

Investigation of the relationships among respiratory syncytial virus infection, T cell immune response and intestinal flora

H.-T. ZHENG^{1,2,3}, Q.-Y. ZHAO^{1,2}, Y. DING^{1,2}, S.-X. MA^{1,2}, W.-X. CHEN^{1,2}, J.-L. QIU^{1,2}, X.-F. LI^{1,2}, X.-X. SUN^{1,2}, Y.-J. ZHANG⁴, B. YUAN³, Y.-B. YAN^{1,2}

¹Department of Pediatrics, The First Affiliated Hospital of Henan University of Chinese Medicine, Zhengzhou, Henan, China

²School of Pediatrics, Henan University of Chinese Medicine, Zhengzhou, Henan, China

³Department of Pediatric, Affiliated Hospital of Nanjing University of Chinese Medicine, Nanjing, Jiangsu, China

⁴Graduate School of Henan University of Chinese Medicine, Zhengzhou, Henan, China

Haitao Zheng, Yongbin Yan and Bin Yuan contributed equally to this work

Abstract. – The aim of this work was to evaluate the relationships among respiratory syncytial virus infection, T cell immune response and intestinal flora.

Peer-reviewed papers published in English were collected through extensive searches performed in PubMed, Web of Science, Google Scholar, and China National Knowledge Infrastructure databases. The articles were reviewed to extract relevant information on the immune responses of Th1/Th2 and Treg/Th17 to respiratory syncytial virus infection in the body.

RSV (Respiratory syncytial virus, RSV) infection leads to imbalance between Th1/Th2 and Treg/Th17 immune cells, resulting in Th2 or Th17 dominant immune responses, which can generate immune disorder and aggravate clinical symptoms. Intestinal micro-organisms play very important roles in maintaining stable immune environment, stimulating immune system maturation and balancing Th1/Th2 and Treg/Th17 immune systems in children. In our review of various papers from around the world, we speculated that the steady state of intestinal bacteria was disturbed after children got infected with RSV, resulting in intestinal flora disorder. Then, the imbalance between Th1/Th2 and Treg/Th17 immune cells was increased. Both intestinal flora disorder and RSV infection could cause cellular immunity imbalance of Th1/Th2 or Treg/Th17, eventually leading to disease deterioration and even a vicious cycle. Normal intestinal flora can maintain immune system stability, regulate the dynamic balance of Th1/Th2 and Treg/Th17 and prevent or mitigate adverse consequences of RSV infection.

Because probiotics can improve intestinal barrier function and regulate immune response, they can effectively be used to treat children

with recurrent respiratory tract infections. Using conventional antiviral therapy strategy supplemented with probiotics in the treatment of clinical RSV infection may be better for the body.

Key Words:

Respiratory syncytial virus, T-helper 1/T-helper 2, Regulatory T cells/T-helper17, Intestinal flora, Cellular immunity.

Introduction

As the most common pathogen of viral pneumonia among children, respiratory syncytial virus (RSV) can cause interstitial pneumonia and bronchiolitis^{1,2}. Based on statistics, more than 30 million children < 5 years old are infected with RSV in the world each year, of which 3-4 million need hospitalization and 0.1-0.2 million hospitalized children experience serious adverse consequences and even death^{1,3}. 70% of children are infected with RSV at the age of about one year and 100% have RSV infection in the age range of 2 to 3^{2,4,5}. Although most children present only mild upper respiratory symptoms, 2-5% develop severe bronchiolitis and require further treatment⁵⁻⁷. In addition, these children have a high risk of repeated wheezing and asthma^{8,9}. Reasonable treatment strategies for children with RSV infection are of great significance in the efficacy and prognosis of therapy.

Immune responses help the immune system of body to identify themselves, eliminate alienation and maintain the balance of body environment.

The physiological function of the immune system is mainly fulfilled by immunizing lymphocytes. Immune response is also described as the process of antigen recognition by immune lymphocytes, based on their own activation, proliferation and differentiation to exert their effect. In immune response, immunogen only contribute to the selection and triggering of immune lymphocytes. During antigen recognition, immune lymphocytes are activated to form B cell-mediated humoral immunity and T cell-mediated cellular immunity, and are mediated in non-activated state to form immune tolerance^{10,11}.

As a research hot topic in recent years, gut microbiota greatly affects human health mainly in regulating digestion, physiology, nutrition and immunity. Imbalance of intestinal flora can disturb the regulation of the immune system of body, damage intestinal mucosal barrier, and induce chronic inflammatory diseases. Different gut bacteria might be involved in the differentiation and functional regulation of various immune cell subsets. It is believed that RSV not only causes lung inflammation, imbalances the development of T cells in lungs and releases related inflammatory agents, but also affects intestinal immune cell differentiation and intestinal flora homeostasis. Therefore, this paper reviews the relationships among the mechanisms of RSV infection, T cell immune response, and intestinal flora, and provides a theoretical reference for clinical prevention and treatment of RSV, as well as new drug development.

