, RhoA level in the miR-126 exosomes group was notably reduced ($p<0.05$) (Figure 7A). Furthermore, the phosphorylation of ERK1/2 was pronounced in the SCI rats treated with miR-126 exosomes (Figure 7B).

Discussion

SCI has been showing a great impact on patients and society. The functional recovery following SCI is usually slight and there is no effective treatment method clinically applied so far. Many studies have indicated the involvement of miRNAs in the pathogenesis of SCI. For example, it has also been proved that miR-126 reduces neuronal cell death and promotes functional recovery after SCI. Long et al¹³ have shown that miR-126 exosomes crucial effects neuronal differentiation, apoptosis, and neurite outgrowth in the CNS¹³. Moreover, further research has revealed that miR-126 overexpression is able to improve the functional recovery in rats with stroke. The exosome is a new type of intercellular communication device which has been applied as a biological carrier for local or systemic miRNA delivery and treatment for stroke and Parkinson's disease¹⁴. In this paper, the role of exosome-mediated miR-126 transfer in SCI therapy was explored. After SCI, evident differences in the functional recovery were identified in each group. Niu et al¹⁵ manifested that there were a variety of treatment methods improving the functional recovery 3 days after SCI. It was observed in this study that at 5 days

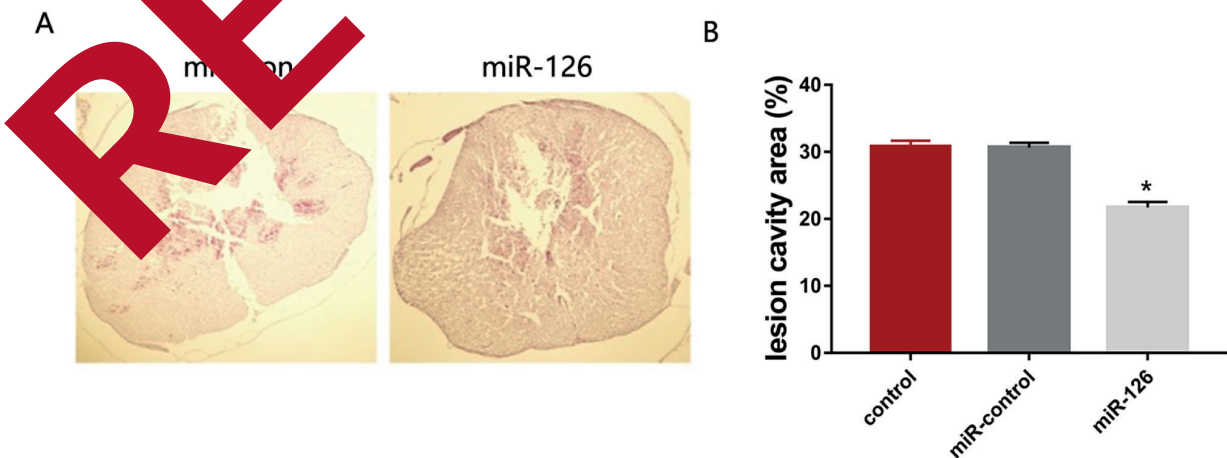


Figure 4. Injection of miR-126 exosomes reduces the lesion volume and retains NeuN after SCI (magnification $\times 10$) (* $p<0.05$).

