

Property and current clinical applications of mammal hyaluronidase

S.-H. LV¹, S.-F. RONG², B.-G. CAI², S.-M. GUAN², Q.-O. LI²

¹Master of Biochemistry, Department of Bioengineering, Shanghai Institute of Technology, China

²Department of Bioengineering, Shanghai Institute of Technology, China

Abstract. – Hyaluronidase (Hyal), which is related to mammalian diseases, is greatly significant for mammal. It is the major enzyme for degrading hyaluronan (HA), which is a linear high molecular polymer that is ubiquitous in mammalian extracellular matrix. Previous studies suggested that the levels of Hyals play significant roles in predicting, determining and curing many diseases.

This review summarizes previous studies on the classification and biophysical and therapeutic applications of mammalian Hyals and focuses on the current medicinal and clinical applications of Hyals.

Key Words:

Mammal hyaluronidase, Medicinal and clinical applications, Tumor cells.

Introduction

Hyaluronidases (Hyals) are endo- β -N-acetylhexosaminidases (EC 3.2.1.35) that predominantly hydrolyze hyaluronan (hyaluronic acid or hyaluronate) and catalyze transglycosylation. They have limited ability to degrade chondroitin sulphate (C4-S, C6-S)¹⁻³. Hyaluronan (HA) is a high-molecular-weight, unsulfated glycosaminoglycan that comprises repeating units of [N-acetylglucosamine (1- β -4) D-glucuronic acid (1- β -3)]_n (Figure 1). HA dominates the macromolecular components of the extracellular matrix (ECM) in mammals; these components include connective tissues, eyes, skin, cartilage, aorta, synovial fluid and splanchnic nerves⁴. Mammalian Hyals can overcome the physical barrier that is formed by HA, and change the penetrability of ECM. They are ubiquitous in stroma and tissues from the liver, brain, testicle, plasma, and ECM⁵. Mammalian Hyals have attracted increasing attention because of their functions in several physiological and pathological reactions, such as

fertilization⁶, embryogenesis⁷, wound healing⁸, tumor growth⁹ and tumor malignancy¹⁰. This review summarized the bio-physical and chemical properties of mammal Hyal as well as the medicinal and clinical applications of mammalian Hyals.

Mammalian Hyaluronidases

Mammalian Hyals are hyaluronan 4-glycanohydrolases, which degrade HA by cleaving the β -1,4-glycosidic bond and produce a tetrasaccharide molecule as their main product. They involve both hydrolytic and transglycosidase activities¹¹. The degradation of mammalian Hyals follows a random biting-type catalytic method¹², which is an endo, non-progressive mechanism, in degrading products and substrates.

Genomic analysis reveals that mammals possess multiple Hyal genes. In humans, the *Hyal* (*hHyal*) -1, *hHyal*-2 and *hHyal*-3 genes are tightly clustered on chromosome 3p21.3 whereas the *hHyal*-4, *hHYALP1* (hyaluronidase pseudogene gene 1) and *hPH*-20 are clustered on chromosome 7q31.3⁵. *HYALP1* is expressed as a pseudogene in humans¹³ but encoded as an active enzyme in mice¹⁴. Meningioma expressed antigen-5 (MGEA5) is the eighth hyaluronidase-like protein coded for by *Mgea5* on chromosome 10q24.1-q24.3. This protein demonstrated a hyaluronidase activity and is identified as a novel immunogenic antigen that is expressed in meningiomas¹⁵. In mice, *Hyal*-1, *Hyal*-2 and *Hyal*-3 are located on chromosome 9F1, whereas *Hyal*-4, *HYALP1*, *PH*-20 and *Hyal*-5 are located on chromosome 6A¹⁶. However, the existence of Hyals provides vast potential for both constitutive and regulated turnover of HA^{12,17}. All mammalian Hyals have different substrate specificities and expression profiles. The properties of these enzymes are listed in Table I.

Hyal-1 is a secreted Hyal that serves as the major type of Hyals in human plasma and is

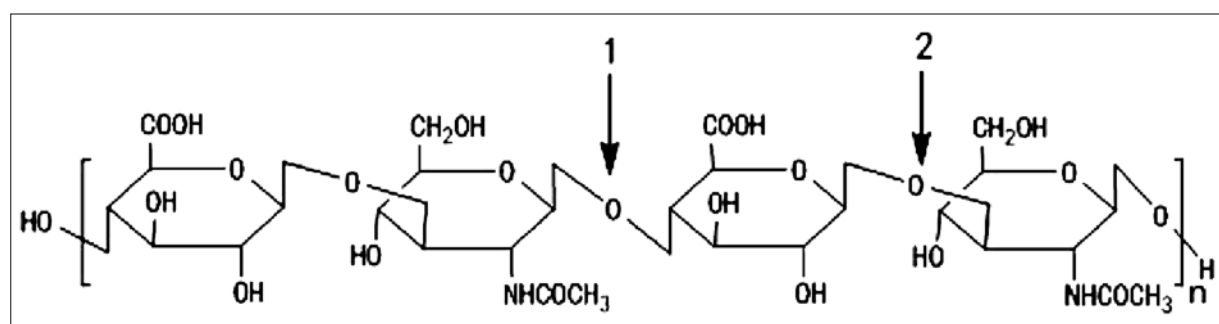


Figure 1. Structure of Hyaluronan. 1, β -1, 3 glycosidic bond; 2, β -1, 4 glycosidic bond.

ubiquitous in somatic tissues. Hyal-1 is responsible for the catabolism of HA in ECM. It is an acid pH optimum enzyme¹¹. Hyal-1 is associated with many diseases, such as tumor growth, invasion and the deterioration. A deficiency in Hyal-1 causes a lysosomal storage disorder in humans called mucopolysaccharidosis IX²⁶. The transformation of Hyal-1 in tissues is involved in the healing of damaged tissue and wound²⁷.