In this research, a detailed review of published studies was performed on the works published in PubMed, Web of Science, Google Scholar, and China National Knowledge Infrastructure databases using RSV as a search term. Considering technological progress and recent reports, focus was placed on the last 30 years and articles published from January 1st, 1992, to January 1st, 2022, were considered. The search returned 176,262 articles. Peer-reviewed papers published in English language were adopted with no restrictions in terms of article type and geography. Papers related to RSV infection, T cell immune responses and intestinal flora were retrieved. To complement this review, electronic search was supplemented with a manual search of the references of these papers. The papers extracted from electronic databases were screened by reading their titles and abstracts and the full texts of those considered likely to be relevant were retrieved. The retrieved papers were reviewed and critically analyzed.

RSV Infection and T Cell Immune Response

T cell-mediated immunity (CMI) is also known as cellular immune response. Initial T cells (naive T cell, T_n), also known as non-sensitized T cells, migrate to peripheral lymphoid tissue after maturation in thymus. T_n cells are gradually differentiated into Th1, Th2, Th17, regulatory T (Treg), effector T and similar types of cells through antigenic stimulation or dendritic cell delivery. Th cells are mainly divided into Th1 and Th2 cells¹¹. Th1 cells, so-called inflammatory T cells, secrete interleukin-2 (IL-2), interferon- α (IFN- α), IFN- γ , tumor necrosis factor- β (TNF- β), etc. These substances mediate cytotoxicity and local inflammation-related immune responses, assist antibody production, participate in cellular immunity, delay hypersensitivity and play an important role in body anti-intracellular pathogen infection^{11,12}. Th2 cells mainly secrete IL-4, IL-5, IL-6, IL-10 and IL-13, which help the differentiation of B cells into antibody-secreting cells, regulate humoral immune response and play a key role in the induction of allergic reactions^{11,13}. Treg cells are a subpopulation of T cells with immuno-suppressive function. The most important function of Treg cells is the suppressing the activation and survival of effector T cells, avoiding tissue damage and maintaining the immune balance of body by eliminating IL-2 and producing IL-10 and TGF- β during anti-infective immune responses¹⁴⁻¹⁶. Th17 cells are specific pro-inflammatory cells that produce IL-17, and their main function is the stimulation of inflammatory responses, enhancement of the acquired cellular immune responses, and resisting bacterial, fungal and viral infections^{17,18}. IL-17 can also induce pro-inflammatory cytokine and chemokine expression. The activation of IL-17 receptor, which is extensively present in epithelial cells, endothelial cells, monocytes and macrophages, induces infiltration and destruction in tissue cells¹⁹. The coordination of various T cells plays a critical role in body protection against external stimuli.

RSV-infected children can cause cellular immune responses and imbalance Th1 and Th2 immune responses, especially Th2-dominant immune responses. The severity of clinical symptoms is correlated with increased degree of Th2 response^{20,21}. Animal experiments showed that in T cell immune response, T_n cells are induced to differentiate into Th1 cells by RSV virus F protein and Th2 by RSV virus G protein^{22,23}. NS1 protein of RSV virus can inhibit Th1, Th2 and

Th17 cell differentiations, while NS2 protein inhibits Th2 and Th17 cell differentiations¹⁶. Th1 response could produce IFN- γ , TNF- α , IL-2 and activate cytotoxic T and NK cells to promote effective virus removal, which is the most expected auxiliary T cell response mode after RSV infection^{2,20}. However, Th2 cells secrete IL-4 and IL-5 which arise obvious and unfavorable inflammatory responses after re-infection with RSV.

Furthermore, Th1 and Th2 can restrain and lower the activation level of each other²⁴. Thus, Th2 superior immune response after RSV infection decreases the efficiency of virus removal increasing its damage to body^{2,25,26}. Clinical examination²⁷ revealed that newborns re-infected with RSV generated Th2-dominant immune responses, but this phenomenon is not generally observed in adults. Detecting sera and nasopharyngeal aspirate samples in RSV-infected children increased IL-4 concentration and decreased IFN- γ concentration, suggesting Th2-dominant immune response^{20,26,28}. A correlation was also found²⁶ between IL-4/IFN- γ ratio and disease severity. In 1960s, the trial of FI-RSV vaccine failed since the vaccine not only could not prevent infants from being infected by RSV, but also aggravated and worsened original disease²⁹. In FI-RSV-vaccinated cases, immune response was Th2-dominant response and Th1/Th2 ratio was one of the important indicators in measuring the severity of clinical symptoms after RSV infection²⁵. It should be noted that, using Th1/Th2 balance to explain the pathogenesis of RSV infection is not absolute. For example, in the severe infection of RSV-induced bronchiolitis various types of immune cells exist.

