

Long noncoding RNA OR3A4 promotes cisplatin resistance of non-small cell lung cancer by upregulating CDK1

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Abstract. – OBJECTIVE: Numerous studies have proved that long non-coding RNAs (lncRNAs) have an important role in malignant tumors, including non-small cell lung cancer (NSCLC). LncRNA olfactory receptor family 3 subfamily A member 4 (OR3A4) was explored to identify how it functions in resistance of NSCLC patients to cisplatin.

MATERIALS AND METHODS: Real Time-quantitative Polymerase Chain Reaction (RT-qPCR) was utilized to detect OR3A4 expression in NSCLC patients. Then, we conducted Cell Counting Kit-8 (CCK-8) assay and flow cytometric analysis to detect the function of OR3A4 on the resistance of NSCLC cells to cisplatin. Furthermore, the potential mechanism was explored by messenger RNA assays.

RESULTS: Compared with control expression of paired A549 cells, OR3A4 expression of A549/DDP cells was higher. Moreover, the functional assay showed that after OR3A4 was overexpressed in A549/DDP cells, cell cycle arrest and cell apoptosis was induced and resistance to cisplatin was reversed. Furthermore, it was found that CDK1 expression was upregulated in A549/DDP cells by knock down of OR3A4.

CONCLUSIONS: The present work suggests that OR3A4 participates in regulating cell cycle, cell apoptosis of NSCLC cells and the resistance to cisplatin via upregulating CDK1, indicating that OR3A4 could be identified as a potential therapeutic target for NSCLC patients.

Keywords:

lncRNA, OR3A4, NSCLC, CDK1, Cisplatin.

Introduction

More than 80% of all lung cancer cases are non-small cell lung cancer (NSCLC)¹. The 5-year overall survival rate is less than 15% in the NS-

CLC patients². Chemotherapy drugs are emerging as the most important strategy of therapeutic strategy, particularly in NSCLC. The resistance to chemotherapy drugs remains an important factor for the prognosis of NSCLC patients. Therefore, there is an urgent need to identify a novel biomarker and therapeutic target for the patient with NSCLC.

Although long noncoding RNAs (lncRNAs) have little potential of protein-coding, they are important regulators in carcinogenesis of cancers. For example, lncRNA H19 is correlated with resistance to cisplatin in lung adenocarcinoma³. lncRNA HOTAIR induces cisplatin resistance by up-regulating miR-34a in gastric cancer⁴. lncRNA CCAT1 regulates SOX4 and promotes cisplatin resistance in NSCLC cells⁵. lncRNA UCA1 promotes cell growth and cisplatin resistance of oral squamous cell carcinoma via downregulating miR-184⁶. However, the clinical role of olfactory receptor family 3 subfamily A member 4 (OR3A4) in cisplatin-resistance remains unknown.

Therefore, we conducted studies and found that OR3A4 could regulate cell apoptosis, cell cycle and cisplatin resistance in NSCLC. Meanwhile, Cyclin-dependent kinase 1 (CDK1) has been identified as a potential marker in many cancers and participates in many tumors. In this work, we found that CDK1 was upregulated by OR3A4 and is associated with resistance to cisplatin.

Materials and Methods

NSCLC Cell Lines

A549/DDP cells and A549 cells were cultured in Roswell Park Memorial Institute-1640

(RPMI-1640) medium (Life Technologies, Gaithersburg, MD, USA) added with 10% fetal bovine serum (FBS; Life Technologies, Gaithersburg, MD, USA). Besides, the incubator for cell culture consisted of 5% CO₂ at 37°C.

Cell Transfection

The complementary deoxyribose nucleic acid (cDNA) oligonucleotides specifically targeting OR3A4 (OR3A4/shRNA) was synthesized by GenePharma (Shanghai, China), and cloned into pGPH1/Neo. Then, OR3A4/shRNA were used for transfection in NSCLC cells. 48 h later, Real Time-quantitative Polymerase Chain Reaction (RT-qPCR) was used to measure the transfection efficiency.