Hyal-2 is also found in most tissues, except in the adult brain. Hyal-2 has a lower activity than Hyal-1 and PH-20^{20,28}. It is localized either in lysosome or is connected to the vascular membrane surface as a glycosphosphatidylinositol (GPI)-anchored cell surface protein²⁹. Hyal-2 also serves as the cell-entry receptor for Jaagsiekte sheep retrovirus which is associated with lung cancer³⁰. Hyal-2 is also an acid pH optimum enzyme. It degrades high molecular weight HA into 20kDa HA fragment, which corresponds to 50-

60 disaccharide units²⁰. This HA fragment has distinctive biological function. In specific Hyal-2 can stimulate angiogenesis, which facilitates the growth of solid tumors²⁸. However, the special function of Hyal-2 in mammals needs further investigation.

Hyal-3 is relatively unexplored as compared with other Hyals. This protein is highly expressed in testis and bone marrow, but is weakly expressed in other organs^{5,31}. Hyal-3 is also expressed in the adult brain, in which Hyal-1 and Hyal-2 are rarely found. A functional study showed that mice without Hyal-3 were viable and fertile, exhibited no gross phenotypic change and showed no indication of HA accumulation³². Hyal-3 may not play a major role in HA degradation at least in mouse³².

Unlike other Hyals, human hyal-4 is a chondroitin sulfate-specific endo- β -N-acetylgalactosaminidase that exerts no hyaluronidase ac-

Table I. Properties of Hyaluronidases.

Type	Expressed sites	Molecular weight	Optima pH	Substrate	Product	Reference
Hyal-1	Widely expressed, (plasma) intracellular	435AA	3.0 -4.5	HA Ch/CS	–	18
Hyal -2	Widely expressed, extracellular	452AA	6.0-7.0	HA (Only high molecular weight HA)	20kDa oligosaccharides	19, 20
Hyal -3	Testis, bone marrow, weak expression in other organs	45.0-56.0kDa	–	HA	–	21
Hyal -4	Restricted to placenta, skeletal muscle, testis	54.0kDa (481AA)	5.0	Ch	–	5, 22
HYALP1	All organs, an expressed pseudogene	–	–	–	–	5
PH-20	Testis extracellular	80.0-97.0 kDa 30-59kDa	4.0, 7.0	HA Ch/CS	–	23, 24
Hyal -5	Testis	–	–	HA	–	25
MGEA5	–	–	–	HA	–	15

tivity³. Hyal-4 is also a GPI-anchored chondroitinase^{3,22}. HYALP1 has been detected translated in many human tissues, but does appear to be active. HYALP1 has a unique function in the adult testis and may compensate for the function of SPAM1 in *Spam1* null mice¹³.

PH-20 (Sperm adhesion molecule 1, SPAM1 or PH-20) is the most active mammalian Hyal. It degrades HA into small fragments, mostly tetrasaccharides. PH-20 is a GPI-anchored protein only located on the plasma membrane of sperm. PH-20 is the major sperm Hyal and plays an important role in sperm adhesion to the oocyte zona pellucida and in sperm penetration through the cumulus matrix³³. Sperm without PH-20 were present in the cumulus matrix at a low rate and the numbers of sperm were lower than normal at the early stages after insemination *in vitro*³⁴. However, these proteins still had the ability to reach the oocyte zona pellucida and undergo fertilization³³. PH-20 may not essential in fertilization in mouse³⁵, but it is still has an exact effect on fertilization.

Hyal-5 is also a GPI-anchored Hyal that contains epididymal sperm that can only be found in mouse testes. This protein helps PH-20 degrade the HA-rich matrix of the cumulus layer that surround the egg³³. The lack of Hyal-5 in mice did not find significantly effect on fertilization. Therefore, Hyal-5 may be considered dispensable for fertilization³³. MGEA5 is the eighth hyaluronidase-like protein coded by *Mgea5* on chromosome 10q24.1-q24.3. MGEA5 is a novel immunogenic antigen that is expressed in meningiomas and demonstrated Hyal activity¹⁵.

Homology Analysis

Mammalian Hyals are widely distributed in mammals. Similarities in the amino acids sequences of mammalian Hyal were analyzed using DNAMAN Version 4.0 (Lynnon LLC., San Ramon, CA, USA). High similarity rates of 83.86%, 88.52%, 88.07%, 66.76%, 90.92%, and 91.72% were observed in Hyal-1, Hyal-2, Hyal-3, Hyal-4, PH-20, and MGEA5, respectively.

Phylogenetic analysis was performed using MEGA6.0. The amino acids sequences of the mammalian Hyals from *Homo sapiens*, *Rattus norvegicus*, *Mus musculus*, *Bos taurus*, *Sus scrofa* and *Pan troglodytes* were chosen to construct a phylogenetic tree. The phylogenetic analysis results in Figure 2 are consistent with the Hyal classification. Mammalian Hyals were divided into seven subgroups. MGEA5 achieved the low-

est similarity with the six other mammalian Hyals. Hyal-1, Hyal-2 and Hyal-3, which were clustered on the same chromosome, had a near-similar homology. PH-20 and Hyal-5, which were expressed in the sperm, also had a near-similar homology. Hyal-4 had a higher similarity with PH-20 and Hyal-5 than with Hyal-1, Hyal-2 and Hyal-3. These subgroups of Hyals were closely associated with their locations on chromosomes and their functions. This homology indicated an ancient gene duplication, which could motivate future studies on the different Hyal subtypes with unknown functions.

Function of Mammalian Hyaluronidases in Cancer Biology

The effect of the ECM degrading enzyme Hyals on interstitial fluid pressure and microvascular pressure was measured in many human tumors³⁶. The levels of Hyals and HA are closely associated with many types of tumor cell lines^{37,38}. The Hyals that are expressed by tumor cells degrade HA to small HA fragments of 2 to 15 disaccharide units which can stimulate angiogenesis, and initiate endothelial cell proliferation via the cell surface receptors (RHAMM and CD44)³⁹. HA-CD44 interactions are important in malignant tumor growth angiogenesis induction and tumor progression *in vivo*¹⁰. Blocking Hyal-1 expression in invasive bladder cancer (BCa) cells decreases tumor growth, inhibits tumor infiltration, and decreases microvessel density²⁹. Adding Hyal-1 and Hyal-2 in a culture dose not inhibit tumor growth *in vitro*. The overexpression of Hyal-2 not only increases the growth rate of tumor cells but also enhances the invasiveness and vascularization in intracerebral glioma³⁷.