Disequilibrium of Treg and Th17 immune cells leads to inflammation and autoimmune diseases^{17,30}. Treg cells differentiation was relatively weakened and Th17 cell differentiation was increased, resulting in intensified inflammation response after RSV infection. Research³¹⁻³³ on RSV-infected mice showed that Treg cells promoted RSV clearance and significantly reduced inflammation degree in mice lungs³⁴. Treg cells can modulate immune microenvironment and avoid excessive inflammatory T cell responses, including the inhibition of Th2-dominant immune responses³⁵. The number of Treg cells was decreased and that of Th17 cells was increased in IL-10 knock out mice³⁶. After IL-10 knock out RSV-infected mice, more severe disease development, increased pro-inflammatory and chemokines levels and increased pathological changes in lungs were observed^{34,36}.

Breakage between Th1/Th2 or Treg/Th17 also affected immune response to Treg/Th17 or Th1/Th2. In RSV-infected mice, the IL-10 produced by Treg cells can control inflammatory responses to ensure effective virus removal, maintain appropriate immune level of the body and reduce clinical symptoms by inhibiting Th2-based immune responses^{31,35,37}. Th17 cells and IL-17 produced by them enhance Th2 response, preventing effective virus removal and enhancing inflammatory responses, eventually leading to more serious clinical symptoms³⁸. In addition, the IL-17 produced by Th17 cells could up-regulate the level of Th2 cytokines in Th2-dominant immune environments, demonstrating that Th17 immune response is coordinated with Th2 immune response^{38,39}. Wang et al⁴⁰ showed that OVA (Ovalbumin, OVA)-sensitized mouse asthma model had increased Th17 and insufficient Treg immune responses, then induced Th2-mediated respiratory inflammation.

The imbalance of Th1/Th2 and Treg/Th17 immune cells resulted in corresponding clinical phenotypes. Th1/Th2 and Treg/Th17 have a complex and intimate relationship. The imbalance of any system can lead to immune system disorders, eventually resulting in the occurrence of diseases.

Intestinal Flora and Immunity

Intestinal flora is a stable microbial community in the intestine of host by long-term evolution, capable of resisting pathogen stimulation and invasion and regulating a series of physiological and metabolic processes⁴¹. Normal intestinal flora and its metabolites contain a great number of immune stimuli including antigens, toxins, etc., while intestinal immune barrier can generate appropriate responses to different antigens from intestinal mucosal surface such as immune clearance and rejection to pathogens. Research has shown⁴² that intestinal mucosa could induce immune responses of different CD4⁺ T cell subsets after colonization and formation of Treg, Th1, Th2, Th17 and T follicular helper (Tfh) cells, revealing the accurate regulatory mechanism of intestinal immunization. Intestinal flora not only is important to the development and activation of intestinal mucosal immune system, but also is significant for extra-intestinal immune system. Constant stimulation of immune system by normal intestinal microbes is required for its maturation. For example, lactic acid bacteria could stimulate the

secretion of IL-12, IL-18, IFN- γ and other cytokines by peripheral blood cells⁴³. Nicaise et al⁴⁴ also suggested that intact intestinal flora was the basis for IL-12 production in spleen, which was an important link to innate immunity and acquired immunity. IL-12 effectively improves cellular immune defense function and promotes the differentiation of CD4⁺ T cells into Th1 cells. In recent years, research⁴⁵⁻⁴⁷ on intestinal microflora effects on immune environment stability has made great progress. Host microbiota can participate in adaptive modulation of intestinal mucosa to maintain normal intestine state while suppressing immune response, resulting in the generation of inflammatory diseases. Furthermore, microbial composition could affect the susceptibility of immune-mediated diseases, such as autoimmune and allergic diseases⁴⁸⁻⁵⁰. Intestinal flora could regulate the reactivity of immune system through immune cells and vice versa. Kawamoto et al⁵¹ showed that intestinal immunosuppressive receptor PD-1 affected the composition of intestinal flora. Under such conditions, intestinal bacteria lose control resulting in excessive breeding and bacterial translocation when intestinal immune activity is reduced. That is, intestinal immune system is inextricably related to systemic immune system.

