

Endothelial constitutive nitric oxide synthase, angiotensin converting enzyme, angiotensin II type 1 receptor gene polymorphisms and endothelial functions in healthy individuals

T.S. AKPINAR, A. OZKOK¹, M. KOSE, R. ATAS, A. SUMNU, O.K. BAKKALOGLU, O. ERK, M.S. KAYACAN, H. OFLAZ², V.A. AKKAYA, A.S. DEMIREL

Department of Internal Medicine, Istanbul University, Istanbul Faculty of Medicine, Istanbul, Turkey

¹Department of Internal Medicine and Nephrology, Istanbul Medeniyet University, Goztepe Training and Research Hospital, Istanbul, Turkey

²Department of Cardiology, Istanbul University, Istanbul Faculty of Medicine, Istanbul, Turkey

Abstract. – **INTRODUCTION:** Endothelial dysfunction is recognized as an early and initiating event in the pathogenesis of coronary artery disease. Gene polymorphisms of endothelial constitutive nitric oxide synthase (ecNOS), angiotensin converting enzyme (ACE) and angiotensin II type 1 receptor (AT1R) have been found to be associated with atherosclerosis. We aimed to investigate the possible effects of ecNOS, ACE and AT1R gene polymorphisms on endothelial functions in healthy population.

MATERIALS AND METHODS: In 255 healthy subjects (male/female: 119/136 mean age 35.1±2.3 years) ecNOS, ACE and AT1R gene polymorphisms were assessed by polymerase chain reaction (PCR). Endothelium dependent (EDD, flow-mediated) and endothelium independent vasodilation (EID) were measured by high resolution brachial artery ultrasound and 0.5mg sublingual nitroglycerine respectively.

RESULTS: ecNOS and ACE genes had no significant effect on EDD and EID. However, subjects with AT1RAC+CC genotypes had lower EDD compared to subjects with AT1RAA genotype in females (19.4 ± 6.6% vs 21.5 ± 7.8%, $p = 0.041$). EDD and EID were significantly negatively associated with age, body mass index, serum creatinine, glucose, uric acid and hemoglobin levels. When the data on age, uric acid, BMI, glucose, creatinine, and hemoglobin were split into 3 as low-1/3, mid-1/3 and high 1/3, there was significant graded decrease in EDD and EID with these parameters. In multiple regression analysis, age and presence of AT1RAC+CC genotype retained as significant independent factors predicting endothelial functions.

CONCLUSIONS: Gene polymorphisms of endothelial constitutive nitric oxide synthase and angiotensin converting enzyme had no effect on endothelial functions. However, the presence of angiotensin II type 1 receptor polymorphism (AT1RAC+CC genotype) seemed to adversely af-

fect the endothelial functions as reflected by impaired endothelium dependent and independent vasodilatation in healthy individuals.

Key Words:

Angiotensin converting enzyme polymorphism, Angiotensin II type 1 receptor polymorphisms, Endothelial constitutive nitric oxide synthase polymorphism, Endothelial dysfunction, Endothelium dependent vasodilatation, Endothelium independent vasodilatation.

Introduction

Endothelial dysfunction is recognized as an early and initiating event in the pathogenesis of coronary artery disease (CAD)¹⁻³. Endothelial function also has a long term prognostic importance in terms of progression of cardiovascular diseases^{4,5}. Gene polymorphisms of endothelial constitutive nitric oxide synthase (ecNOS)^{6,7}, angiotensin converting enzyme (ACE)^{8,9} and angiotensin II type 1 receptor (AT1R)⁹ have been found to be associated with atherosclerosis and CAD. Herein, we aimed to investigate the possible effects of ecNOS, ACE and AT1R gene polymorphisms on endothelial functions measured as both endothelium dependent and independent vasodilatation in healthy population.

Subjects and Methods

Study Population

Two hundred and fifty-five healthy subjects (male/female: 119/136, mean age 35.1 ± 12.3 years) were enrolled into the study. Each subject

was evaluated by history, physical examination, and screening laboratory tests. Exclusion criteria included evidence or history of diabetes mellitus, hypertension, cardiovascular disease and chronic medication use including hormone replacement therapy. The study was approved by the Ethics Committee of the Istanbul Medical Faculty, University of Istanbul (No: 2009/925). Informed consent was obtained from each subject and the Declaration of Helsinki on Biomedical Research on Humans was followed for the study.