Some studies suggested that Hyals exhibited a very strong inhibiting effect^{10,40}. In human genomes, Hyals genes clustered on chromosome 3p21.2 were considered as conserved candidate tumor suppressors¹⁶. Wang et al⁴⁰ found that Hyal-2 showed some tumor suppressor activity in SCID mice. Chang et al⁴¹ reported that Hyal-1 and Hyal-2 increased the susceptibility of tumor necrosis factor in L929 cells. This study proved that the transforming growth factor β 1 only promoted the growth of L929 cells but not the growth of Hyal-2 and Hyal-2-expressing L929 cells⁴². The use of Hyals can enhance the action of immune effector cells via degradation of the HA-formed halos around cancer cells that look similar to adherent fibrosarcoma cells⁴³.

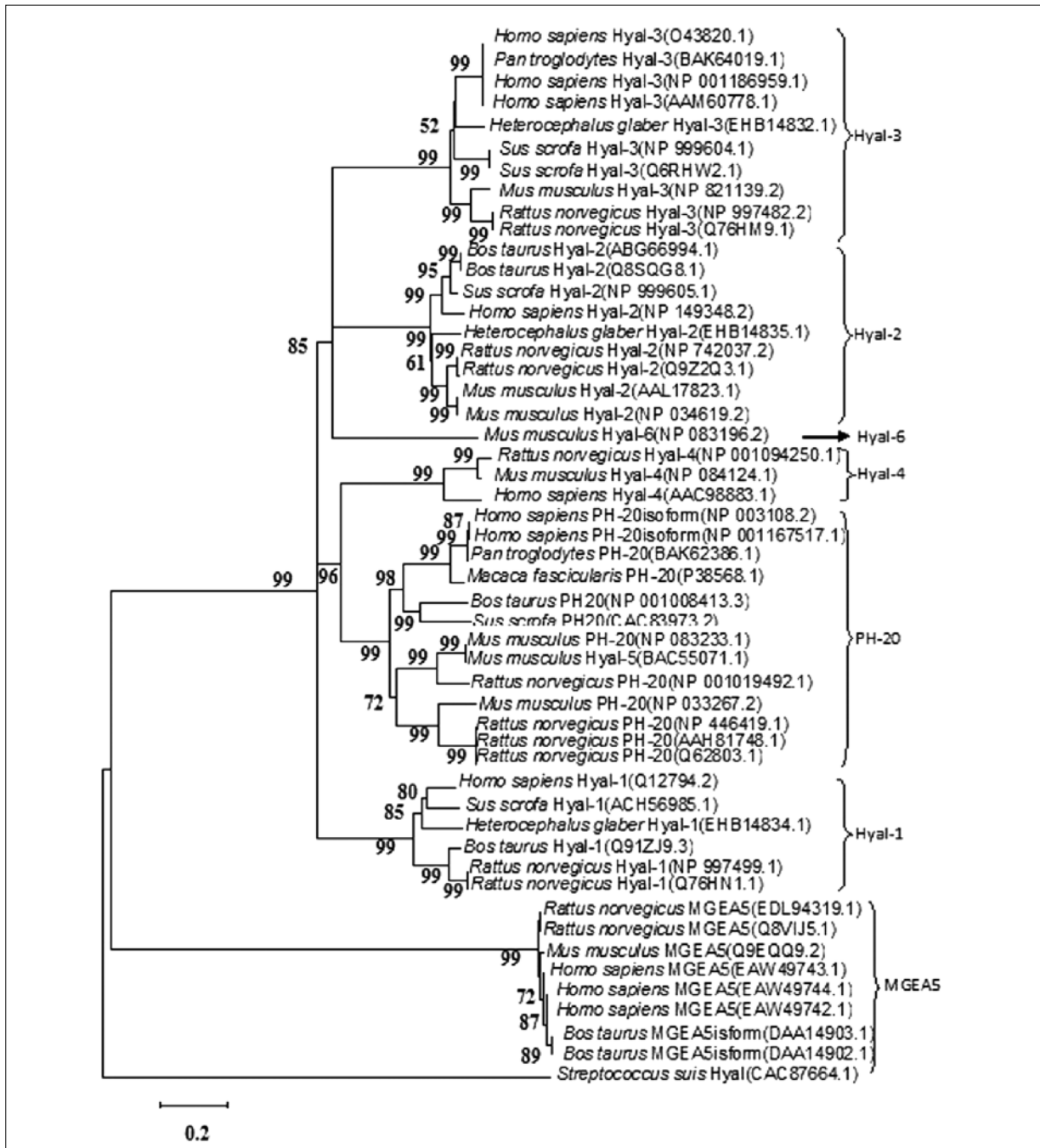


Figure 2. Phylogenetic trees showing the affiliations of Hyal amino acid sequences (GenBank number). The tree was constructed by neighbor-joining using full-length aligned amino acids sequences with *Streptococcus suis* as an outgroup. The bootstrap value is based on 1000 replicates and is shown at the nodes with more than 50% bootstrap support. The scale bar represents 20% sequence divergence.

However, some studies suggested that Hyals exhibited both inhibiting and facilitating effects on cancer. This finding is closely related to the cellular concentration of Hyals¹⁸. Hyal variants were also found in tumor cells. For example, Hyal-1 variants were found in tumor cells of BCa⁴⁴. Eleven types of

Hyal-wild (1, 2, 3) or variants were found in adenocarcinomas and squamous cell⁴⁵. Previous studies concluded that the function of Hyals in normal cells and their variants in tumor cells remains unclear.