Flora Imbalance and T Cell Immune Response

The type, number and distribution of intestinal flora are not constant and are affected by several factors such as eating habits, diseases and drugs⁵². Under normal circumstances, intestinal flora and host are in a dynamic equilibrium state through precise regulatory mechanism. When the number, type and proportion of normal intestinal flora are changed, the original stable state of intestinal flora is broken causing flora imbalance^{41,53}. The main reason for flora imbalance is irrational application of antibiotics, especially broad-spectrum antibiotics. Experiments have revealed the correlation between intestinal flora imbalance due to the application of antibiotics and occurrence of immune diseases^{41,53}. Antibiotics-induced intestinal flora imbalance can generate Th2 and Th17 cell-dominant immune responses and lead to the deficiency of Treg cells immune response⁵⁴. Therefore, antibiotic-induced intestinal flora imbalance might be a risk factor for asthma and other lung diseases. Immune system could be regulated by probiotics through balancing Th1/Th2 ratio⁵⁵. For example,

Lactobacillus pentosus produces IL-10 exerting an anti-allergic effect by regulating Th1/Th2 balance⁵⁶. Studies⁵⁷⁻⁵⁹ have shown that application of probiotics to newborn asthma mice induced the generation of Treg cells, inhibiting allergic and respiratory diseases. Normal intestinal flora could induce Th1 immune response and inhibit Th2 immune response, reducing the occurrence of allergic diseases. However, intestinal flora imbalance can prevent the maturation of Th1 cells and promote Th2 cell differentiation increasing the risk of respiratory allergies and infectious diseases. Therefore, probiotics can prevent inflammation by retrieving immune imbalance^{43,60}.

The relationships among RSV, T cell immune response and intestinal flora are shown in Figure 1.

Discussion

Based on the above discussion, it was concluded that intestinal microflora possessed regulatory ability to develop and activate immune system. If the normal colonization process of neonatal intestinal flora is damaged, abnormal reactions occur causing lung allergic immune response when exposed to allergens through ingestion or inhalation^{61,62}. Children, especially infants and young children, are prone to the establishment of normal microbes. During these periods, the flora with poor stability and diversity are fragile and susceptible to various unwanted factors. However, the establishment of normal microbes is closely affected by some important physiological functions, such as development and maturation of immune, metabolism, and nutrition systems. If the formation of normal microbes is disturbed during this period, acute and chronic diseases and even some adult diseases could occur. Both intestinal flora disorder and RSV infection cause cellular immunity imbalance in Th1/Th2 and Treg/Th17, eventually resulting in disease deterioration and even a vicious cycle. Normal intestinal flora can maintain the stability of immune system, regulate Th1/Th2 and Treg/Th17 dynamic balance and prevent or mitigate adverse consequences of RSV infection. Therefore, application of conventional antiviral therapy strategies supplemented with probiotics to clinical RSV infection treatment may achieve better treatment results.

Conclusions

Intestinal dysbiosis can aggravate RSV airway inflammation by altering local immune