Laboratory Analysis

Major cardiovascular risk factors such as body mass index (BMI), smoking, family history (myocardial infarction or sudden death in first degree consanguinity in men before 55, in women before 65 years of age), glucose, total cholesterol, high density lipoprotein (HDL), low density lipoprotein (LDL), triglyceride and uric acid were measured and recorded. Biochemical analyses were performed by standard methods in the clinical biochemical laboratory of Istanbul Faculty of Medicine.

Analysis of the Variable Number of Tandem repeats (VNTRs) Polymorphism of the Endothelial Constitutive Nitric Oxide Synthase (ecNOS) Gene

ecNOS genotypes were determined by polymerase chain reaction (PCR) using oligonucleotide primers (sense: 5'-AGGCCCTATGGTAGTGCCTTT-3'; antisense: 5'-TCTCTAGTGCTGTGG TCAT-3') that flank the region of the 27 base pair (bp) direct repeat in intron 4 as described previously with minor modifications¹⁰. Reactions were performed in a total volume of 50 (l containing 500 ng genomic DNA, 10 pmol of each primer, 0.2 mM dNTP, 0.5 U Taq DNA polymerase (MBI Fermentas Inc., New York, NY, USA), 5(l PCR buffer (500 mmol/l KCl, 100 mmol trihydroxymethylaminomethane hydrochloride and 0.8% Nonidet

P40; MBI Fermentas Inc.). The thermocycling procedure consisted of initial denaturation at 94°C for 5 minutes, 35 cycles of denaturation for 1 minute, annealing at 55°C for 1 minute, extension at 72°C for 1 minute. The PCR products were analyzed using 3% agarose gel electrophoresis and visualized by ethidium bromide staining.

Analysis of the Insertion/Deletion Polymorphism of the Angiotensin Converting enzyme (ACE) Gene

Genomic DNA was isolated from leukocytes obtained from peripheral venous blood samples using the method described by Miller et al¹¹. Sixteenth intron of the ACE gene was amplified with PCR as described by Rigat et al¹². In the first PCR, a 490-bp fragment representing the I allele and a 190-bp fragment representing the D allele were obtained (sense: 5'-CTGGAGACCACTCCCATCCTTTCT-3'; antisense: 5'-GATGTGGCCATCACATTC GTCAGAT-3'). Second PCR reaction is necessary for the confirmation of the ACE genotype by an insertion specific PCR amplification with the primer pair of Lindpaintner et al¹³ obtained (sense: 5'-TGGGAC-CACA GCGCCCGCCACTAC-3'; antisense: 5'-TCGCCAGCC CTCCCATGCCCATAA-3'). The first PCR product was electrophoresed on a 3% agarose gel. A 1.5% agarose gel was used for visualizing the second PCR product. The gels were stained with ethidium bromide and photographed under UV light.

Analysis of the A1166C Polymorphism of the Angiotensin II Type 1 Receptor (AT1R) Gene

Genomic DNA was extracted from white blood cells. The region of the AT1R located between nucleotides 423 and 1278 of the cDNA (14) was amplified by using oligonucleotides 5'-GGCTT TGCTTTGTCTTGTTG and 5'-AATGCTTGTAGCC AAAGTCACCT as sense and

Table I. Genotype and allele frequencies (n = 255).

ecNOS Gene Polymorphism		AT1R gene polymorphism		ACE gene polymorphism	
aa genotype	3	AA genotype	184	II genotype	45
ab genotype	42	AC genotype	68	ID genotype	111
bb genotype	210	CC genotype	3	DD genotype	99
a allele	0.09	A allele	0.85	I allele	0.39
b allele	0.91	C allele	0.15	D allele	0.61

Table II. The comparison of cardiovascular risk factors according to sex (*Myocardial infarction or sudden death in first degree consanguinity in men before 55, in women before 65 years of age).