Although the function of Hyals in cancers has yet been clarified, they can be used as markers

and therapeutic adjuncts in cancer. Hyals can also be used as a non-invasion method for determining cancer prognosis. Eissa et al⁴⁶ showed that the measuring Hyal-1 mRNA by semiquantitative PCR can be performed to diagnoses BCa. Moreover, Hyal achieves high accuracy in detecting cancer and evaluating its grade⁴⁶. Hyals can also serve as markers to accurately detect early and high-grade BCa as well as to predict prostate cancer progression. Virotherapy, studies suggested that oncolytic adenoviruses that expressing PH-20 have great antitumor efficacy after systemic administration in virotherapy for cancer. The co-application of Hyals with oncolytic adenovirus can enhance the anti-cancer activity^{47, 48}. When chemically treatment of malignant tumors, chemotherapy drugs that are leaked into the surrounding tissue or intravenous injection of such drugs into the extravasation accident can damage tissues, nerve and tendons. Hyals can reduce intratumoral pressure, which allows the drug to penetration into the malignant tumor and relieve the chemical drug out of vessels⁴⁹.

Clinical Application and Prospect of Mammalian Hyaluronidases

The biological function and therapeutic applications of Hyals have recently attracted considerable academic attention. Hyals have potential application in postoperative operation, giving of macromolecular drugs, trauma treatment and wound healing⁵⁰.

Hyaluronidases as Adjuvant

Mammalian Hyals have been widely used as adjuvants in clinical settings to administer local anesthetics^{51,52}, replacement fluid⁵³, insulin⁵⁴, corticosteroid and large molecules⁵⁵. Hyal can reduce the viscosity of ECM. A lesser viscosity of ECM facilitates the diffusion of the anesthetic agent. Adding Hyals with lidocaine can enlarge the anesthetized area, shorten the drug diffusion time, increase the permeability of tissues, and enhance the anesthesia effect⁵⁶. Therefore, the use of Hyals in cardiac patients is beneficial because they strengthen the anesthetic effect and decrease the volume of applied anesthetic. Hyals can also effectively improve the absorption of X-ray opaque at drugs. Injecting Hyals has also been considered safe agent without any combined applications⁵¹. Recombinant human Hyals (rHuHyals) has advantage of high purity, favorable efficacy and safety⁵⁷. Hyals may be suitable alternatives for clinical use than compounded animal-derived products⁵⁸.

Hyaluronidases in Ophthalmology

Hyals have been used in ophthalmology for many years⁵⁹. Hyals can be injected with anaesthetic for retrobulbar, peribulbar and sub-Tenon's blocks⁵⁹⁻⁶¹. They can promote the spread of the anaesthetic (bupivacaine or lidocaine or ropivacaine et al⁶²) through the tissues or (and) reduce the volumes of local anaesthesia⁶⁰. It was found that Hyals can be used for pharmacologic vitreolysis to avoid the complication of surgery such as cataract, endophthalmitis and anesthesia related complications⁶³. The advantages of using Hyal also include enhancing speed of onset and improving operating conditions⁶⁴.

Previous studies^{58,62,65} indicated that the purity and optimal dosage of Hyals were crucial in ophthalmology. Important factors that influence the clinic use of Hyals still need investigation, such as injection site and techniques, the choice of anaesthetic solution and the optimal dosage^{58,62}. In addition, the toxicity of Hyals still needs further studies^{61,66,67}.

Hyaluronidases in cancer forecast and Therapy

Elevated levels of Hyals and HA in plasma and urine have been reported to be associated with the presence of tumors, like BCa⁶⁸, epithelial ovarian cancer³⁸, lung cancer, prostate cancer, and breast cancer⁶⁹. Therefore, Hyals may have potential advantages for tumor diagnosis⁴⁶. Jamshldlan et al⁷⁰ devised a highly accurate and non-invasive cancer diagnosis method that involves measuring the urinary levels of HA and Hyals and evaluating its grade in BCa. Hyal-1 is expressed in bladder cancer lines and serves as an accurate marker for high-level bladder cancer.

Hyal can be used to treat intravenous extravasations. They can also be used as antidote to the extravasation of vinca alkaloids; this condition results from the complication of severe tissue necrosis during administration⁴⁹. The absorption of Hyals can also prevent local necrosis from exuding further. The subcutaneous administration of Ondansetron with rHuHyals is generally tolerated in minipigs and healthy volunteers⁷¹.

Hyaluronidases in Plastic Surgery

Hyals are commonly applied in plastic surgery to reverse the effects of fillers overdose, eliminate nodules, and reduce the odds of ischemic complications. Hyals can also reduce skin damage and prevent postoperative wounds from swelling⁷².

HA fillers are widely used in plastic surgery. However, these fillers can cause several complications that range from superficial placement, uneven placement, granulomatous reactions, and skin necrosis⁷³. Hyals can effectively solve such problems. The rapid response of Hyals is particularly helpful in compromise skin malposition or overfill. Given their safety and effectiveness, Hyals are generally used as HA filler to prevent severe complications⁷⁴. Several experiments have proven that infiltration of Hyals can rapidly, effectively, and safely solve eyelid edema following the injections of HA fillers⁵⁸. Cavallini et al⁷⁵ reported that ovine Hyal dosages below 14U did not affect fibroblast viability.

Hyaluronidases in Fertilization

PH-20 is the major type of Hyal in testis that significantly increases the ability of null sperm to penetrate the cumulus of oocytes. The relation between PH-20 and fertilizing ability indicates that PH-20 is a marker of sperm maturation⁷⁶. Although some experiments have shown that PH-20 null mice are still fertile, their fertilization efficiency has significantly decreased^{34,77}. Besides from these functions, PH-20 also has high contraceptive properties because of its strong immunogenicity and essential role in fertilization⁷⁸. Hyal-1 and Hyal-3 also play important roles in ovarian folliculogenesis and female fertility⁷⁹.

Hyaluronidases in Immunoreaction

Combining human immunoglobulin with rHuPH-20 offers a new method for subcutaneous delivery of human immunoglobulin replacement therapy in patients with primary immunodeficiency disorders. rHuPH-20 increases the volumes of infused subcutaneous immunoglobulin and facilitates the absorption immunoglobulin in the lymphatic, particularly in pediatric population^{80,81}. This method is equally effective in the administration of intravenous immunoglobulin and induces fewer complications⁵⁵. This novel method of delivering immunoglobulin is considered safe, effective and well tolerated by patients. It still needs more data to declare the long-term safety⁵⁵.