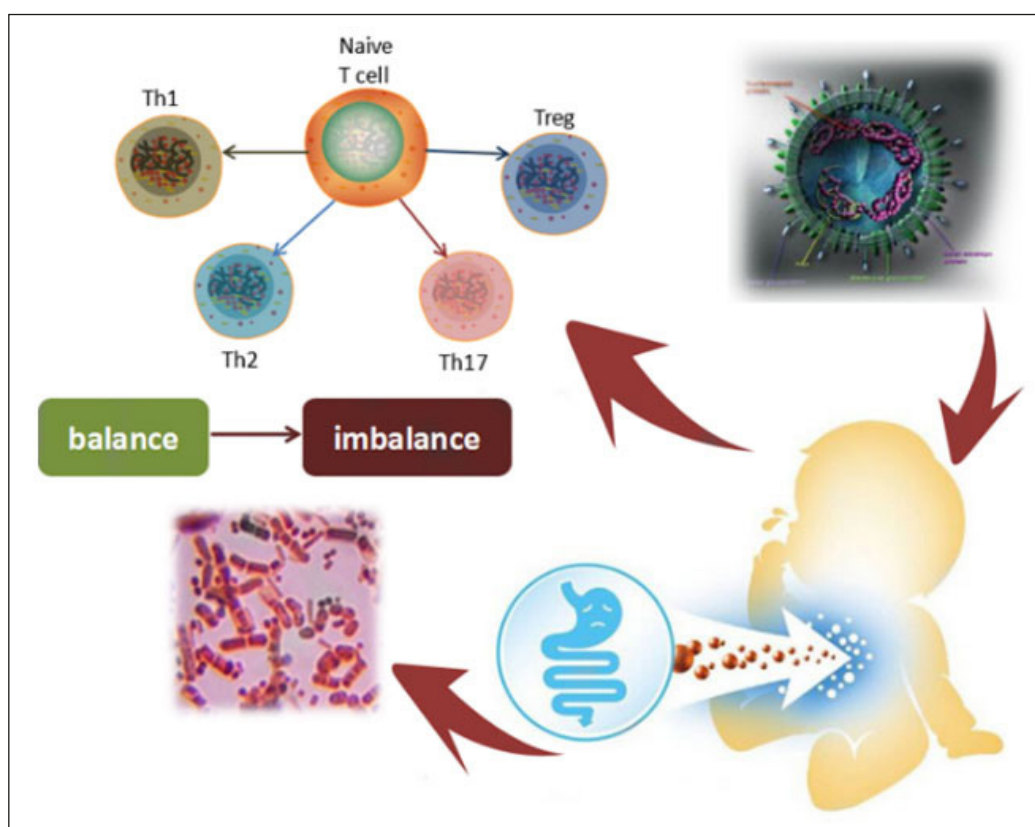


Figure 1. The relationships among RSV, T cell immunity and intestinal flora.

response in lungs. Supplementation with beneficial bacteria can restore immune dysbiosis induced by intestinal dysbiosis and reduce RSV airway inflammation.

Conflict of Interest

The authors declared that they have no conflicts of interest to this work.

Acknowledgements

We thank Xiaona Hu (Henan Provincial Hospital of Chinese Medicine, China) for reading this manuscript and providing helpful feedback.

Funding

This work was supported by grants from National Natural Science Foundation of China (NSFC) (Grant No. 81574025, 81973903, 82174438), the Henan of Chinese Medicine Special Research Project (Grant No. 2019JDZX2031).

Informed Consent

Not applicable.

Authors' Contribution

HTZ YBY and BY participated in study conception, design, and preparation of the manuscript. QYZ, YD, SXM, WXC, JLQ, XFL, XXS and YJZ reviewed the manuscript. HTZ YBY and BY revised the manuscript and coordinated the whole project. All authors read and reviewed the final manuscript.

ORCID ID

H.-T. Zheng: 0000-0003-3160-2082

References

- 1) Paluck A, Osan J, Hollingsworth L, Talukdar SN, Saegh AA, Mehedi M. Role of ARP2/3 Complex-Driven Actin Polymerization in RSV Infection. *Pathogens* 2021; 11: 26.
- 2) Ananworanich J, Heaton PM. Bringing Preventive RSV Monoclonal Antibodies to Infants in Low-and Middle-Income Countries: Challenges and Opportunities. *Vaccines (Basel)* 2021; 9: 961.
- 3) Rossey I, Gilman MS, Kabeche SC, Sedeyn K, Wrapp D, Kanekiyo M, Chen M, Mas V, Spitaels J,

