Lack of association between CXCL9 and CXCL10 gene polymorphisms and the outcome of rheumatoid arthritis treatment with methotrexate

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Abstract. – OBJECTIVE: Methotrexate (MTX) in low doses is used in the therapy of rheumatoid arthritis (RA). The aim of many studies is to identify factors predicting the outcome of treatment with methotrexate in rheumatoid arthritis. The action of MTX in RA is associated with the inhibition of inflammatory mediators synthesis. CXCL9 and CXCL10 chemokines play the important role in inflammatory response in RA patients. The aim of this study was to examine the association between CXCL9/10 gene polymorphisms and response to therapy of RA patients with MTX.

PATIENTS AND METHODS: The study included 422 patients diagnosed with rheumatoid arthritis, treated with MTX in doses 20 mg weekly. Good responders were defined as patients who were receiving MTX and had a DAS28 of \leq 2.5 at 6 months of therapy. Poor-responders were defined as patients who were receiving MTX and had a DAS28 of > 2.5.

RESULTS: There were not statistically significant associations between studied polymorphisms and the outcome of rheumatoid arthritis treatment with methotrexate.

CONCLUSIONS: The results of this study suggest lack of associations between the polymorphisms in CXCL9 and CXCL10 genes and the response to MTX in RA patients.

Key Words:

Chemokines, Methotrexate, Polymorphism, Rheumatoid arthritis.

Introduction

Rheumatoid arthritis (RA) is a multifactorial disease and its treatment is mainly based on disease modifying antirheumatic drugs (DMARDs)¹. The optimal strategy in RA therapy is to use (DMARDs) in an early stage of disease in order

to reduce disease activity and to prevent joint destruction. This treatment goal is frequently achieved by methotrexate (MTX)². MTX is a foliate antagonist used in the therapy of malignant disorders and low doses of methotrexate were introduced for the treatment of rheumatoid arthritis because of its antiproliferative, immunosuppressive and antiinflammatory properties^{3,4}. MTX is however not effective in all patients and a significant proportion of patients stop the therapy because of inefficacy or adverse events. Identification of genetic determinants of drug efficacy and toxicity will be valuable because they may help in the individualization of the therapy in the patient before initiation of treatment. Previous studies have revealed that therapy with MTX causes the decreased production of mediators which are involved in the inflammatory process such as numerous cytokines and chemokines^{5,6}.

Chemokines have been classified into four families. One of these are the CXC chemokines. To these chemokines belong CXCL9 and CX-CL10, secreted by various cell types, such as, neutrophils, monocytes, endothelial cells, fibroblasts, keratinocytes7. The high level of CX-CL9/10 in peripheral liquids is a marker of immune response, especially involving Th1-cells. In tissues Th1 lymphocytes may be responsible for increased IFN- γ and TNF- α production, which in turn stimulates CXCL9/10 secretion⁸⁻¹⁰. CXCL9 and CXCL10 have been detected in sera, synovial fluid, and synovial tissue in RA patients¹¹⁻¹³. In CXCL9/10 genes several polymorphisms have been detected and correlated with various diseases¹⁴⁻¹⁶. CXCL9 and CXCL10 are the important mediators determining RA activity, the anti-inflammatory action of MTX is associated with the influence on chemokine synthe-

| | Poor responders | | Good responders | | | | | |
|-----------------------------|--------------------|---------|--------------------|--------|-----------------------|-------------|------------|------------------|
| | n . | % | n | % | p ^a | | ρ^{b} | OR (95%) CI) |
| CXCL9 rs3733236 genotype | | | | | | | | |
| GG | 218 | 59.73% | 147 | 40.27% | 0.31 | AA+GA vs GG | 0.47 | 1.24 (0.71-2.18) |
| GA | 29 | 52.73% | 26 | 47.27% | 0.01 | AA vs GA+GG | 0.52 | - |
| AA | 2 | 100.00% | 0 | 0.00% | | AA vs GG | 0.52 | - |
| | | | | | | GA vs GG | 0.38 | 1.33 (0.75-2.35) |
| | | | | | | AA vs GA | 0.49 | - |
| CXCL9 rs3733236 allele | | | | | | | | |
| G | 465 | 93.37% | 320 | 92.49% | | | | |
| А | 33 | 6.63% | 26 | 7.51% | | A vs G | 0.68 | 1.14 (0.67-1.95) |
| CXCL10 rs8878 genotype | | | | | | | | |
| GG | 84 | 60.43% | 55 | 39.57% | 0.46 | AA+AG vs GG | 0.75 | 1.09 (0.72-1.65) |
| AG | 120 | 56.34% | 93 | 43.66% | | AA vs AG+GG | 0.35 | 0.77 (0.45-1.30) |
| AA | 45 | 64.29% | 25 | 35.71% | | AA vs GG | 0.65 | 0.85 (0.47-1.54) |
| | | | | | | AG vs GG | 0.51 | 1.18 (0.77-1.83) |
| | | | | | | AA vs AG | 0.27 | 0.72 (0.41-1.25) |
| CXCL10 rs8878 allele | | | | | | | | |
| G | 288 | 57.83% | 203 | 58.67% | | | | |
| А | 210 | 42.17% | 143 | 41.33% | | A vs G | 0.83 | 0.97 (0.73-1.28) |