RNA Extraction and Real Time-Quantitative Polymerase Chain Reaction

TRIzol reagent (Invitrogen, Carlsbad, CA, USA) was utilized for extracting total RNA, which was then reversely transcribed to cDNA through reverse Transcription Kit (TaKaRa Biotechnology, Otsu, Shiga, Japan). Following are the primers used for RT-qPCR: OR3A4, forward 5'-CCTATCCCTTCTCTAAGAA-3' and reverse 5'-ACTTCTGCAAAAACGTGC-3'; and for glyceraldehyde 3-phosphate dehydrogenase (GAPDH), forward 5'-CCACATCGCAGACACCAT-3' and reverse 5'-CCAGGATGCCCCAATACG-3'. The thermal cycle was as follows: 95°C for 30 sec, 1 min for 40 cycles at 95°C, 60°C for 35 sec.

Cell Counting Kit-8 (CCK8) Assay

0, 1, 5, 8, 10, 15, 18, 20, 22 or 25 μg/mL cisplatin were used to treat the cells in cell status, respectively. Then, cell viability was detected by Cell Counting Kit-8 (Dojindo Molecular Technologies, Kumamoto, Japan) at 0, 48, 72 and 96 h. The absorbance was measured at 450 nm.

Cell Cycle Analysis

2 × 10⁵ cells were diluted by RNase A in ice-cold PBS overnight, and these cells were stained with Propidium Iodide (PI; 50 mg/mL; Beyotime, Shanghai, China) in the dark for 2 h. Then, they were measured with flow cytometer (FACScan, BD Biosciences, Franklin Lakes, NJ, USA).

Cell Apoptosis Analysis

Harvested cells were washed twice using ice-cold. Then 100 μL of flow cytometry binding

buffer was added. After Annexin V/fluorescein isothiocyanate (FITC; 5 μL; Beyotime, Shanghai, China) and PI (5 μL) were mixed, these cells were stained for 15 min in the dark. Each tube was added with four hundred microliters binding buffer. FACSCalibur flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) was performed to analyze cell apoptosis.

Western Blotting Analysis

After being separated with 12% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE), the gel was replaced to polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA). Rabbit anti-CDK1 (Abcam, Cambridge, MA, USA) and anti-GAPDH (Abcam, Cambridge, MA, USA) were used as the primary antibodies, which were utilized by incubating the membranes overnight. Goat anti-rabbit secondary antibody (Abcam, Cambridge, MA, USA) was then applied for incubation. Image software (NIH, Bethesda, MD, USA) was performed for data analysis.

Statistical Analysis

Statistical Product and Service Solutions 17.0 (SPSS, Chicago, IL, USA) was utilized to perform statistical analysis. The method of 2^{-ΔΔCT} and two-tailed Student's *t*-test were used. It was considered statistically significant when *p* < 0.05.

Results

The Expression of OR3A4 in A549/DDP and A549 Cells

As half of the maximal inhibitory concentration (IC₅₀) of cisplatin was an important factor in resistance to cisplatin, it was detected through the CCK-8 assay in NSCLC cells. As a result, the IC₅₀ of cisplatin in A549/DDP cells was remarkably increased compared with A549 cells (Figure 1A). Furthermore, OR3A4 was lower-expressed in A549 cells than A549/DDP cell line (Figure 1B).

OR3A4 Was Upregulated in A549 Cells Treated With Cisplatin

Then, A549 cells were treated with 0.0, 0.5, 1.0, 1.5, 2.0 or 2.5 μg/ml cisplatin. RT-qPCR was utilized to monitor OR3A4 expression in these treated cells. After cisplatin concentrations were increased, the OR3A4 expression of these treated cells was increased (Figure 2).

