Letter to the Editor

Transparency of disinfectant and hand sanitizer contents in the context of COVID-19

Dear Editor,

Soave et al¹ have provided a review of disinfectant exposure during the COVID-19 pandemic and related concerns. One of the practical aspects for infection control of COVID-19 is the role that the environmental presence of virus may play in transmission from whatever source and under whatever conditions. Spread from several sources is an accepted mode of transmission albeit it has proven challenging to quantitate the same comparatively². In this regard, it is yet imperative that disinfection and hand sanitization should garner careful consideration in healthcare settings and in the environment of those infected. Many countries have established simplified protocols for efficacy and suitability of disinfectants and hand sanitizers. Public postings are often used by industry, scientific communities, and public to determine applications or to better understand the science. Any presumed approval by regulatory authorities has the tendency to make the community believe that products should be efficacious even if endorsement is waived.

A web-based national listing of hard-surface disinfectants and hand sanitizers in Canada exemplifies some issues³. Those for approval are designated with drug identification numbers. The listing so made is also said to be effective for SARS-CoV-2. When first reviewed in May 2020, 414 products were listed, and 60.1% were deemed to have quaternary ammonium compounds as sole active ingredients. Another 8.7% included quaternary ammonium compounds with other concomitant active agents. Other active agents included alcohols, peroxides, phenolics, citric acid, chlorhexidine, hydrogen peroxide, acetic acid, peracetic acid, chlorine dioxide, sodium hypochlorite, hypochlorous acid, hydrochloric acid, lactic acid, silver dihydrogen citrate, thymol, sodium dichloroisocyanurate, and potassium peroxymonosulfate. For 21/414 (5.1%), published scientific evidence of efficacy against one or more coronaviruses was lacking.

At the same time, a smaller representative sample of 58 (90% confidence level \pm 10%) was chosen with random numbers for further analysis. For 3/58 (5.1%), no data were accessible online. Of the remaining, 36/58 (62.1%) were available in liquid containers for direct use or dilution, 8/58 (13.8%) were surface wipes, and 11/58 (19%) were sprays. None of the products listed working temperatures. Ready-to-use status was apparent for one-half. For nine solutions, there was either no access data or no working dilutions listed on instruction or material safety data sheets. Product pH was commonly stated – alkaline, acidic, and neutral pH ranges for 27/58 (46.6%), 1/58 (1.7%), and 19/58 (32.8%). Of those with neutral pH, 16/19 (84.2%) were viewed as only containing quaternary ammonium-based disinfectants. Alcohol ethoxylates or other surfactants and EDTA were occasionally found as quantitatively significant constituents although the database did not cite them. In December 2020, 551 such products were listed, and a sample of 77 (90% confidence level \pm 10%) were randomly assessed in similar fashion. For 12/77 (15.6%), again no reliable data were accessible on-line. The frequency of formulations, ready-to-use status, temperature use indications, and pH variation were essentially the same in comparison to the first sample assessment.

Disinfectants and decontamination agents have been assessed for coronaviruses generally⁴. Higher temperature and adverse pH favor inactivation. Working diluent temperatures are not usually detailed, but temperatures at or above room temperature theoretically favor better activity albeit high extremes of temperature raise concern over chemical volatility. Most adverse pH were alkaline and usually greater than 10. In combination with disinfectants, adverse pH would theoretically add another measure of presumed efficacy. Neutral quaternary ammonium or phenolic products have raised some concern for lesser efficacy.

Potential for SARS-CoV-2 presence in the environment of patients is now clearly evident⁵⁻⁹. This is not surprising given the same for other respiratory viruses¹⁰. Most of the studies used RNA amplification for virus detection among environmental samples and thus will have overstated the presence of putative infectious virus. Nevertheless, some have found that the environmental burden of virus increases with patient disease severity and thus patterns of and burdens for dissemination have variable influence¹¹. Environmental contamination can occur from symptomatic or asymptomatic patients¹². Public domains may also be at risk¹³. Viable virus can remain on surfaces even when dessicated or under various adverse conditions^{4,14}. Survival on human skin has also been modeled¹⁵.