Table I. Association between CXCL9 and CXCL10 gene polymorphisms and response to therapy of RA patients with MTX.

 ${}^{a}\chi^{2}$ test; ${}^{b}Fisher$ exact test

sis and therefore the aim of this study was to examine the association between *CXCL9/10* gene polymorphisms and response to therapy of RA patients with MTX.

Patients and Methods

The study included 422 patients (340 female, 82 male, mean age 57.5 \pm 12.4 years) diagnosed with rheumatoid arthritis, treated with MTX in doses 20 mg weekly. RA was diagnosed according to the ACR/EULAR criteria¹⁷.

Evaluation of clinical efficacy

Good responders were defined as patients who were receiving MTX and had a DAS28 of ≤ 2.5 at 6 months of therapy (patients with remission of disease symptoms). Poor-responders were defined as patients who were receiving MTX and had a DAS28 of $> 2.5^{18,19}$. The study was approved by the local Ethics Committee and written informed consent was obtained from all subjects.

Genotyping

DNA was extracted from 200 μ L of whole blood samples using a GeneMATRIX Quick Blood DNA Purification Kit (EURx, Poland). SNPs within the *CXCL9* (rs3733236 G>A) and *CXCL10* (rs8878 A>G) genes were genotyped using TaqMan genotyping assays from Life Technologies Genomic. Fluorescence data were captured using a 7500 FAST Real-Time PCR System (Applied Biosystems, Waltham, MA, USA).

Statistical Analysis

Chi-square or Fisher exact tests were used to compare genotype and allele frequencies between the study groups. p < 0.05 was considered statistically significant.

Results

The efficacy of RA therapy with MTX is presented in Table I. Under MTX therapy remission of RA symptoms was achieved in 40.27% of *CXCL9 GG* genotype carriers and in 47.27% of subjects with GA genotype. The differences were not statistically significant (Table I).

The remission of RA symptoms was observed in 39.57% of *CXCL10* GG genotype carriers, in 43.66% of subjects with AG genotype and in 35.71% with AA genotype. The differences were not statistically significant (Table I).

Discussion

MTX is the drug commonly used in the therapy of RA. The mechanisms by which MTX at low doses modulates inflammation in RA are still not fully explained. Many pharmacological mechanisms have been suggested, including inhibition of purine synthesis, promotion of adenosine release, inhibition of production of proinflammatory cytokines and chemokines, suppression of lymphocyte proliferation³⁻⁶. In this study we analyzed the CXCL9 and CXCL10 gene polymorphisms as the factors predisposing to the achieving of RA remission during therapy with MTX. Our results indicate that these polymorphisms are not the factors associated with the response to the therapy in RA patients treated with MTX. Many previous studies revealed the significant role of CXCL9 and CXC10 in the pathogenesis and inflammatory process in RA. Kuan et al¹¹ examined the association between CXCL9, CXCL10 serum levels and disease activity in RA. The authors observed that the CX-CL9 and CXCL10 serum concentrations were significantly increased in patients with active RA compared to controls. The improvement in clinical activity in patients with RA was accompanied by a significant reduction in the serum concentration of these chemokines. Yoshida et al¹² analyzed gene expression in the microdissected synovial lining cells of RA patients, using those of OA patients as the control. Expression levels of the chemokines CXCL9 and CXCL10 were significantly higher in the synovium of RA than in that of OA. Ruschpler et al²⁰ investigated gene expression patterns in synovial tissue from RA and OA patients. These authors observed that CXCR3 mRNAs, as well as CXCL9 and CX-CL10 mRNAs, were significantly increased in RA as compared with OA. Elevated CXCR3 protein was found to be especially expressed on mast cells within synovial tissue from RA patients. These findings suggest that substantial ex-