- Melero JA, Graham BS, Schepens B, McLellan JS, Saelens X. Potent single-domain antibodies that arrest respiratory syncytial virus fusion protein in its prefusion state. *Nat Commun* 2017; 8: 14158.
- 4) Di Mattia G, Nenna R, Mancino E, Rizzo V, Pierangeli A, Villani A, Midulla F. During the COVID-19 pandemic where has respiratory syncytial virus gone? *Pediatr Pulmonol* 2021; 56: 3106-3109.
 - 5) Chatterjee A, Mavunda K, Krilov LR. Current State of Respiratory Syncytial Virus Disease and Management. *Infect Dis Ther* 2021; 10 (Suppl 1): 5-16.
Meng J, Stobart CC, Hotard AL, Moore ML. An overview of respiratory syncytial virus. *PLoS Pathog* 2014; 10: e1004016.
 - 6) Rodriguez R, Ramilo O. Respiratory syncytial virus: how, why and what to do. *J Infect* 2014; 68 Suppl 1: S115-118.
 - 7) Borchers AT, Chang C, Gershwin ME, Gershwin LJ. Respiratory syncytial virus--a comprehensive review. *Clin Rev Allergy Immunol* 2013; 45: 331-379.
 - 8) Piedimonte G, Perez MK. Alternative mechanisms for respiratory syncytial virus (RSV) infection and persistence: could RSV be transmitted through the placenta and persist into developing fetal lungs? *Curr Opin Pharmacol* 2014; 16: 82-88.
 - 9) Simoes EA, Carbonell-Estrany X, Rieger CH, Mitchell I, Fredrick L, Groothuis JR, Palivizumab Long-Term Respiratory Outcomes Study G. The effect of respiratory syncytial virus on subsequent recurrent wheezing in atopic and nonatopic children. *J Allergy Clin Immunol* 2010; 126: 256-262.
 - 10) Roozendaal R, Mebius RE. Stromal cell-immune cell interactions. *Annu Rev Immunol* 2011; 29: 23-43.
 - 11) Akdis M, Akdis CA. Therapeutic manipulation of immune tolerance in allergic disease. *Nat Rev Drug Discov* 2009; 8: 645-660.
 - 12) O'Garra A, Vieira P. T(H)1 cells control themselves by producing interleukin-10. *Nat Rev Immunol* 2007; 7: 425-428.
 - 13) Pulendran B, Artis D. New paradigms in type 2 immunity. *Science* 2012; 337: 431-435.
 - 14) Shalev I, Schmelzle M, Robson SC, Levy G. Making sense of regulatory T cell suppressive function. *Semin Immunol* 2011; 23: 282-292.
 - 15) Afzali B, Mitchell P, Lechler RI, John S, Lombardi G. Translational mini-review series on Th17 cells: induction of interleukin-17 production by regulatory T cells. *Clin Exp Immunol* 2010; 159: 120-130.
 - 16) O'Connor RA, Taams LS, Anderton SM. Translational mini-review series on Th17 cells: CD4 T helper cells: functional plasticity and differential sensitivity to regulatory T cell-mediated regulation. *Clin Exp Immunol* 2010; 159: 137-147.
 - 17) Kimura A, Kishimoto T. IL-6: regulator of Treg/Th17 balance. *Eur J Immunol* 2010; 40: 1830-1835.
 - 18) Crome SQ, Wang AY, Levings MK. Translational mini-review series on Th17 cells: function and regulation of human T helper 17 cells in health and disease. *Clin Exp Immunol* 2010; 159: 109-119.
 - 19) Awasthi A, Kuchroo VK. Th17 cells: from precursors to players in inflammation and infection. *Int Immunol* 2009; 21: 489-498.
 - 20) Pinto RA, Arredondo SM, Bono MR, Gaggero AA, Diaz PV. T helper 1/T helper 2 cytokine imbalance in respiratory syncytial virus infection is associated with increased endogenous plasma cortisol. *Pediatrics* 2006; 117: e878-886.
 - 21) Legg JP, Hussain IR, Warner JA, Johnston SL, Warner JO. Type 1 and type 2 cytokine imbalance in acute respiratory syncytial virus bronchiolitis. *Am J Respir Crit Care Med* 2003; 168: 633-639.
 - 22) Bembridge GP, Garcia-Beato R, Lopez JA, Melero JA, Taylor G. Subcellular site of expression and route of vaccination influence pulmonary eosinophilia following respiratory syncytial virus challenge in BALB/c mice sensitized to the attachment G protein. *J Immunol* 1998; 161: 2473-2480.
 - 23) Johnson TR, Graham BS. Secreted respiratory syncytial virus G glycoprotein induces interleukin-5 (IL-5), IL-13, and eosinophilia by an IL-4-independent mechanism. *J Virol* 1999; 73: 8485-8495.
 - 24) Raveh D, Kruskal BA, Farland J, Ezekowitz RA. Th1 and Th2 cytokines cooperate to stimulate mannose-receptor-mediated phagocytosis. *J Leukoc Biol* 1998; 64: 108-113.
 - 25) Becker Y. Respiratory syncytial virus (RSV) evades the human adaptive immune system by skewing the Th1/Th2 cytokine balance toward increased levels of Th2 cytokines and IgE, markers of allergy--a review. *Virus Genes* 2006; 33: 235-252.
 - 26) Hassan MA, Eldin AM, Ahmed MM. T-helper2 /T-helper1 imbalance in respiratory syncytial virus bronchiolitis in relation to disease severity and outcome. *Egypt J Immunol* 2008; 15: 153-160.
 - 27) Dulek DE, Newcomb DC, Toki S, Goliniewska K, Cephus J, Reiss S, Bates JT, Crowe JE Jr, Boyd KL, Moore ML, Zhou W, Peebles RS Jr. STAT4 deficiency fails to induce lung Th2 or Th17 immunity following primary or secondary respiratory syncytial virus (RSV) challenge but enhances the lung RSV-specific CD8+ T cell immune response to secondary challenge. *J Virol* 2014; 88: 9655-9672.
 - 28) Bendelja K, Gagro A, Bace A, Lokar-Kolbas R, Krsulovic-Hresic V, Drazenovic V, Mlinaric-Galinovic G, Rabatic S. Predominant type-2 response in infants with respiratory syncytial virus (RSV) infection demonstrated by cytokine flow cytometry. *Clin Exp Immunol* 2000; 121: 332-338.
 - 29) Loebbermann J, Durant L, Thornton H, Johansson C, Openshaw PJ. Defective immunoregulation in RSV vaccine-augmented viral lung disease restored by selective chemoattraction of regulatory T cells. *Proc Natl Acad Sci USA* 2013; 110: 2987-2992.
 - 30) Heylen M, Ruysers NE, Gielis EM, Vanhomwegen E, Pelckmans PA, Moreels TG, De Man JG, De Winter BY. Of worms, mice and man: an overview of experimental and clinical helminth-based therapy for inflammatory bowel disease. *Pharmacol Ther* 2014; 143: 153-167.