Experimental assessments of disinfectants and antiseptics are at best estimates, and it is often difficult to simulate variability that will occur in real-life applications⁴. Variability is influenced by virus species, temperature, relative humidity, contact time, concomitant organic load, viral load, precision of disinfectant stock dilution, effects of mechanical lavage, nature of the environmental surface, and inactivation of the test agent by the materials. In situ field assessments are uncommon. Several assessments have examined non-SARS-CoV-2 coronaviruses. For cleaning and disinfection (quaternary ammonium compounds and ethanol ethoxylates mixture – pH 9) of toys in a daycare nursery, viral load was determined with quantitative amplification¹⁶. RNA of several respiratory viruses was reduced, but coronavirus RNA was stable regardless of control or intervention group. Others assessed daily morning interventions with a cleaner (an anionic surfactant and ethanol ethoxylate mixture – unknown pH) in a university classroom¹⁷. Although there was a reduction in cultivable virus counts after one week, coronavirus 229E remained viable on environmental surfaces. Whereas most disinfecting or antiseptic agents are associated with benefit if not only from the mechanical application and removal, there are also product-specific ingredients which may vary in efficacy¹⁸.

SAR-CoV-2-specific publications have emerged and several themes prevail¹⁹⁻³⁰. Most such studies have also used viral RNA as the determinant for outcome, and success has been achieved in either lowering the burden of viral RNA or eliminating its detection altogether. Frequencies of disinfection and their timings have been variable as have concentrations of the same chemical. There have been diverse products used, but complex solutions have generally been more efficacious. The greater the intensity of disinfection efforts, the greater the reduction of environmental viral RNA. Full product disclosure is often missing, and it is clear that studies would benefit from assessing the full composite of a commercial product in its prescribed working dilution.

Variability in studies and outcomes raises risks for scientist and consumer. Safety is also a potential concern with some products. Studies referable to coronaviruses provide a measure of confidence but may exclude other microbial pathogens that products should affect. More consistency is required for creating measurable standards. From a consumers' perspectives, more information, ease of application, and practical efficacy are desirable. In the least, commercial products should list their active ingredients, pH, recommended temperatures for use, working dilutions, and application standards. As well, electronic media provide ample opportunity for safety data sheets to be widely accessible. In the current environment with COVID-19, there is considerable potential for trial studies of efficacy.