pression of CXCR3 protein on mast cells within synovial tissue from RA patients plays a significant role in the pathophysiology of RA, accompanied by elevated levels of the chemokines CX-CL9 and CXCL10. Above studies indicate that chemokines CXCL9 and CXCL10 are the important mediators associated with disease activity in RA patients. Many studies revealed that disease activity is the important factor determining the response of RA patients to MTX²¹. In our previous study we have detect the association between CXCL10 (rs8878) G allele and RA²². In this study we examined whether CXCL9 and CXCL10 gene polymorphisms are associated with the response to the therapy in RA patients treated with MTX.

Conclusions

Although CXCL9 and CXCL10 play the important role in the RA pathogenesis, the results of this study suggest lack of associations between the polymorphisms in *CXCL9* and *CX-CL10* genes and the response to MTX in RA patients. Nevertheless this hypothesis requires further investigations.

Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- ALETAHA D, SMOLEN JS. The rheumatoid arthritis patient in the clinic: comparing more than 1300 consecutive DMARD courses. Rheumatology 2002; 41: 1367-1374.
- SMOLEN JS, ALETAHA D, MACHOLD KP. Therapeutic strategies in early rheumatoid arthritis. Best Pract Res Clin Rheumatol 2005; 19: 163-177.
- BANNWARTH B, LABAT L, MORIDE Y, SCHAEVERBEKE T. Methotrexate in rheumatoid arthritis. An update. Drugs 1994; 47: 25-50.
- CRONSTEIN BN. Going with the flow: methotrexate, adenosine, and blood flow. Ann Rheum Dis 2006; 65: 421-422.
- CONSTANTIN A, LOUBET-LESCOULIÉ P, LAMBERT N, YAS-SINE- DIAB B, ABBAL M, MAZIERES B, DE PREVAL C, CAN-TAGREL A. Antiinflammatory and immunoregulatory action of methotrexatein the treatment of rheumatoid arthritis. Arthritis Rheum 1998; 41: 48-57.
- CUTOLO M, SULLI A, PIZZORNI C, SERIOLO B, STRAUB RH. Anti-inflammatory mechanisms of methotrexate in rheumatoid arthritis. Ann Rheum Dis 2001; 60: 729-735.

- RUTH JH, ROTTMAN JB, KATSCHKE JR KJ, QIN S, WU L, LAROSA G, PONATH P, POPE RM, KOCH AE. Selective lymphocyte chemokine receptor expression in the rheumatoid joint. Arthritis Rheum 2001; 44: 2750-2760.
- ROSSI D, ZLOTNIK A. The biology of chemokines and their receptors. Annu Rev Immunol 2000; 18: 217-242.
- 9) GARCIA-LOPEZ MA, SANCHEZ-MADRID F, RODRIGUEZ-FRADE JM, MELLADO M, ACEVEDO A, GARCÍA MI, ALBAR JP, MARTÍNEZ C, MARAZUELA M. CXCR3 chemokine receptor distribution in normal and inflamed tissues: expression on activated lymphocytes, endothelial cells, and dendritic cells. Lab Invest 2001; 81: 409-418.
- 10) HANAOKA R, KASAMA T, MURAMATSU M, YAJIMA N, SH-IOZAWA F, MIWA Y, NEGISHI M, IDE H, MIYAOKA H, UCHI-DA H, ADACHI M. A novel mechanism for the regulation of IFN-gamma inducible protein-10 expression in rheumatoid arthritis. Arthritis Res Ther 2003; 5: R74-81.
- 11) KUAN WP, TAM LS, WONG CK, KO FW, LI T, ZHU T, LI EK. CXCL 9 and CXCL10 as Sensitive markers of disease activity in patients with rheumatoid arthritis. J Rheumatol 2010; 37: 257-264.
- 12) YOSHIDA S, ARAKAWA F, HIGUCHI F, ISHIBASHI Y, GOTO M, SUGITA Y, NOMURA Y, NIINO D, SHIMIZU K, AOKI R, HASHIKAWA K, KIMURA Y, YASUDA K, TASHIRO K, KUHARA S, NAGATA K, OHSHIMA K. Gene expression analysis of rheumatoid arthritis synovial lining regions by cDNA microarray combined with laser microdissection: up-regulation of inflammation-associated STAT1, IRF1, CXCL9, CX-CL10, and CCL5. Scand J Rheumatol 2012; 41: 170-179.
- 13) LEE EY, SEO MR, JUHN YS, KIM JY, HONG YJ, LEE YJ, LEE EB, SONG YW. Potential role and mechanism of IFN-gamma inducible protein-10 on receptor activator of nuclear factor kappa-B ligand (RANKL) expression in rheumatoid arthritis. Arthritis Res Ther 2011; 13: R104.
- 14) KLICH I, FENDLER W, WYKA K, MŁYNARSKI W. Effect of the IP10 (CXCL10) and HLA genotype on the risk of type 1 diabetes in children. Pediatr Endocrinol Diabetes Metab 2011; 17: 10-13.
- 15) NOGUEIRA LG, SANTOS RH, IANNI BM, FIORELLI AI, MAIRENA EC, BENVENUTI LA, FRADE A, DONADI E, DIAS F, SABA B, WANG HT, FRAGATA A, SAMPAIO M, HIRATA MH, BUCK P, MADY C, BOCCHI EA, STOLF NA, KALIL J, CUNHA-NETO E. Myocardial chemokine expression