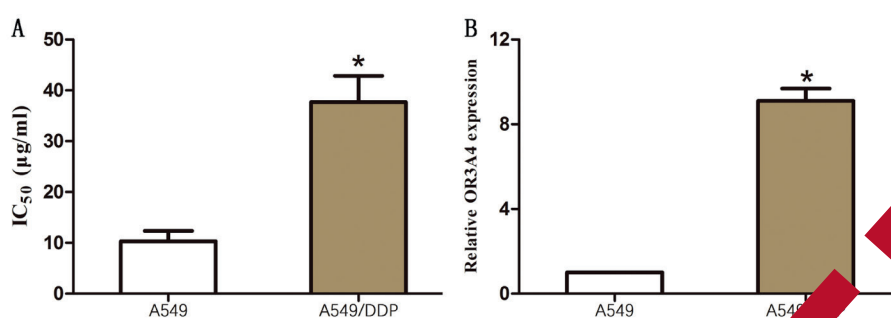


Figure 1. The expression levels of OR3A4 were increased in A549/DDP cells. **A**, IC_{50} value of cisplatin was higher in A549/DDP cells, compared with that of the A549 cells. **B**, The expression levels of OR3A4 relative to GAPDH were determined in the A549/DDP and A549 cells by RT-qPCR. GAPDH was used as an internal control. * $p < 0.05$.

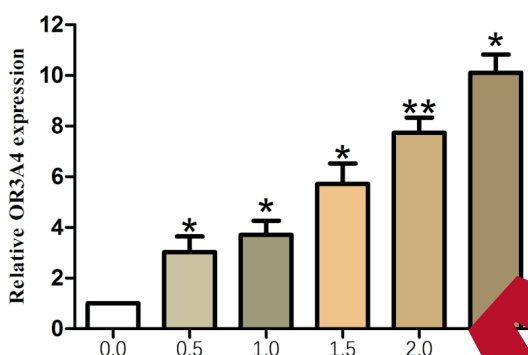


Figure 2. OR3A4 was upregulated in A549 cells treated with cisplatin. A549 cells were cultured in various concentrations of cisplatin (0.0, 0.5, 1.0, 1.5, 2.0, 2.5 μg/mL) for 24 h. OR3A4 expression was evaluated by RT-qPCR. GAPDH was used as an internal control. * $p < 0.05$, ** $p < 0.01$.

Knockdown of OR3A4 Reduced Cisplatin Resistance in A549/DDP Cells

After A549/DDP cells were silenced through OR3A4/shRNA, the infection efficiency was

monitored by RT-qPCR (Figure 3A). We conducted the CCK-8 assay and found that IC_{50} of cisplatin was decreased through knockdown of OR3A4 in A549/DDP cells (Figure 3B). Moreover, after these cells were treated with cisplatin (0.5, 1.0 or 2.0 μg/ml), cell apoptosis of OR3A4/shRNA group was promoted compared with the control group (Figure 4A). In addition, these treated cells were in subG0/G1 phases in the OR3A4/shRNA group was increased compared with that in the control group (Figure 4B).

Knockdown of OR3A4 Decreased Cisplatin Resistance in A549/DDP Cells by Downregulating CDK1

RT-qPCR results revealed that CDK1 of A549/DDP cells was remarkably lower-expressed in the OR3A4/shRNA group compared with that in the control group (Figure 5A). Western blot analysis results also revealed that CDK1 of

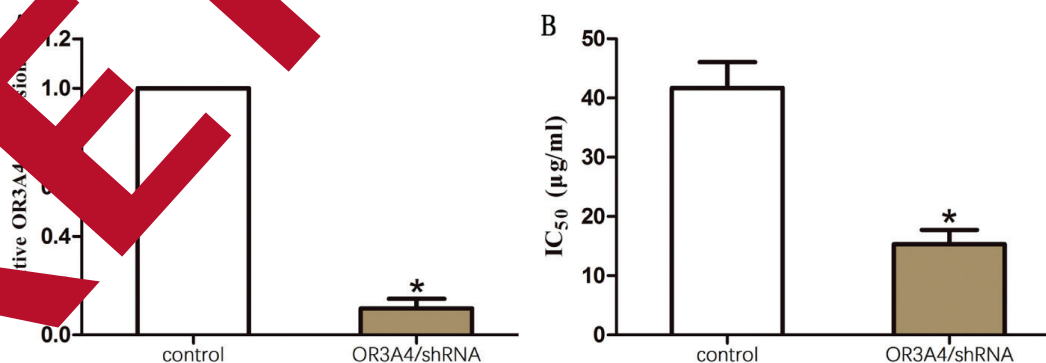


Figure 3. The knockdown of OR3A4 reduced IC_{50} values of cisplatin in A549/DDP. **A**, 48 h after A549/DDP cells were transfected with OR3A4/shRNA, the inhibition efficiency was detected by RT-qPCR. GAPDH was used as an internal control. **B**, IC_{50} values of cisplatin in A549/DDP cells transfected with control and OR3A4/shRNA were analyzed by CCK-8 assay. The results represent the average of three independent experiments. * $p < 0.05$.