Conflict of Interest

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References

- Soave PM, Grasso S, Oliva A, Romano B, Di Stasio E, Dominici L, Pascali V, Antonelli M. Household disinfectant exposure during the COVID-19 pandemic: a retrospective study of the data from an Italian poison control center. Eur Rev Med Pharmacol Sci 2021; 25: 1738-1742.
- 2) Cimolai N. The semantics of airborne microbial spread and environmental relevance: back to Anderson and Cox. Environ Res 2021; 193: 110448.
- 3) Health Canada, Government of Canada. Hard-surface disinfectants and hand sanitizers (COVID-19): list of disinfectants with evidence for use against COVID-19. Accessed May 23, 2020 and December 30, 2020. https://www.canada.ca/en/health-canada/services/drugs-health-products/disinfectants/covid-19/list.html
- 4) Cimolai N. Environmental and decontamination issues for human coronaviruses and their potential surrogates. J Med Virol 2020; 92: 2498-2510.
- Lomont A, Boubaya M, Khamis W, Deslandes S, Cordel H, Seytre D, Alloui C, Malaure C, Bonnet N, Carbonnelle E, Cohen Y, Nunes H, Bouchaud O, Zahar J-R, Trandjaoui-Lambiotte Y. Environmental contamination related to SARS-CoV-2 in ICU patients. ERJ Open Res 2020; 6: 00595-2020.
- 6) Nelson A, Kassimatis J, Estoque J, Yang C, McKee G, Bryce E, Hoang L, Daly P, Lysyshyn M, Hayden AS, Harding J, Boraston S, Dawar M, Schwandt M. Environmental detection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) from medical equipment in long-term care facilities undergoing COVID-19 outbreaks. Am J Infect Control 2021; 49: 265-268.
- 7) Rahimi NR, Fouladi-Fard R, Aali R. Bidirectional association between COVID-19 and the environment: a systematic review. Environ Res 2020; 194: 110692.
- Hu X, Ni W, Wang Z, Ma G, Pan B, Dong L, Gao R, Jiang F. The distribution of SARS-CoV-2 contamination on the environmental surfaces during incubation period of COVID-19 patients. Ecotoxicol Environ Saf 2021; 208: 111438.
- Orenes-Piñero E, Baño F, Navas-Carillo D, Moreno-Docó A, Marin JM, Misiego R, Ramirez P. Evidence of SARS-CoV-2 virus air transmission indoors using several untouched surfaces: a pilot study. Sci Total Environ 2021; 751: 142317.
- Zulli A, Bakker A, Racharaks R, Nieto-Caballero M, Hernandez M, Shaughnessy R, Haverinen-Shaughnessy U, Ijaz K, Rubino J, Peccia J. Occurrence of respiratory viruses on school desks. Am J Infect Control 2020; S0196-6553(20)31042-7. Doi: 10.106/j.ajic.2020.12.006. Online ahead of print.
- 11) Ge XY, Pu Y, Liao CH, Huang WF, Zeng Q, Zhou H, Yi B, Wang AM, Dou QY, Zhou PC, Chen HL, Liu HX, Xu DM, Chen X, Huang X. Evaluation of the exposure risk of SARS-CoV-2 in different hospital environment. Sustain Cities Soc 2020; 61: 102413.
- 12) Yamagishi T, Ohnishi M, Matsunaga,N, Kakimoto K, Kamiya H, Okamoto K, Suzuki M, Gu Y, Sakaguchi M, Tajima T, Takaya S, Ohmagan N, Takeda M, Matsuyama S, Shirato K, Nao N, Hasegawa H, Kageyama T, Takayama I, Saito S, Wada K, Fujita R, Saito H, Okinaka K, Griffith M, Parry AE, Barnetson B, Leonard J, Wakita T. Environmental sampling for severe acute respiratory syndrome coronavirus 2 during COVID-19 outbreak in the Diamond Princess cruise ship. J Infect Dis 2020; 222: 1098-1102.
- 13) Abrahão JS, Sacchetto L, Rezende IM, Araújo Lima Rodrigues R, Correia Crispim AP, Moura C, Correia Mendonça D, Reis E, Souza F, Garcia Oliveira GF, Domingos I, de Miranda Boratto PV, Bastos Silva PH, Queiroz VF, Bastos Machado T, Flores Andrade LA, Lima Lourenço K, Silva T, Pereira Oliveira G, de Souza Alves V, Alves PA, Geessien Kroon E, de Souza Trindade G, Paiva Drumond B. Detection of SARS-CoV-2 RNA on public surfaces in a densely populated urban area of Brazil: a potential tool for monitoring the circulation of infected patients. Sci Total Environ 2021; 766: 142645.
- 14) Jang H, Ross TM. Dried SARS-CoV-2 virus maintains infectivity to Vero E6 cells for up to 48 h. Vet Microbiol 2020; 251: 108907.
- 15) Hirose R, Ikegaya H, Naito Y, Watanabe N, Yoshida T, Bandou R, Daidoji T, Itoh Y, Nakaya T. Survival of SARS-CoV-2 and influenza virus on the human skin: importance of hand hygiene in COVID-19. Clin Infect Dis 2020; ciaa1517. Doi: 10.1093/cid/ciaa1517. Online ahead of print.
- 16) Ibfelt T, Engelund EH, Schultz AC, Andersen LP. Effect of cleaning and disinfection of toys on infectious diseases and micro-organisms in daycare nurseries. J Hosp Infect 2015; 89: 109-115.
- 17) Bonny TS, Yezli S, Lednicky JA. Isolation and identification of human coronavirus 229E from frequently touched environmental surfaces of a university classroom that is cleaned daily. Am J Infect Control 2018; 46: 105-107.
- 18) Malenovská H. Coronavirus persistence on a plastic carrier under refrigeration conditions and its reduction using wet wiping technique, with respect to food safety. Food Environ Virol 2020; 12: 361-366.
- 19) Ben-Shmuel A, Brosh-Nissimov T, Glinert I, Bar-David E, Sittner A, Poni R, Cohen R, Achdout H, Tamir H, Yahalom-Ronen Y, Politi B, Melamed S, Vitner E, Cherry L, Israeli O, Beth-Din A, Paran N, Israely T, Yitzhaki S, Levy H, Weiss S. Detection and infectivity potential of severe acute respiratory syndrome coronavirus

2 (SARS-CoV-2) environmental contamination in isolation units and quarantine facilities. Clin Microbiol Infect 2020; 26: 1658-1662.