- 31) Fulton RB, Meyerholz DK, Varga SM. Foxp3+ CD4 regulatory T cells limit pulmonary immunopathology by modulating the CD8 T cell response during respiratory syncytial virus infection. *J Immunol* 2010; 185: 2382-2392.
- 32) Lee DC, Harker JA, Tregoning JS, Atabani SF, Johansson C, Schwarze J, Openshaw PJ. CD25+ natural regulatory T cells are critical in limiting innate and adaptive immunity and resolving disease following respiratory syncytial virus infection. *J Virol* 2010; 84: 8790-8798.
- 33) Ruckwardt TJ, Bonaparte KL, Nason MC, Graham BS. Regulatory T cells promote early influx of CD8+ T cells in the lungs of respiratory syncytial virus-infected mice and diminish immunodominance disparities. *J Virol* 2009; 83: 3019-3028.
- 34) Loebbermann J, Schnoeller C, Thornton H, Durant L, Sweeney NP, Schuijs M, et al. IL-10 regulates viral lung immunopathology during acute respiratory syncytial virus infection in mice. *PLoS One* 2012; 7: e32371.
- 35) Durant LR, Makris S, Voorburg CM, Loebbermann J, Johansson C, Openshaw PJ. Regulatory T cells prevent Th2 immune responses and pulmonary eosinophilia during respiratory syncytial virus infection in mice. *J Virol* 2013; 87: 10946-10954.
- 36) Weiss KA, Christiaansen AF, Fulton RB, Meyerholz DK, Varga SM. Multiple CD4+ T cell subsets produce immunomodulatory IL-10 during respiratory syncytial virus infection. *J Immunol* 2011; 187: 3145-3154.
- 37) Loebbermann J, Thornton H, Durant L, Sparwasser T, Webster KE, Sprent J, Culley FJ, Johansson C, Openshaw PJ. Regulatory T cells expressing granzyme B play a critical role in controlling lung inflammation during acute viral infection. *Mucosal Immunol* 2012; 5: 161-172.
- 38) Mukherjee S, Lindell DM, Berlin AA, Morris SB, Shanley TP, Hershenson MB, Lukacs NW. IL-17-induced pulmonary pathogenesis during respiratory viral infection and exacerbation of allergic disease. *Am J Pathol* 2011; 179: 248-258.
- 39) Mangoldt TC, Van Herck MA, Nullens S, Ramet J, De Dooy JJ, Jorens PG, De Winter BY. The role of Th17 and Treg responses in the pathogenesis of RSV infection. *Pediatr Res* 2015; 78: 483-491.
- 40) Wang J, Kong L, Luo Q, Li B, Wu J, Liu B, Wu X, Dong J. Dual effects of respiratory syncytial virus infections on airway inflammation by regulation of Th17/Treg responses in ovalbumin-challenged mice. *Inflammation* 2014; 37: 1984-2005.
- 41) Noverr MC, Falkowski NR, McDonald RA, McKenzie AN, Huffnagle GB. Development of allergic airway disease in mice following antibiotic therapy and fungal microbiota increase: role of host genetics, antigen, and interleukin-13. *Infect Immun* 2005; 73: 30-38.
- 42) Shale, M., Schiering, C. and Powrie, F. CD4(+) T-cell subsets in intestinal inflammation. *Immunol Rev* 2013; 252: 164-82.
- 43) Miettinen M, Matikainen S, Vuopio-Varkila J, Pirhonen J, Varkila K, Kurimoto M, Julkunen I. Lactobacilli and streptococci induce interleukin-12 (IL-12), IL-18, and gamma interferon production in human peripheral blood mononuclear cells. *Infect Immun* 1998; 66: 6058-6062.
- 44) Nicaise P, Gleizes A, Sandre C, Kergot R, Lebrech H, Forestier F, Labarre C. The intestinal microflora regulates cytokine production positively in spleen-derived macrophages but negatively in bone marrow-derived macrophages. *Eur Cytokine Netw* 1999; 10: 365-372.
- 45) Abreu NA, Nagalingam NA, Song Y, Roediger FC, Pletcher SD, Goldberg AN, Lynch SV. Sinus microbiome diversity depletion and *Corynebacterium tuberculo-stearicum* enrichment mediates rhinosinusitis. *Sci Transl Med* 2012; 4: 151ra124.
- 46) Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, Cheng G, Yamasaki S, Saito T, Ohba Y, Taniguchi T, Takeda K, Hori S, Ivanov II, Umesaki Y, Itoh K, Honda K. Induction of colonic regulatory T cells by indigenous *Clostridium* species. *Science* 2011; 331: 337-341.
- 47) Hansen CH, Nielsen DS, Kverka M, Zakostelska Z, Klimesova K, Hudcovic T, Tlaskalova-Hogenova H, Hansen AK. Patterns of early gut colonization shape future immune responses of the host. *PLoS One* 2012; 7: e34043.
- 48) Mathis D, Benoist C. The influence of the microbiota on type-1 diabetes: on the threshold of a leap forward in our understanding. *Immunol Rev* 2012; 245: 239-249.
- 49) Scher JU, Abramson SB. The microbiome and rheumatoid arthritis. *Nat Rev Rheumatol* 2011; 7: 569-578.
- 50) Hanski I, von Hertzen L, Fyhrquist N, Koskinen K, Torppa K, Laatikainen T, Karisola P, Auvinen P, Paulin L, Mäkelä MJ, Vartiainen E, Kosunen TU, Alenius H, Haahtela T. Environmental biodiversity, human microbiota, and allergy are interrelated. *Proc Natl Acad Sci USA* 2012; 109: 8334-8339.
- 51) Kawamoto S, Tran TH, Maruya M, Suzuki K, Doi Y, Tsutsui Y, Kato LM, Fagarasan S. The inhibitory receptor PD-1 regulates IgA selection and bacterial composition in the gut. *Science* 2012; 336: 485-489.
- 52) Thaiss CA, Zmora N, Levy M, Elinav E. The microbiome and innate immunity. *Nature* 2016; 535: 65-74.
- 53) Noverr MC, Noggle RM, Toews GB, Huffnagle GB. Role of antibiotics and fungal microbiota in driving pulmonary allergic responses. *Infect Immun* 2004; 72: 4996-5003.
- 54) Russell SL, Gold MJ, Hartmann M, Willing BP, Thorson L, Wlodarska M, Gill N, Blanchet MR, Mohn WW, McNagny KM, Finlay BB. Early life antibiotic-driven changes in microbiota enhance susceptibility to allergic asthma. *EMBO Rep* 2012; 13: 440-447.
- 55) Ghadimi D, Folster-Holst R, de Vrese M, Winkler P, Heller KJ, Schrezenmeier J. Effects of probiotic bacteria and their genomic DNA on TH1/TH2-cytokine production by peripheral blood mononuclear cells (PBMCs) of healthy and allergic subjects. *Immunobiology* 2008; 213: 677-692.