Potential Functions

Hyals have potential applications in the DNA vaccine. Hyals can also improve conversion efficiency and promote antibody expression during skeletal muscle inflammation treatment among mice via DNA vaccine. Previous studies have al-

so indicated that Hyal improved the initial complement of interleukin (IL)-1 β expression, cancer cell necrosis factor α , interleukin-6 cytokine in inflammatory cells, thereby enhancing the power transfer effect.

Conclusions

Mammalian Hyals has a lots of important physiological processes. However, compared with the Hyals from bacteria or venom, those from mammals remain unexplored in terms of their structure and metabolic process in HA function⁸². Understanding the mechanism of Hyals in cancer progression may be useful for future cancer prognosis and may offer novel pharmacological opportunities for cancer treatment. A guideline must also be devised for using Hyals in clinical application, particularly on their optimal dosage and treatment⁸³. Future work on mammalian Hyals and their clinical use must: (1) elucidate the relationship between structure and function, (2) reveal the mechanism of Hyals in related diseases, (3) develop genetic methods for the rapid production of safe and long-term Hyals, (4) establish the assay of using Hyals and the appropriate dosage.

Acknowledgements

This paper was funded by Shanghai Municipal Commission of Economy and Informatization, grant number CXY-2013-78; Science and Technology Commission of Shanghai Municipality, grant number 11DZ1921405; National Natural Science Foundation of China, grant Number 31301475; Shanghai Institute of Technology, grant number YJ2013-23.

Conflict of Interest

The Authors declare that there are no conflicts of interest.

References

- 1) KOBAYASHI S, FUJIKAWA S, OHMAE M. Enzymatic synthesis of chondroitin and its derivatives catalyzed by hyaluronidase. *J Am Chem Soc* 2003; 125: 14357-14369.
- 2) GUSHULAK L, HEMMING R, MARTIN D, SEYRANTEPE V, PSHEZHETSKY A, TRIGGS-RAINE B. Hyaluronidase 1 and β -hexosaminidase have redundant functions in hyaluronan and chondroitin sulfate degradation. *J Biol Chem* 2012; 287: 16689-16697.
- 3) KANEIWA T, MIZUMOTO S, SUGAHARA K, YAMADA S. Identification of human hyaluronidase-4 as a novel chondroitin sulfate hydrolase that preferentially