	Male (n=119) Mean ± SD	Female (n=136) Mean ± SD	p value
Age (years)	37 ± 12	33 ± 13	0.009
Smoking	36.1%	20.1%	0.002
Family History*	20.2%	23.5%	NS
Glucose	86 ± 9	85 ± 9	NS
Total Cholesterol	172 ± 35	166 ± 27	NS
HDL	41 ± 9	51 ± 12	<0.001
LDL	103 ± 28	96 ± 23	0.024
Triglyceride	139 ± 70	92 ± 47	<0.001
Uric acid	4.9 ± 1.0	3.6 ± 0.8	<0.001
BMI	25.2 ± 3.1	22.9 ± 3.9	<0.001

antisense primers, respectively. The reaction conditions were: 2 ng/L of genomic DNA, 0.75 (mol/L of each oligonucleotide, 75 (mol/L of each dNTP, 1.5 mmol/L of MgCl₂, 75 mmol/L Tris-HCl (pH 9.0), 5 mmol/L KCl, 20 mmol/L (NH₄)₂SO₄, 0.2 U Taq DNA polymerase (MBI Fermentas Inc., New York, NY, USA), in a final volume of 10 (L DNA was amplified with an initial denaturation step at 94°C for 3 min, followed by 40 cycles of 94°C for 30 sec, 60°C for 30 sec, 72°C for 90 sec, and a final elongation step at 72°C for 5 min. The A1166C polymorphism was detected by digestion of the PCR product with Ddel (Dopachrome Tautomerase Distal Enhancer 1), which cuts at positions 1167, when C is present at 1166 and 1023.

Endothelial Function Assessment

Brachial artery was imaged on a commercially available ultrasound system (VINGMED Technology, System Five, Horten, Norway) using a 10.0 MHz linear phased-array ultrasound transducer longitudinally just above the antecubital fossa as previously described^{15,16}. Two hundred and thirty-two subjects were assessed for endothelial function. Blood pressure cuff was wrapped around the upper arm¹⁶, inflated to 250 mmHg and held for 5 minutes to induce ischemia. The cuff was released and brachial artery diameter was measured every minute for 5 minutes to assess maximal endothelium dependent vasodilation (EDD) in response to reactive hyperemia. After vessel diameter returned to baseline values (~ 7-10 minutes), endothelium independent vasodilation (EID) was assessed af-

ter 0.5 mg sublingual nitroglycerine, every minute for 5 minutes. Vessel diameters were measured at the end diastole coincident with the onset of the R-wave of the simultaneously obtained ECG trace. During the measurements particular attention was paid to the temperature of the laboratory, menstrual cycle, exercise, drugs, food, and sympathetic stimuli as recommended by the guidelines¹⁷. The percent vasodilation was calculated with the following formula:

$$\text{Percent EDD or EID} = 100 \times \frac{(\text{pBAD} - \text{bBAD})}{(\text{bBAD})}$$

pBAD: Peak brachial artery diameter after intervention
bBAD: Baseline brachial artery diameter

The intra and inter-observer variability of the measurements in our laboratory was 1-3%. Brachial artery ultrasound evaluation was done by an experienced physician.

Statistical Analysis

The statistical analysis was done with a commercially available Statistical Package for Social Sciences for Windows version 10.0 (SPSS Inc, Chicago, IL, USA). The means of two groups of numerical variables were compared with independent samples Student's *t* test. The variables were log-transformed if the assumptions for normal distribution were violated. Ordinal variables were analyzed with χ^2 test. For 2↔2 contingency tables Yates correction was done. If expected frequencies in the cells of 2↔2 table were less than

Table III. The correlation of risk factors with endothelium dependent (EDD) and independent (EID) vasodilation

	EDD		EID	
	r	p value	r	p value
Age	-0.34	<0.001	-0.37	<0.001
Sex	0.33	<0.001	0.30	<0.001
Glucose levels	-0.16	0.016	-0.16	0.015
Uric acid levels	-0.30	<0.001	-0.31	<0.001
Body mass index	-0.26	<0.001	-0.25	<0.001
Creatinine levels	-0.21	0.001	-0.19	0.004
Hemoglobin levels	-0.28	<0.001	-0.31	<0.001
AT1R	-0.13	0.041	-0.09	NS

5, Fisher’s exact test was used. If tables examined were greater than 2x2 then appropriate rows or columns were combined to obtain expected frequencies of more than 5. Correlation between two numerical variables with normal distribution was sought with Pearson’s bivariate correlation test, else correlation was done with Spearman’s correlation test. The patients were divided into lower, mid and higher thirds for age, hemoglobin, uric acid, glucose, body mass index (BMI), creatinine and analyzed with one-way ANOVA to examine the effect of the variability of the parameters on EDD and EID. Factors that were significantly related to EDD and EID in univariate testing were examined with multivariate linear regression analysis with forward inclusion to identify the principal causes of the variability observed in EDD and EID. *p* value < 0.05 was accepted as statistically significant.