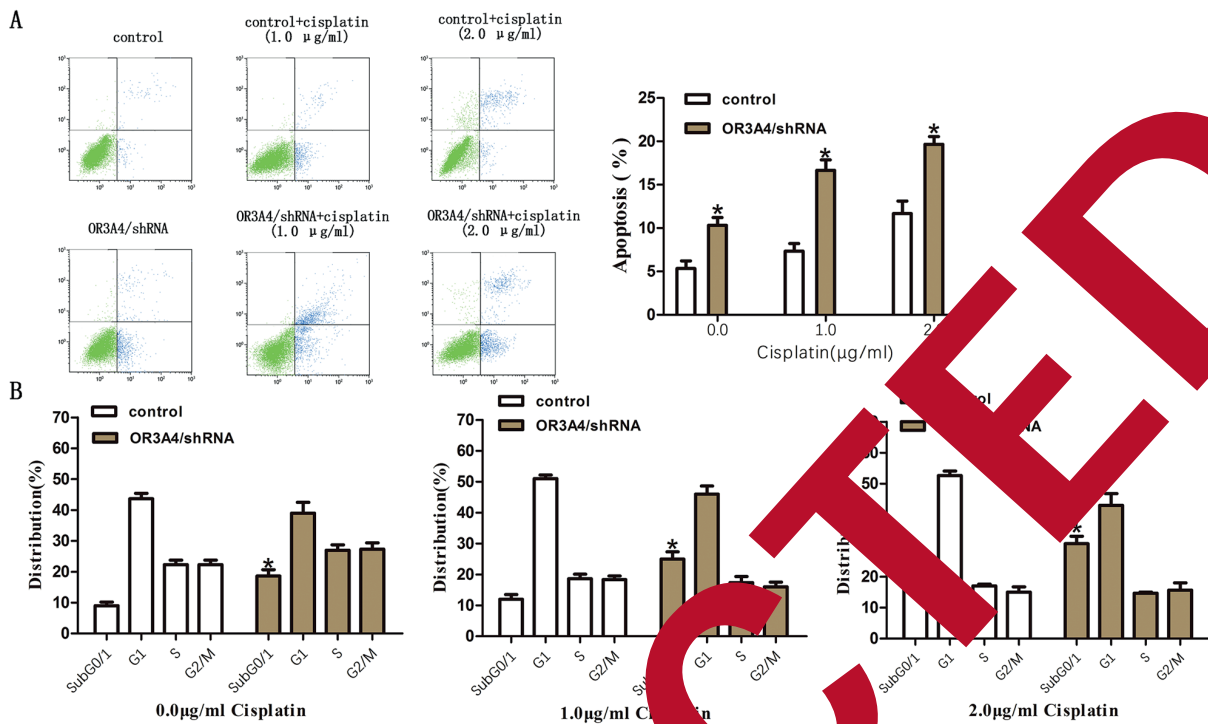


Figure 4. Knockdown of OR3A4 promoted the sensitivity of A549/DDP cells to cisplatin. **A**, Cell apoptosis induced by different concentrations (0.0, 1.0 or 2.0 µg/mL) of cisplatin was detected in A549/DDP cells transfected with control and OR3A4/shRNA by flow cytometric analysis. **B**, Cell cycle distribution induced by different concentrations (0.0, 1.0 or 2.0 µg/mL) of cisplatin was detected in A549/DDP cells transfected with control and OR3A4/shRNA by flow cytometric analysis. The results represent the average of three independent experiments. * $p < 0.05$.