- cleaves the galactosaminidic linkage in the trisulfated tetrasaccharide sequence. *Glycobiology* 2010; 20: 300-309.
- 4) NECAS J, BARTOSIKOVA L, BRAUNER P, KOLAR J. Hyaluronic acid (hyaluronan) a review. *Vet Med-Czech* 2008; 53: 397-411.
 - 5) CSÓKA AB, SCHERER SW, STERN R. Expression analysis of six paralogous human hyaluronidase genes clustered on chromosomes 3p21 and 7q31. *Genomics* 1999; 60: 356-361.
 - 6) BROWN CR. Distribution of hyaluronidase in the ram apermatozoon. *J Reprod Fertil* 1975; 45: 537-546.
 - 7) HYDE CE, OLD RW. Expression pattern of a novel hyaluronidase during *Xenopus* embryogenesis. *Mech Develop* 1999; 82: 213-217.
 - 8) YUFA Y, KASAHIMA Y, KUWANO A, SATO K, HATTORI S, ARAI K. Active hyaluronidase 2 expression in the granulation tissue formed in the healing process of equine superficial digital flexor tendonitis. *J Vet Med Sci* 2012; 75: 219-223.
 - 9) TAN J, WANG X, LI H, SU X, WANG L, RAN L, ZHENG K, REN G. HYAL1 overexpression is correlated with the malignant behavior of human breast cancer. *Int J Cancer* 2011; 128: 1303-1315.
 - 10) STERN R. Hyaluronidases in cancer biology. *Semin Cancer Biol* 2008; 18: 7-14.
 - 11) HOFINGER ES, HOECHSTETTER J, OETTL M, BERNHARDT G, BUSCHAUER A. Isoenzyme-specific differences in the degradation of hyaluronic acid by mammalian-type hyaluronidases. *Glycoconj J* 2008; 25: 101-109.
 - 12) MENZEL EJ, FARR C. Hyaluronidase and its substrate hyaluronan biochemistry, biological activities and therapeutic uses. *Cancer Lett* 1998; 131: 3-11.
 - 13) MILLER AK, SHAO M, MARTIN-DELEON AP. Hyalp1 in murine sperm function: evidence for unique and overlapping functions with other reproductive hyaluronidases. *J Androl* 2007; 28: 67-76.
 - 14) CSOKA AB, FROST GI, STERN R. The six hyaluronidase-like genes in the human and mouse genomes. *Matrix Biol* 2001; 20: 499-508.
 - 15) COMTESSE N, MALDENER E, MEESE E. Identification of a nuclear variant of MGEA5, a cytoplasmic hyaluronidase and a β -N-acetylglucosaminidase. *Biochem Biophys Res Commun* 2001; 283: 634-640.
 - 16) CSÓKA TB, FROST GI, HENG HHQ, SCHERER SW, MOHAPATRA G, STERN R. The hyaluronidase gene HYAL1 maps to chromosome 3p21.2-p21.3 in human and 9F1-F2 in mouse, a conserved candidate tumor suppressor locus. *Genomics* 1998; 48: 63-70.
 - 17) ERICKSON M, STERN R. Chain gangs: new aspects of hyaluronan metabolism. *Biochem Res Int* 2011; 2012: 893947-894055.
 - 18) CHAO KL, MUTHUKUMAR L, HERZBERG O. Structure of human hyaluronidase-1, a hyaluronan hydrolyzing enzyme involved in tumor growth and angiogenesis. *Biochemistry* 2007; 46: 6911-6920.
 - 19) MILLER AD, VIGDOROVICH V, STRONG RK, FERNANDES RJ, LERMAN MI. Hyal2, where are you? *Osteoarthritis Cartilage* 2006; 14: 1315-1317.
 - 20) LEPPERDINGER G, STROBL B, KREIL G. HYAL2, a Human gene expressed in many cells, encodes a lysosomal hyaluronidase with a novel type of specificity. *J Biol Chem* 1998; 273: 22466-22470.
 - 21) HEMMING R, MARTIN DC, SLOMINSKI E, NAGY JI, HALAYKO AJ, PIND S, TRIGGS-RAINE B. Mouse Hyal3 encodes a 45- to 56-kDa glycoprotein whose overexpression increases hyaluronidase 1 activity in cultured cells. *Glycobiology* 2008; 18: 280-289.
 - 22) KANEIWA T, MIYAZAKI A, KOGAWA R, MIZUMOTO S, SUGAHARA K, YAMADA S. Identification of amino acid residues required for the substrate specificity of human and mouse chondroitin sulfate hydrolase (conventional hyaluronidase-4). *J Biol Chem* 2012; 287: 42119-42128.
 - 23) HARRISON RA. Preliminary characterization of the multiple forms of ram sperm hyaluronidase. *Biochem J* 1988; 252: 875-882.
 - 24) MEYERS SA. Equine sperm-oocyte interaction: the role of sperm surface hyaluronidase. *Anim Reprod Sci* 2001; 68: 291-303.
 - 25) KIM E, BABA D, KIMURA M, YAMASHITA M, KASHIWABARA S, BABA T. Identification of a hyaluronidase, Hyal5, involved in penetration of mouse sperm through cumulus mass. *Proc Natl Acad Sci USA* 2005; 102: 18028-18033.
 - 26) TRIGGS-RAINE B, SALO TJ, ZHANG H, WICKLOW BA, NATOWICZ MR. Mutations in HYAL1, a member of a tandemly distributed multigene family encoding disparate hyaluronidase activities, cause a newly described lysosomal disorder, mucopolysaccharidosis IX. *Proc Natl Acad Sci USA* 1999; 96: 6296-6300.
 - 27) BERTOLAMI C, BRUCEDONOFF R. Hyaluronidase activity during open wound healing in rabbits: a preliminary report. *J Surg Res* 1978; 25: 256-259.
 - 28) LEPPERDINGER G, MÜLLEGGGER J, KREIL G. Hyal2-less active, but more versatile? *Matrix Biol* 2001; 20: 509-514.
 - 29) RAI SK, DUH F, VIGDOROVICH V, DANILKOVITCH-MIAGKOVA A, LERMAN MI, MILLER AD. Candidate tumor suppressor HYAL2 is a glycosylphosphatidylinositol (GPI)-anchored cell-surface receptor for jaagsiekte sheep retrovirus, the envelope protein of which mediates oncogenic transformation. *PNAS* 2001; 98: 4443-4448.
 - 30) LIU S, DUH F, LERMAN MI, MILLER AD. Role of virus receptor Hyal2 in oncogenic transformation of rodent fibroblasts by sheep betaretrovirus env proteins. *J Virol* 2002; 77: 2850-2858.
 - 31) LEPPERDINGER G, KREIL G, CHEMISTRY AND BIOLOGY OF HYALURONAN. Chapter 27 Functional, structural and biological properties of hyaluronidases, ed. G. HG, H. CA. Elsevier Press: Amsterdam, 2004; pp. 586-594.
 - 32) ATMURI V, MARTIN DC, HEMMING R, GUTSOL A, BYERS S, SAHEBIAMF S, THLIVERIS JA, MORT JS, CARMONA E, ANDERSON JE, DAKSHINAMURTI S, TRIGGS-RAINE B.