Results

The frequencies of genotypes and alleles of ACE, AT1R, and ecNOS genes were presented in Table I. The distributions of the genotypes were in Hardy-Weinberg equilibrium. The presence of ecNOS and ACE gene polymorphisms had no significant effect on EDD and EID. However, female subjects with AT1RAC+ CC genotype had significantly lower EDD compared to female subjects with AT1RAA genotype ($19.4 \pm 6.6\%$ vs $21.5 \pm 7.8\%$, *p* = 0.041) (Figure 1).

The comparison of common cardiovascular risk factors are shown according to sex in Table II. Accordingly, females had a more favorable cardiovascular risk profile compared to males.

EDD and EID were found to be strongly correlated with each other (*r* = 0.88, *p* < 0.001). The correlations found with biochemical parameters and other risk factors are listed in Table III. Accordingly EDD and EID showed significant correlations with age, sex, BMI, hemoglobin, serum creatinine, uric acid and glucose. The EDD and EID were significantly related to sex with males having lower EDD ($18.4 \pm 5.7\%$ vs $23.2 \pm 8.2\%$, *p* < 0.001) and EID ($21.7 \pm 6.6\%$ vs $27.1 \pm 9.4\%$, *p* < 0.001) compared to females. When females were compared with age matched male controls for EDD and EID in premenopause (EDD $18.9 \pm 5.3\%$ vs $24.0 \pm 7.9\%$, *p* < 0.001; EID $22.6 \pm 6.2\%$ vs $28.1 \pm 8.8\%$, *p* < 0.001) and in postmenopause (EDD $16.3 \pm 6.9\%$ vs $16.7 \pm 8.3\%$, *p* = NS; EID $18.2 \pm 7.1\%$ vs $19.3 \pm 10.1\%$, *p* = NS), the difference between sexes were apparent only in premenopause.

When the data on age, uric acid, BMI, glucose, creatinine, and hemoglobin were split into 3 (low-1/3, mid-1/3 and high 1/3), there was significant graded decrease in EDD and EID with

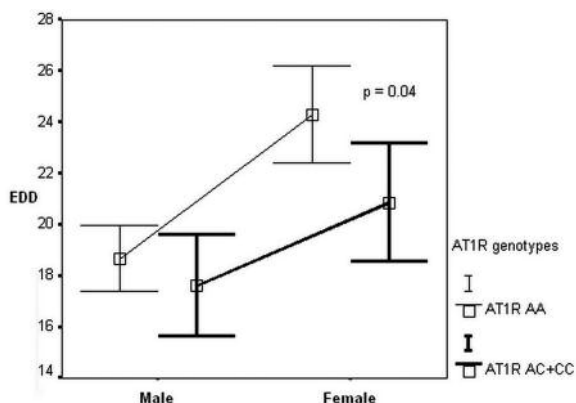


Figure 1. Female subjects with AT1RAC+ CC genotype had significantly lower EDD compared to female subjects with AT1RAA genotype. No significant difference was detected in males in terms of AT1R gene polymorphism.

Table IV. The effect of the distribution of biochemical parameters on endothelium dependent (EDD) and independent (EID) vasodilatation.