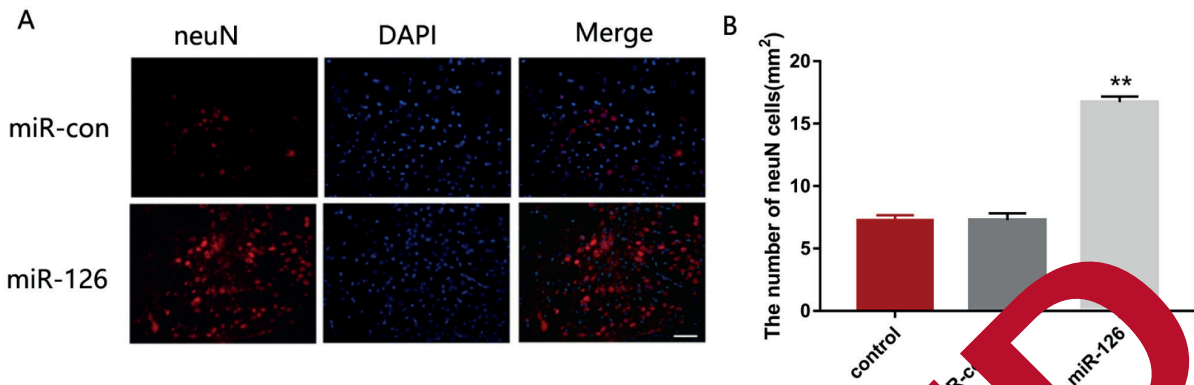


Figure 5. Effect of miR-126 exosomes on NeuN evaluated by calculating the cell number through immunofluorescence staining (magnification $\times 40$) (** $p < 0.01$).

after SCI, the BBB score of the rats injected with miR-126 exosomes was remarkably higher than that of the rats injected with miR-con exosomes. Lentiviral delivery of miR-126b promotes the recovery in the SCI mice, which was consistent with our results¹⁶.

Several mRNAs, such as RhoA and MST2, have been identified as the target genes of miR-126¹⁷. Here, we found that the RhoA level exhibited a significant decrease in SCI rats receiving miR-126 exosomes. Together with the results of our previous study *in vitro*, our data suggested that RhoA is a target of miR-126¹⁸. RhoA, a member of the Rho family, is up-regulated in the SCI rats and act on the downstream effector Rho-associated protein kinase (ROCK). Besides, Umezaki et al¹⁹ indicated that RhoA-ROCK pathway exerts a vital effect on the neuronal death in SCI mice. Thus, whether the survival of neurons after the injection of miR-126 exosomes was enhanced *in vivo* was further investigated. Within 24 hours, the neurons were exposed to mechanical forces and

secondary injuries followed SCI. In this study, miR-126 exosome group showed a notably increased number of mature neurons on the 4th day after SCI. Our results revealed that the silence of RhoA evidently reduced cell death following SCI. Besides, the treatment of miR-126 exosomes markedly enhanced the ERK1/2 phosphorylation level at the injured site of the spinal cord. MiR-126 activated ERK1/2 phosphorylation *via* RhoA. Our results also demonstrated a neuroprotective effect of miR-126 exosomes attributing to RhoA downregulation and ERK1/2 activation.

It is determined that the presence of the inhibitory molecules, including oligodendrocyte myelin glycoproteins and myelin-associated glycoproteins, inhibits axonal regeneration in the CNS. To reveal the effect of miR-126 exosomes on neurite outgrowth, the immunohistochemical staining was performed to analyze the NF expression in the spinal cord. The NF expression was observed to be increased in miR-126 exosome group, which is consistent with the results of the WB. GAP43

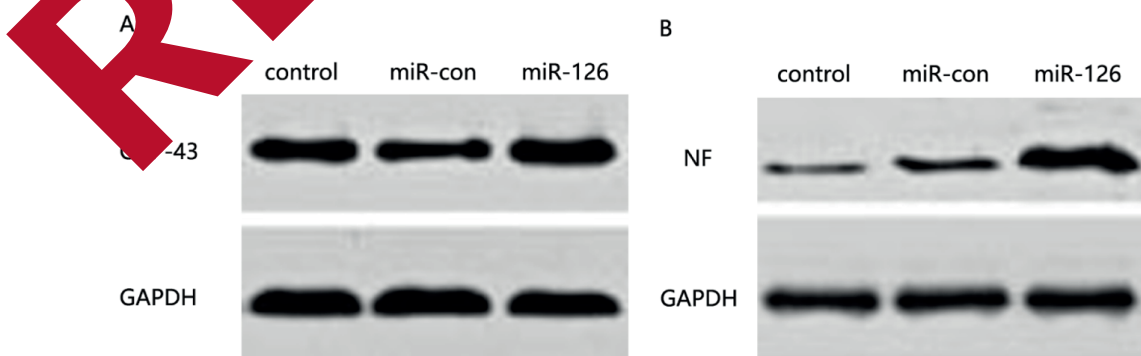


Figure 6. WB results show that miR-126 exosomes can increase the expressions of GAP43 (A) and NF (B).