- Hyaluronidase 3 (HYAL3) knockout mice do not display evidence of hyaluronan accumulation. *Matrix Biol* 2008; 27: 653-660.
- 33) KIMURA M, KIM E, KANG W, YAMASHITA M, SAIGO M, YAMAZAKI T, NAKANISHI T, KASHIWABARA S, BABA T. Functional roles of mouse sperm hyaluronidases, HYAL5 and SPAM1, in fertilization. *Biol Reprod* 2009; 81: 939-947.
 - 34) BABA D, KASHIWABARA S, HONDA A, YAMAGATA K, WU Q, IKAWA M, OKABE M, BABA T. Mouse sperm lacking cell surface hyaluronidase PH-20 can pass through the layer of cumulus cells and fertilize the egg. *J Biol Chem* 2002; 277: 30310-30314.
 - 35) KANG W, ZHOU C, KOGA Y, BABA T. Hyaluronan-degrading activity of mouse sperm hyaluronidase is not required for fertilization? *J Reprod Dev* 2010; 56: 140-144.
 - 36) EIKENES L, TARI M, TUFTO I, BRULAND OS, DE LANGE DAVIES C. Hyaluronidase induces a transcapillary pressure gradient and improves the distribution and uptake of liposomal doxorubicin (Caelyx) in human osteosarcoma xenografts. *Br J Cancer* 2005; 93: 81-88.
 - 37) NOVAK U, STYLLI SS, KAYE AH, LEPPERDINGER G. Hyaluronidase-2 overexpression accelerates intracerebral but not subcutaneous tumor formation of murine astrocytoma cells. *Cancer Res* 1999; 46: 6246-6250.
 - 38) YOFFOU PH, EDJEKOUANE L, MEUNIER L, TREMBLAY A, PROVENCHER DM, MES-MASSON A, CARMONA E. Subtype specific elevated expression of hyaluronidase-1 (HYAL-1) in epithelial ovarian cancer. *PLoS One* 2011; 6: e20705.
 - 39) KULTTI A, LI X, JIANG P, THOMPSON CB, FROST GI, SHEPARD HM. Therapeutic targeting of hyaluronan in the tumor stroma. *Cancers (Basel)* 2012; 4: 873-903.
 - 40) WANG F, GRIGORIEVA EV, LI J, SENCHENKO VN, PAVLOVA TV, ANEDCHENKO EA, KUDRYAVTSEVA AV, TSIMANIS A, ANGELONI D, LERMAN MI, KASHUBA VI, KLEIN G, ZABAROVSKY ER. HYAL1 and HYAL2 inhibit tumour growth *in vivo* but not *in vitro*. *PLoS One* 2008; 3: e3031.
 - 41) CHANG N. Transforming growth factor- β 1 blocks the enhancement of tumor necrosis factor cytotoxicity by hyaluronidase Hyal-2 in L929 fibroblasts. *BMC Cell Biol* 2002; 3: 8-17.
 - 42) CHANG N. Hyaluronidase induces murine L929 fibrosarcoma cells resistant to tumor necrosis factor and fas cytotoxicity in the presence of actinomycin D. *Cell Biochem Biophys* 1995; 27: 109-132.
 - 43) MCBRIDE WH, BARD JB. Hyaluronidase-sensitive halos around adherent cells. Their role in blocking lymphocyte-mediated cytotoxicity. *J Exp Med* 1979; 149: 507-515.
 - 44) LOKESHWAR VB, ESTRELLA V, LOPEZ L, KRAMER M, GOMEZ P, SOLOWAY MS, LOKESHWAR BL. HYAL1-v1, an alternatively spliced variant of HYAL1 hyaluronidase: a negative regulator of bladder cancer. *Cancer Res* 2006; 66: 11219-11227.
 - 45) DE SÁ VK, OLIVIERI E, PARRA ER, AB'SABER AM, TAKAGAKI T, SOARES FA, CARRARO D, CARVALHO L, CAPELOZZI VL. Hyaluronidase splice variants are associated with histology and outcome in adenocarcinoma and squamous cell carcinoma of the lung. *Hum Pathol* 2012; 43: 675-683.
 - 46) EISSA S, ZOHNY SF, SHEHATA HH, HEGAZY MG, SALEM AM, ESMAT M. Urinary retinoic acid receptor- β 2 gene promoter methylation and hyaluronidase activity as noninvasive tests for diagnosis of bladder cancer. *Clin Biochem* 2012; 45: 402-407.
 - 47) GUEDAN S, ROJAS JJ, GROS A, MERCADE E, CASCALLO M, ALEMANY R. Hyaluronidase expression by an oncolytic adenovirus enhances its intratumoral spread and suppresses tumor growth. *Mol Ther* 2010; 18: 1275-1283.
 - 48) GANESH S, GONZALEZ-EDICK M, GIBBONS D, VAN RM, JOOSS K. Intratumoral coadministration of hyaluronidase enzyme and oncolytic adenoviruses enhances virus potency in metastatic tumor models. *Clin Cancer Res* 2008; 14: 3933-3941.
 - 49) BERTELLI G, DINI D, FORON GB, GOZZA A, SILVEATRO S, VWNTURIN M, ROSSO R, PRONZATO P. Hyaluronidase as an antidote to extravasation of Vinca alkaloids: clinical results. *J Cancer Res Clin Oncol* 1994; 120: 505-506.
 - 50) COLACIOPPO PM, RIESCO MLG, KOIFFMAN MD. Use of hyaluronidase to prevent perineal trauma during spontaneous births a randomized, placebo-controlled, double-blind, clinical trial. *J Midwifery Wom Heal* 2011; 56: 436-445.
 - 51) WOHLRAB J, FINKE R, FRANKE WG, WOHLRAB A. Clinical trial for safety evaluation of hyaluronidase as diffusion enhancing adjuvant for infiltration analgesia of skin with lidocaine. *Dermatol Surg* 2012; 38: 91-96.
 - 52) PACELLA E, COLLINI S, PACELLA F, PIRAINO DC, SANTAMARIA V, DE BLASI RA. Levobupivacaine vs. racemic bupivacaine in peribulbar anaesthesia: a randomized double blind study in ophthalmic surgery. *Eur Rev Med Pharmacol Sci* 2010; 14: 539-544.
 - 53) LENSTRUP J. Hyaluronidase in subcutaneous infusion of fluid. *Acta Pharmacol Toxicol* 1951; 7: 143-152.
 - 54) MUCHMORE DB, VAUGHN DE. Review of the mechanism of action and clinical efficacy of recombinant human hyaluronidase coadministration with current prandial insulin formulations. *J Diabetes Sci Technol* 2010; 4: 419-428.
 - 55) JOLLES S. Hyaluronidase facilitated subcutaneous immunoglobulin in primary immunodeficiency. *ImmunoTargets Therapy* 2013; 2: 125-133.
 - 56) WOHLRAB J, FINKE R, FRANKE WALTER G, WOHLRAB A. Efficacy study of hyaluronidase as a diffusion promoter for lidocaine in infiltration analgesia of skin. *Plast Reconstr Surg* 2012; 129: 771e-772e.
 - 57) HUANG Z, ZHAO C, CHEN Y, COWELL JA, WEI G, KULTTI A, HUANG L, THOMPSON CB, ROSENGREN S, FROST GI, SHEPARD HM. Recombinant human hyaluronidase PH20 does not stimulate an acute inflammatory response and inhibits