		EDD %	EID %
Age	<i>p</i> value	<0.001	<0.001
	Low 1/3	23.2 ± 8.2	27.3 ± 9.2
	Mid 1/3	20.6 ± 6.6	24.1 ± 7.4
	High 1/3	18.4 ± 6.6	21.6 ± 8.0
Uric acid	<i>p</i> value	<0.001	<0.001
	Low 1/3	23.6 ± 8.5	27.5 ± 9.5
	Mid 1/3	20.4 ± 7.7	24.1 ± 8.8
	High 1/3	18.4 ± 5.0	21.8 ± 6.1
Body mass index	<i>p</i> value	<0.001	<0.001
	Low 1/3	23.6 ± 8.3	27.4 ± 8.8
	Mid 1/3	19.6 ± 6.9	23.2 ± 8.5
	High 1/3	19.2 ± 6.4	22.7 ± 7.6
Serum creatinine	<i>p</i> value	0.029	0.025
	Low 1/3	22.5 ± 8.9	26.6 ± 10.1
	Mid 1/3	21.2 ± 7.0	24.6 ± 8.1
	High 1/3	18.9 ± 6.8	22.4 ± 7.5
Hemoglobin	<i>p</i> value	0.002	<0.001
	Low 1/3	22.7 ± 8.4	27.1 ± 9.5
	Mid 1/3	21.3 ± 7.2	24.9 ± 8.0
	High 1/3	18.6 ± 6.2	21.4 ± 7.1
Glucose	<i>p</i> value	0.005	0.008
	Low 1/3	23.2 ± 9.4	27.0 ± 10.4
	Mid 1/3	19.8 ± 6.3	23.7 ± 7.6
	High 1/3	19.8 ± 6.2	22.9 ± 7.1

increasing age, glucose, BMI, creatinine, hemoglobin and uric acid levels (Table IV).

When factors found significantly related to EDD in univariate testing included in multiple regression test, only age, sex and presence of AT1RAC+CC genotypes were retained as significant (multiple $R = 0.461$, adjusted $R^2 = 0.202$, $F = 20.47$, $p < 0.001$). When the same analysis was performed for EID, age, hemoglobin levels and presence of AT1RAC+CC genotype were retained as significant (multiple $R = 0.484$, adjusted $R^2 = 0.224$, $F = 23.202$, $p < 0.001$).

Discussion

In this study, possible influences of eNOS, ACE and AT1R gene polymorphisms on endothelial functions were investigated in healthy individuals. AT1RAC+CC genotype was found to adversely affect the endothelial functions as both endothelium dependent and independent vasodilatation in healthy individuals. Reports assessing the effect of the genes on endothelial function in

healthy humans are scarce in the literature^{6-9,18-22}. In a study that endothelial functions were measured non-invasively, no significant influence of ACE genotypes on EDD and EID was found¹⁸. However, Butler et al²¹ ($n = 68$) found that both, EDD and EID were impaired in young healthy men with D allele. Later Rossi et al²² investigated another polymorphism and showed that the T-786C promoter polymorphism and its interaction with exon 7 Glu298Asp affected EDD in healthy normotensive Caucasian subjects. In the study by Gururajan et al⁶, eNOS gene polymorphism was investigated in 106 patients with acute coronary syndrome and 100 healthy controls. eNOS gene polymorphism was significantly associated with acute coronary syndrome. In another study⁷, the eNOS4a allele was found to be related to carotid atherosclerosis in type 2 diabetic patients. In our work, we evaluated 3 gene polymorphisms and AT1R allele was shown to have deleterious effect on EDD in women. As reported above, our findings about ACE gene are in agreement with the literature. In contrast to Rossi et al study²² we investigated eNOS gene polymorphism in intron 4 and found no effect on EDD and EID. Studies performed with AT1R antagonists have shown that AT1R blockage may improve endothelial function endothelial function. The putative mechanism involved was the prevention of the inactivation of NO by superoxide anions. This hypothesis was supported by the finding that NOS inhibitor, L-N-monomethyl-arginine prevented the improvement in endothelial function seen with losartan²³ and AT1R antagonists increased the antioxidative potential of the vessel wall by increasing the activity of endothelial superoxide dismutase²⁴.

In this study, we observed that women had better EDD compared to men. This difference in EDD disappeared after menopause as reported elsewhere²⁵⁻²⁷. These findings may imply the importance of sex hormones in protecting the endothelium. All these studies are in line with our findings.

We also found that increasing glucose levels adversely affected endothelial functions as reflected by impaired EDD and EID. These findings were in parallel to the findings of the study by Sarabi et al²⁷.

In this report only 20% of the variability of EDD and EID was explained by the parameters collected in the study which implies that the greater part of variability in EDD and EID remained unexplained. Further studies with novel parameters are required to explain the remaining variability in EDD and EID.

Conclusions

Gene polymorphisms of endothelial constitutive nitric oxide synthase and angiotensin converting enzyme had no effect on endothelial functions. However, the presence of angiotensin II type 1 receptor polymorphism (AT1RAC+CC genotype) seemed to adversely affect the endothelial functions as reflected by impaired endothelium dependent and independent vasodilatation in healthy individuals.