- 20) Chin AWH, Chu JTS, Perera MRA, Hui KPY, Yen HL, Chan MCW, Peiris M, Poon LLM. Stability of SARS-CoV-2 in different environmental conditions. Lancet Microbe 2020; 1: e10.
- Ge T, Lu Y, Zheng S, Zhuo L, Yu L, Ni Z, Zhou Y, Ni L, Qu T, Zhong Z. Evaluation of disinfection procedures in a designated hospital for COVID-19. Am J Infect Control 2020; S0196-6553(20)30812-9.
- 22) Ijaz MK, Nims RW, Whitehead K, McKinney J, Rubino JR, Ripley M, Jones C, Nims RW, Charlesworth B. Microbicidal actives with virucidal efficacy against SARS-CoV-2. Am J Infect Control 2020; 48: 972-973.
- 23) Kim UJ, Lee SY, Lee JY, Lee A, Kim SE, Choi OJ, Lee JS, Kee SJ, Jang HC. Air and environmental contamination caused by COVID-19 patients: a multi-center study. J Korean Med Sci 2020; 35: e332.
- 24) Lee S-E, Lee D-Y, Lee W-G, Kang BH, Jang YS, Ryu B, Lee SJ, Bahk H, Lee E. Detection of novel coronavirus on the surface of environmental materials contaminated by COVID-19 patients in the Republic of Korea. Osong Public Health Res Perspect 2020; 11: 128-132.
- 25) Lei H, Ye F, Liu X, Huang Z, Ling S, Jiang Z, Cheng J, Huang X, Wu Q, Wu S, Xie Y, Xiao C, Ye D, Yang Z, Li Y, Leung NHL, Cowling BJ, He J, Wong S-S, Zanin M. SARS-CoV-2 environmental contamination associated with persistently infected COVID-19 patients. Influenza Other Resp Viruses 2020; 14: 688-699.
- 26) Peyrony O, Ellouze S, Fontaine, JP, Thegat-Le Cam M, Salmona M, Feghoui L, Mahjoub N, Mercier-Delarue S, Gabassi A, Delaugerre C, Le Goff J; Saint-Louis CORE group. Surfaces and equipment contamination by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in the emergency department at a university hospital. Int J Hyg Environ Health 2020; 230: 113600.
- 27) Redmond SN, Dousa KM, Jones LD, Li DF, Cadnum JL, Navas ME, Kachaluba NM, Silva SY, Zabarsky TF, Eckstein EC, Procop GW, Donskey CJ. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) nucleic acid contamination of surfaces on a coronavirus disease 2019 ward and intensive care unit. Infect Control Hosp Epidemiol 2021; 40: 215-217.
- 28) Song ZG, Chen YM, Wu F, Xu L, Wang BF, Chen X, Dai FH, She JL, Holmes EC, Zhu TY, Zhang YZ. Identifying the risk of SARS-CoV-2 infection and environmental monitoring in airborne infectious isolation rooms (AIIRS). Virol Sin 2020; 35: 785-792.
- 29) Wong SC, Leung M, Tong DW, Lee LL, Leung WL, Chan FW, Chen JH, Hung IF, Yuen KY, Yeung DT, Chung KL, Cheng VC. Infection control challenges in setting up community isolation and treatment facilities for patients with coronavirus disease 2019 (COVID-19): implementation of directly observed environmental disinfection. Infect Control Hosp Epidemiol 2020; 1-29. Doi: 10.1017/ice.2020.1355. Online ahead of print.
- 30) Jin T, Li J, Yang J, Li J, Hong F, Long H, Deng Q, Qin Y, Jiang J, Zhou X, Song Q, Pan C, Luo P. SARS-CoV-2 presented in the air of an intensive care unit (ICU). Sustain Cities Soc 2021; 65: 102446.

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