- lipopolysaccharide-induced neutrophil recruitment in the air pouch model of inflammation. *J Immunol* 2014; 192: 5285-5295.
- 58) HILTON S, SCHRUMPF H, BUHREN BA, BÖLKE E, GERBER PA. Hyaluronidase injection for the treatment of eyelid edema a retrospective analysis of 20 patients. *Eur J Med Res* 2014; 19: 30-34.
 - 59) LEE A. Adjuvants to local anaesthesia in ophthalmic surgery. *Br J Ophthalmol* 2011; 95: 1345-1349.
 - 60) SCHULENBURG HE, SRI-CHANDANA C, LYONS G, COLUMB MO, MCLURE HA. Hyaluronidase reduces local anaesthetic volumes for sub-Tenon's anaesthesia. *Br J Anaesth* 2007; 99: 717-720.
 - 61) SILVERSTEIN AM, GREENBAUM S, STERN R. Hyaluronidase in ophthalmology. *J Applied Res* 2012; 12: 1-13.
 - 62) MANTOVANI C, BRYANT AE, NICHOLSON G. Efficacy of varying concentrations of hyaluronidase in peribulbar anaesthesia. *Br J Anaesth* 2001; 86: 876-878.
 - 63) NARAYANAN R, KUPPERMANN BD. Hyaluronidase for pharmacologic vitreolysis. *Dev Ophthalmol* 2009; 44: 20-25.
 - 64) KHANDWALA M, AHMED S, GOEL S, SIMMONS IG, MCLURE HA. The effect of hyaluronidase on ultrasound-measured dispersal of local anaesthetic following sub-Tenon injection. *Eye* 2008; 22: 1065-1068.
 - 65) JUMPER JM, MCCAULEY MB, EQUI RA, DUNCAN KG, DUNCAN J, SCHWARTZ DM. Corneal toxicity of intraocular hyaluronidase. *J Ocul Pharmacol Ther* 2002; 18: 89-97.
 - 66) DELAERE L, ZEYEN T, FOETS B, VAN CJ, STALMANS I. Allergic reaction to hyaluronidase after retrobulbar anaesthesia: a case series and review. *Int Ophthalmol* 2009; 29: 521-528.
 - 67) ZAMORA-ALEJO K, MOORE S, LEATHERBARROW B, NORRIS JH, LAKE DB, MALHOTRA R, SELVA D, GOGGIN M. Hyaluronidase toxicity a possible cause of post-operative periorbital inflammation. *Clin Experiment Ophthalmol* 2013; 41: 122-126.
 - 68) KRAMER MW, GOLSHANI R, MERSEBURGER AS, KNAPP J, GARCIA A, HENNENLOTTER J, DUNCAN RC, SOLOWAY MS, JORDA M, KUCZYK MA, STENZL A, LOKESHWAR VB. HYAL-1 hyaluronidase: a potential prognostic indicator for progression to muscle invasion and recurrence in bladder cancer. *Eur Urol* 2010; 57: 86-94.
 - 69) TAN J, WANG X, SU X, LI H, SHI Y, WANG L, REN G. Upregulation of HYAL1 expression in breast cancer promoted tumor cell proliferation, migration, invasion and angiogenesis. *PLoS One* 2011; 6: e22836.
 - 70) JAMSHIDIAN H, HASHEMI M, NOWROOZI MR, AYATI M, BONYADI M, TOUSI VN. Sensitivity and specificity of urinary hyaluronic acid and hyaluronidase in detection of bladder transitional cell carcinoma. *Urol J* 2011; 11: 1232-1237.
 - 71) DYCHTER SS, HARRIGAN R, BAHN JD, PRINTZ MA, SUGARMAN BJ, DENOIA E, HAUGHEY DB, FELLOWS D, MANEVAL DC. Tolerability and pharmacokinetic properties of ondansetron administered subcutaneously with recombinant human hyaluronidase in minipigs and healthy volunteers. *Clin Ther* 2014; 36: 211-224.
 - 72) KIM D, YOON E, JI Y, PARK S, LEE B, DHONG E. Vascular complications of hyaluronic acid fillers and the role of hyaluronidase in management. *J Plast Reconstr Aesthet Surg* 2011; 64: 1590-1595.
 - 73) BAILEY SH, FAGIEN S, ROHRICH RJ. Changing role of hyaluronidase in plastic surgery. *Plast Reconstr Surg* 2014; 133: 127e-132e.
 - 74) CAVALLINI M, GAZZOLA R, METALLA M, VAIENTI L. The role of hyaluronidase in the treatment of complications from hyaluronic acid dermal fillers. *Aesthet Surg J* 2013; 33: 1167-1174.
 - 75) CAVALLINI M, ANTONIOLI B, GAZZOLA R, TOSCA M, GALUZZI M, RAPISARDA V, CIANCIO F, MARAZZI M. Hyaluronidases for treating complications by hyaluronic acid dermal fillers evaluation of the effects on cell cultures and human skin. *Eur J Plast Surg* 2013; 36: 477-484.
 - 76) MATIN-DELEON PA. Epididymal SPAM1 and its impact on sperm function. *Mol Cell Endocrinol* 2006; 250: 114-121.
 - 77) ZHOU C, KANG W, BABA T. Functional characterization of double-knockout mouse sperm Lacking SPAM1 and ACR or SPAM1 and PRSS21 in fertilization. *J Reprod Dev* 2012; 58: 330-337.
 - 78) DUMARESO-DOIRON K, EDJEKOUANE L, ORIMOTO AM, YOFFOU PH, GUSHULAK L, TRIGGS-RAINE B, CARMONA E. Hyal-1 but not Hyal-3 deficiency has an impact on ovarian folliculogenesis and female fertility by altering the follistatin/activin/Smad3 pathway and the apoptotic process. *J Cell Physiol* 2012; 227: 1911-1922.
 - 79) PRIMAKOFF P, LATHROP W, WOOLMAN L, COWAN A, MYLES D. Fully effective contraception in male and female guinea pigs immunized with the sperm protein PH-20. *Nature* 1988; 334: 543-546.
 - 80) WASSERMAN RL, MELAMED I, STEIN MR, GUPTA S, PUCK J, ENGL W, LEIBL H, MCCOY B, EMPSON VG, GELMONT D, SCHIFF RI. Recombinant human hyaluronidase-facilitated subcutaneous infusion of human immunoglobulins for primary immunodeficiency. *J Allergy Clin Immunol* 2012; 130: 951-957.
 - 81) HANRAHAN K. Hyaluronidase for treatment of intravenous extravasations: implementation of an evidence-based guideline in a pediatric population. *J Spec Pediatr Nurs* 2013; 18: 253-262.
 - 82) EL-SAFORY NS, FAZARY AE, LEE C. Hyaluronidases, a group of glycosidases: current and future perspectives. *Carbohydr Polym* 2010; 81: 165-181.
 - 83) ZAMORA-ALEJO K, GOGGIN M. Recombinant hyaluronidase: response. *Clin Experiment Ophthalmol* 2014; 42: 299.