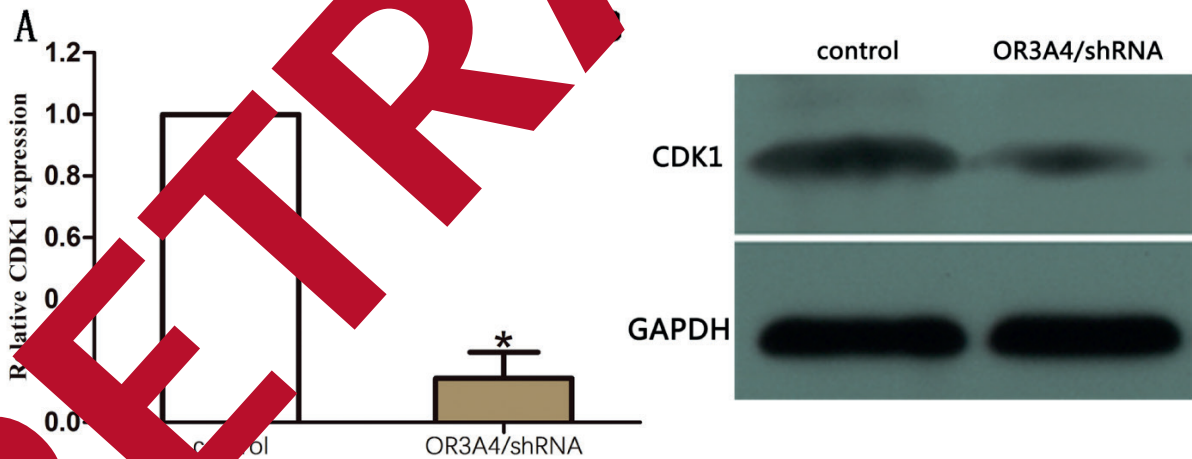


Figure 5. Knockdown of OR3A4 downregulated CDK1 expression in A549/DDP cells. **A**, The mRNA expression levels of CDK1 in A549/DDP cells transfected with control and OR3A4/shRNA were analyzed by RT-qPCR. **B**, Protein expression levels of CDK1 in A549/DDP cells transfected with control and OR3A4/shRNA were analyzed by Western blot assay. GAPDH was used as a control. * $p < 0.05$.

A549/DDP cells was markedly downregulated in the OR3A4/shRNA group compared with that in the control group (Figure 5B).

Discussion

The resistance to chemotherapy drugs remains a vital factor of patients' prognosis de-

spite the fact that various chemotherapy drugs are available for lung cancers^{7,8}. Recently, many studies have been conducted to explain the mechanism of cisplatin resistance, among which the regulation of cell apoptosis is a vital progression in drug resistance for cancers. Moreover, lncRNAs could modulate cell apoptosis and further regulate drug resistance in cancers⁹⁻¹².

Recent studies¹³ revealed that olfactory receptor family 3 subfamily A member 4 (OR3A4) is abnormally expressed and participates in the progression of many cancers. For instance, OR3A4 promotes cell proliferation and acts as an oncogene in gastric cancer, and cell growth ability and invaded ability are promoted in breast cancer by upregulating OR3A4¹⁴. However, the role of OR3A4 in resistance to cisplatin in cancers has not been studied so far. In this work, OR3A4 was upregulated in A549/DDP cells compared with A549 cells. Besides, the OR3A4 was upregulated after the dose of cisplatin for treating A549 cells increased. In addition, cisplatin-induced apoptosis of A549/DDP cells was promoted *via* knockdown of OR3A4. The percent of A549/DDP cells in subG0/G1 phases was increased in OR3A4/shRNA group after treating with different doses of cisplatin.

Cyclin-dependent kinase 1 (CDK1) has been identified as potential marker in many cancers. For example, CDK1 expression is correlated to clinical stage and treatment efficacy of breast cancer¹³. CDK1 is high-expressed in lung cancer and inhibits cell apoptosis *via* p53 apoptosis pathway¹⁴. CDK1 protein is associated with cell growth and cell apoptosis in ovarian cancer by regulating Cdk1-Cyclin B and P53-P21WAF1 signaling pathway¹⁵. CDK1 has been proved to function in the regulation of cell cycle induced by cisplatin¹⁶. Our paper showed that CDK1 was down-regulated in A549/DDP cells *via* knockdown of OR3A4.

Conclusions

We observed that OR3A4 regulated cell apoptosis, cell cycle, and enhanced the resistance of NSCLC cells to cisplatin *via* upregulating CDK1. These findings imply that OR3A4 can be served as a promising mark for NSCLC.

Conflict of interest

The authors declare no conflicts of interest.

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