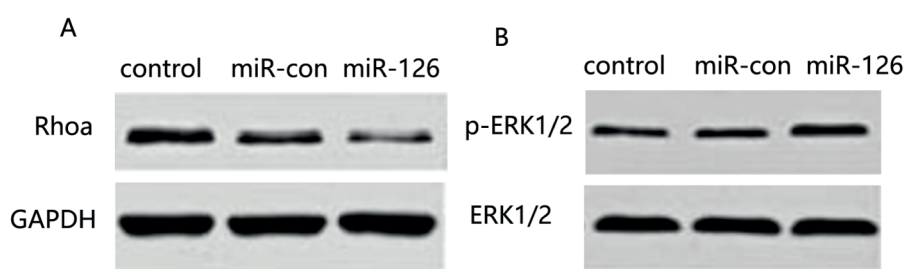


Figure 7. WB results indicate that miR-126 exosomes are capable of lowering the expression of RhoA (A) and decreasing the phosphorylation of ERK1/2 (B).

is a regeneration-related gene that is up-regulated in regenerative neurons²⁰. We found that the GAP43 expression in SCI rats injected with miR-126 exosomes was remarkably raised on the 4th day following SCI, indicating that miR-126 exosomes promote axon regeneration in the injured spinal cord.

Zhang et al²¹ have indicated that the transcription factor CREB played a vital role in axon regeneration. The activation of CREB has been shown to counteract myelin inhibitors and trigger the regeneration of spinal axons. In this aspect, it was discovered that miR-126 exosomes activated CREB expression in SCI rats. In addition, phosphorylated STAT3 (involved in spinal axon regeneration) in miR-126 exosome-treated SCI rats markedly increased. It can be seen that miR-126 exosomes enhanced the axon regeneration after SCI by enhancing the phosphorylated levels of CREB and STAT3.

Conclusion

The systemic injection of miR-126 exosomes could up-regulate the miR-126 level on the lesion spinal cord and promote the functional recovery following SCI. Additionally, it was also observed that miR-126 exosomes retained residential characteristics and increased axon regeneration, which partly resulted from the activation of ERK1/2, STAT3, and CREB and the inhibition of the RhoA expression. The above findings imply that miRNA-modified MSC-exos can be used as a new treatment method for SCI.

Conflict of Interests

The Authors declare that they have no conflict of interests.

References

- 1) BARTEL DP, ZHANG CZ. MicroRNAs: regulators of gene expression in development and disease. *Nat Rev Genet* 2004; 5: 396-400.
- 2) ZHANG X, CATALUCCI M, FELICETTI F, BONCI D, ADDARIO A, GALLO P, BANG ML, SERRALINI P, GU Y, DALTON ND, ELIA L, LATRONICO MM, HOYDAL M, AUTORE C, RUSSO MA, DORN GN, ELLINGBOE O, RUIZ-LOZANO P, PETERSON KL, CROCE TM, PESCHLE C, CONDORELLI G. MicroRNA-133 controls cardiac hypertrophy. *Nat Med* 2007; 13: 613-618.
- 3) DATTO SJ, BASTIDAS JC, MILLER NL, SHAH AK, ARHEART KL, MARCILLO AE, DIETRICH WD, PEARSE DD. Female sex hormones demonstrate improved locomotor recovery and greater preservation of white and gray matter after traumatic spinal cord injury compared to males. *J Neurotrauma* 2015; 32: 1146-1157.
- 4) ESPOSITO E, RINALDI B, MAZZON E, DONNICUO M, IMPELLIZZERI D, PATERNITI I, CAPUANO A, BRAMANTI P, CUZZOCREA S. Anti-inflammatory effect of simvastatin in an experimental model of spinal cord trauma: involvement of PPAR- α . *J Neuroinflammation* 2012; 9: 81.
- 5) YU YM, GIBBS KM, DAVILA J, CAMPBELL N, SUNG S, TODOROVA TI, OTSUKA S, SABAAYW HE, HART RP, SCHACHNER M. MicroRNA miR-133b is essential for functional recovery after spinal cord injury in adult zebrafish. *Eur J Neurosci* 2011; 33: 1587-1597.
- 6) GORDON BS, KAZI AA, COLEMAN CS, DENNIS MD, CHAU V, JEFFERSON LS, KIMBALL SR. RhoA modulates signaling through the mechanistic target of rapamycin complex 1 (mTORC1) in mammalian cells. *Cell Signal* 2014; 26: 461-467.
- 7) HU J, ZENG L, HUANG J, WANG G, LU H. MiR-126 promotes angiogenesis and attenuates inflammation after contusion spinal cord injury in rats. *Brain Res* 2015; 1608: 191-202.
- 8) HU JZ, HUANG JH, ZENG L, WANG G, CAO M, LU HB. Anti-apoptotic effect of microRNA-21 after contusion spinal cord injury in rats. *J Neurotrauma* 2013; 30: 1349-1360.
- 9) JEE MK, JUNG JS, CHOI JI, JANG JA, KANG KS, IM YB, KANG SK. MicroRNA 486 is a potentially novel target for the treatment of spinal cord injury. *Brain* 2012; 135: 1237-1252.