and intensity of myocarditis in Chagas cardiomyopathy are controlled by polymorphisms in CX-CL9 and CXCL10. PLoS Negl Trop Dis 2012; 6: e1867.

- 16) LACHER M, KAPPLER R, BERKHOLZ S, BAURECHT H, VON SCHWEINITZ D, KOLETZKO S. Association of a CXCL9 polymorphism with pediatric Crohn's disease. Biochem Biophys Res Commun 2007; 363: 701-707.
- 17) ALETAHA D, NEOGI T, SILMAN AJ, FUNOVITS J, FELSON DT, BINGHAM CO 3RD, BIRNBAUM NS, BURMESTER GR, BYKERK VP, COHEN MD, COMBE B, COSTENBADER KH, DOUGADOS M, EMERY P, FERRACCIOLI G, HAZES JM, HOBBS K, HUIZINGA TW, KAVANAUGH A, KAY J, KVIEN TK, LAING T, MEASE P, MÉNARD HA, MORELAND LW, NADEN RL, PINCUS T, SMOLEN JS, STANISLAWSKA-BIERNAT E, SYMMONS D, TAK PP, UPCHURCH KS, VENCOVSKÝ J, WOLFE F, HAWKER G. Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League against Rheumatism collaborative initiative. Arthritis Rheum 2010; 62: 2569-2581.
- 18) FELSON DT, ANDERSON JJ, BOERS M, BOMBARDIER C, CHERNOFF M, FRIED B, FURST D, GOLDSMITH C, KIESZAK S, LIGHTFOOT R, ET AL. The American College of Rheumatology preliminary core set of disease activity measures for rheumatoid arthritis clinical trials. The Committee on Outcome Measures in Rheumatoid Arthritis Clinical Trials. Arthritis Rheum 1993; 36: 729-740.
- FUCHS HA, BROOKS RH, CALLAHAN LF, PINCUS T. A simplified twenty-eight-joint quantitative articular index in rheumatoid arthritis. Arthritis Rheum 1989; 32: 531-537.
- 20) RUSCHPLER P, LORENZ P, EICHLER W, KOCZAN D, HÄNEL C, SCHOLZ R, MELZER C, THIESEN HJ, STIEHL P. High CXCR3 expression in synovial mast cells associated with CXCL9 and CXCL10 expression in inflammatory synovial tissues of patients with rheumatoid arthritis. Arthritis Res Ther 2003; 5: R241-252.
- 21) HOEKSTRA M, VAN EDE AE, HAAGSMA CJ, VAN DE LAAR MA, HUIZINGA TW, KRUIJSEN MW. Factors associated with toxicity, final dose, and efficacy of methotrexate in patients with rheumatoid arthritis. Ann Rheum Dis 2003; 62: 423-426.
- 22) KOTRYCH D, DZIEDZIEJKO V, SAFRANOW K, DROZDZIK M, PAWLIK A. CXCL9 and CXCL10 gene polymorphisms in patients with rheumatoid arthritis. Rheumatol Int 2015; 35: 1319-1323.

3040