- 56) Nonaka Y, Izumo T, Izumi F, Maekawa T, Shibata H, Nakano A, Kishi A, Akatani K, Kiso Y. Antiallergic effects of *Lactobacillus pentosus* strain S-PT84 mediated by modulation of Th1/Th2 immunobalance and induction of IL-10 production. *Int Arch Allergy Immunol* 2008; 145: 249-257.
- 57) Feleszko W, Jaworska J, Rha RD, Steinhausen S, Avagyan A, Jaudszus A, Ahrens B, Groneberg DA, Wahn U, Hamelmann E. Probiotic-induced suppression of allergic sensitization and airway inflammation is associated with an increase of T regulatory-dependent mechanisms in a murine model of asthma. *Clin Exp Allergy* 2007; 37: 498-505.
- 58) Karimi K, Inman MD, Bienenstock J, Forsythe P. *Lactobacillus reuteri*-induced regulatory T cells protect against an allergic airway response in mice. *Am J Respir Crit Care Med* 2009; 179: 186-193.
- 59) Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, Nakanishi Y, Uetake C, Kato K, Kato T, Takahashi M, Fukuda NN, Murakami S, Miyachi E, Hino S, Atarashi K, Onawa S, Fujimura Y, Lockett T, Clarke JM, Topping DL, Tomita M, Hori S, Ohara O, Morita T, Koseki H, Kikuchi J, Honda K, Hase K, Ohno H. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 2013; 504: 446-450.
- 60) Oyama N, Sudo N, Sogawa H, Kubo C. Antibiotic use during infancy promotes a shift in the T(H)1/T(H)2 balance toward T(H)2-dominant immunity in mice. *J Allergy Clin Immunol* 2001; 107: 153-159.
- 61) Li CX, Liu HY, Lin YX, Pan JB, Su J. The Gut Microbiota and Respiratory Diseases: New Evidence. *J Immunol Res* 2020; 31; 2020: 2340670.
- 62) Liu BG, Xie M, Dong Y, Wu H, He DD, Hu GZ, Xu EP. Antimicrobial mechanisms of traditional Chinese medicine and reversal of drug resistance: a narrative review. *Eur Rev Med Pharmacol Sci* 2022; 26: 5553-5561.