- 10) LI F, JIANG Q, SHI KJ, LUO H, YANG Y, XU CM. RhoA modulates functional and physical interaction between ROCK1 and Erk1/2 in selenite-induced apoptosis of leukaemia cells. *Cell Death Dis* 2013; 4: e708.
- 11) LIU D, HUANG Y, JIA C, LI Y, LIANG F, FU Q. Administration of antagomir-223 inhibits apoptosis, promotes angiogenesis and functional recovery in rats with spinal cord injury. *Cell Mol Neurobiol* 2015; 35: 483-491.
- 12) LIN CA, DUAN KY, WANG XW, ZHANG ZS. MicroRNA-409 promotes recovery of spinal cord injury by regulating ZNF366. *Eur Rev Med Pharmacol Sci* 2018; 22: 3649-3655.
- 13) LONG Q, UPADHYA D, HATTIANGADY B, KIM DK, AN SY, SHUAI B, PROCKOP DJ, SHETTY AK. Intranasal MSC-derived A1-exosomes ease inflammation, and prevent abnormal neurogenesis and memory dysfunction after status epilepticus. *Proc Natl Acad Sci U S A* 2017; 114: E3536-E3545.
- 14) LOU G, SONG X, YANG F, WU S, WANG J, CHEN Z, LIU Y. Exosomes derived from miR-122-modified adipose tissue-derived MSCs increase chemosensitivity of hepatocellular carcinoma. *J Hematol Oncol* 2015; 8: 122.
- 15) NIU M, XU R, WANG J, HOU B, XIE A. MiR-133b ameliorates axon degeneration induced by MPP(+) via targeting RhoA. *Neuroscience* 2016; 325: 39-49.
- 16) POPOVICH PG. Neuroimmunology of traumatic spinal cord injury: a brief history and overview. *Exp Neurol* 2014; 258: 1-4.
- 17) STENUDD M, SABELSTRÖM H, FRISEN J. Role of endogenous neural stem cells in spinal cord injury and repair. *JAMA Neurol* 2015; 72: 235-237.
- 18) THEIS T, YOO M, PARK CS, CHEN J, KUGLER S, GIBBS KM, SCHACHNER M. Lentiviral delivery of miR-133b improves functional recovery after spinal cord injury in mice. *Mol Neurobiol* 2017; 50: 4659-4670.
- 19) UMEZU T, TADOKORO H, AZUMA M, SHIZAWA S, OHTSUKI K, OHYASHIKI JH. Exosomal miR-133b shed from hypoxic multiple myeloma cells enhances angiogenesis by targeting factor-inhibiting of blood 2014; 124: 3748-3757.
- 20) WU X, WALKER CL, LIU M, WU W, EDDERMAN DB, PARISH JM, XU XM. RhoA/RhoGase mediates neuronal death through regulation of PI3K activation. *Mol Neurobiol* 2017; 54: 6885-6893.
- 21) ZHANG Y, CHEN M, MENG Y, KATAKOWSKI M, XIN H, WOOD A, XIONG Y. Effect of exosomes derived from multipotential mesenchymal stromal cells on functional recovery and neurovascular plasticity in rats after traumatic brain injury. *J Neurosurg* 2015; 122: 86-867.