Conflict of Interest

The Authors declare that there are no conflicts of interest.

References

- 1) PANZA JA, QUYYUMI AA, BRUSH JE, EPSTEIN SE. Abnormal endothelium-dependent vascular relaxation in patients with essential hypertension. *N Engl J Med* 1990; 323: 22-27.
- 2) WILLIAMS SB, CUSCO JA, RODDY MA, JOHNSTONE MT, CREAGER MA. Impaired nitric oxide mediated vasodilation in patients with non-insulin-dependent diabetes mellitus. *J Am Coll Cardiol* 1996; 27: 567-574.
- 3) RAMSEY MW, GOODFELLOW J, JONES CJH, LUDDINGTON LA, LEWIS MJ, HENDERSON AH. Endothelial control of arterial distensibility is impaired in chronic heart failure. *Circulation* 1995; 92: 3212-3219.
- 4) SUWAIDI JA, HAMASAKI S, HIGANO ST, NISHIMURA RA, HOLMES DR JR, LERMAN A. Long term follow-up of patients with mild coronary artery disease and endothelial dysfunction. *Circulation* 2000; 101: 948-954.
- 5) PERTICONE F, CERAVOLO R, PUJIA A, VENTURA G, JACOPINO S, SCOZZAFAWA A, FERRARO A, CHELLO M, MASTROBERTO P, VERDECCHIA P, SCHILLACI G. Prognostic significance of endothelial dysfunction in hypertensive patients. *Circulation* 2001; 104: 191-196.
- 6) GURURAJAN P, GURUMURTHY P, VICTOR D, SRINIVASA NAGESWARA RAO G, SAI BABU R, SARASA BHARATI A, CHERIAN KM. Plasma total nitric oxide and endothelial constitutive nitric oxide synthase (eNOS) gene polymorphism: a study in a South Indian population. *Biochem Genet.* 2011;49:96-103.
- 7) PARK JH, KIM JH, KIM WH, KIM DS, PARK TS, BAEK HS. Association of the endothelial nitric oxide synthase (eNOS) gene polymorphism with carotid atherosclerosis in type 2 diabetes. *Diabetes Res Clin Pract.* 2006;72:322-327.
- 8) ZHANG Z, XU G, LIU D, FAN X, ZHU W, LIU X. Angiotensin-converting enzyme insertion/deletion polymorphism contributes to ischemic stroke risk: a meta-analysis of 50 case-control studies. *PLoS One.* 2012;7:e46495.
- 9) KAUR R, DAS R, AHLUWALIA J, KUMAR RM, TALWAR KK. Synergistic effect of angiotensin II type-1 receptor 1166A/C with angiotensin-converting enzyme polymorphism on risk of acute myocardial infarction in north Indians. *J Renin Angiotensin Aldosterone Syst.* 2012;13:440-445.
- 10) WANG XL, SIM AS, BODENHOP RF, MCCREDIE RM, WILCKEN DE. A smoking- dependent risk of coronary artery disease associated with a polymorphism of the nitric oxide synthase gene. *Nat Med.* 1996; 2: 41-45.
- 11) MILLER SA, DYKES DD, POLLESKY HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res* 1988; 16: 1215.
- 12) RIGAT B, HUBERT C, CORVOL P, SOUBRIER F. PCR detection of the insertion/deletion polymorphism of the human angiotensin converting enzyme (DCP1) (dipeptidylcarboxypeptidase 1). *Nucleic Acids Res* 1992; 20: 1433.
- 13) LINDPAINTEKNER K, PFEFFER MA, KREUTZ R, STAMPFER MJ, GRODSTEIN F, LAMOTTE F, BURING J, HENNEKENS CH. A prospective evaluation of an angiotensin converting enzyme gene polymorphism and the risk of ischemic heart disease. *N Engl J Med* 1995; 332: 706-711.
- 14) NAWATA H, TAKAYANAGA R, OHNAKA K, SAKAI Y, IMASAKI K, YANASE T, IKUYAMA S, TANAKA S, OHE K. Type 1 angiotensin II receptors of adrenal tumors. *Steroids* 1995; 60: 28-34.
- 15) CELERMAJER DS, SORENSEN KE, GOOCH VM, SPIEGELHALTER DJ, MILLER OI, SULLIVAN ID, LLOYD JK, DEANFIELD JE. Non-invasive detection of endothelial dysfunction in children and adults at risk of atherosclerosis. *Lancet* 1992; 340: 1111-1115.
- 16) ANDERSON TJ, UEHATA A, GERHARD MD, MEREDITH IT, KNAB S, DELAGRANGE D, LIEBERMAN EH, GANZ P, CREAGER MA, YEUNG AC, SELWYN AP. Close relation of endothelial function in the human coronary and peripheral circulations. *J Am Coll Cardiol* 1995; 26: 1235-1241.
- 17) CORRETTI MC, ANDERSON TJ, BENJAMIN EJ, CELERMAJER DS, CHARBONNEAU F, CREAGER MA, DEANFIELD J, DREXLER H, GERHARD-HERMAN M, HERRINGTON D, VALLANCE P, VITA J, VOGEL R. Guidelines for the ultrasound assessment of endothelial-dependent flow-mediated vasodilation of the brachial artery. A Report of the International Brachial Artery Reactivity Task Force. *J Am Coll Cardiol* 2002; 39: 257-265.
- 18) CELERMAJER DS, SORENSEN KE, BARLEY J, JEFFREY S, CARTER N, DEANFIELD J. Angiotensin-converting enzyme genotype is not associated with endothelial dysfunction in subjects without other coronary risk factors. *Atherosclerosis* 1994; 111: 121-126.
- 19) PERTICONE F, CERAVOLO R, MAIO R, VENTURA G, ZINGONE A, PERROTTI N, MATTIOLI PL. Angiotensin-converting enzyme gene polymorphism is associated

- with endothelium-dependent vasodilation in never treated hypertensive patients. *Hypertension* 1998; 31: 900-905.
- 20) ROSSI GP, TADDEI S, VIRDIS A, GHIADONI L, ALBERTIN G, FAVILLA S, SUDANO I, PESSINA AC, SALVETTI A. Exclusion of the ACE D/I gene polymorphism as a determinant of endothelial dysfunction. *Hypertension* 2001; 37: 293-300.
- 21) BUTLER R, MORRIS AD, BURCHELL B, STRUTHERS AD. DD angiotensin-converting enzyme gene polymorphism is associated with endothelial dysfunction in normal humans. *Hypertension* 1999; 33: 1164-1168.
- 22) ROSSI GP, TADDEI S, VIRDIS A, CAVALLIN M, GHIADONI L, FAVILLA S, VERSARI D, SUDANO I, PESSINA AC, SALVETTI A. The T-786C and Glu298Asp polymorphism of the endothelial nitric oxide gene affect the forearm blood flow responses of Caucasian hypertensive patients. *J Am Coll Cardiol* 2003; 41: 946-948.
- 23) SUDHIR K, MACGREGOR JS, GUPTA M, BARBANT SD, REDBERG R, YOCK PG, CHATTERJEE K. Effect of selective angiotensin II receptor antagonism and angiotensin converting enzyme inhibition on the coronary vasculature in vivo. Intravascular two-dimensional and Doppler ultrasound studies. *Circulation* 1993; 87: 931-938.
- 24) HORNIG B, LANDMESSER U, KOHLER C, AHLERSMANN D, SPIEKERMANN S, CRISTOPH A, TATGE H, DREXLER H. Comparative effect of ACE-inhibition and angiotensin II-type 1 receptor antagonism on bioavailability of nitric oxide in patients with coronary artery disease-role of superoxide dismutase. *Circulation* 2001; 103: 799-805.
- 25) TADDEI S, VIRDIS A, GHIADONI L, MATTEI P, SUDANO I, BERNINI G, PINTO S, SALVETTI A. Menopause is associated with endothelial dysfunction in women. *Hypertension* 1996; 28: 576-582.
- 26) CELERMAJER DS, SORENSEN KE, SPIEGELHALTER DJ, GEORGAKOPOULOS D, ROBINSON J, DEANFIELD JE. Aging is associated with endothelial dysfunction in healthy men years before the age related decline in women. *J Am Coll Cardiol* 1994; 24: 471-476.
- 27) SARABI M, MILLGARD J, LIND L. Effects of age, gender and metabolic factors on endothelium-dependent vasodilation: A population-based study. *J Intern Med* 1999; 246: 265-274.