# Post-liver transplant intrahepatic cholestasis: etiology, clinical presentation, therapy

F.R. PONZIANI<sup>1</sup>, S. BHOORI<sup>2</sup>, M. POMPILI<sup>1</sup>, M.A. ZOCCO<sup>1</sup>, M. BIOLATO<sup>1</sup>, G. MARRONE<sup>1</sup>, A. GASBARRINI<sup>1</sup>, V. MAZZAFERRO<sup>2</sup>, A. GRIECO<sup>1</sup>

<sup>1</sup>Internal Medicine, Gastroenterology, Hepatology, Agostino Gemelli Hospital, Rome, Italy <sup>2</sup>Liver Transplant, Hepatobiliary and Gastrointestinal Surgery, Istituto Nazionale Tumori, Milan, Italy

**Abstract.** - Post-liver transplant intrahepatic cholestasis is consequent to the impairment of bile flow or formation. It may develop in the early (within 6 months) or in the late (more than 6 months) post-liver transplant period and different causes may be recognized according to the time elapsed from a liver transplant. The raise at various degrees of serum bilirubin, alkaline phosphatase, and gamma-glutamyl transpeptidase, with or without increased transaminases levels, are common hematochemical findings. Liver histology is helpful for diagnostic assessment, and sometimes crucial to differentiate among possible causes of cholestasis. Although timely treatment of underling conditions as well as supportive care may resolve post-liver transplant intrahepatic cholestasis, the risk of graft loss and retransplantation are remarkable. For this reason, post-liver transplant intrahepatic cholestasis should be managed in collaboration with the LT center, and treatment should be devolved to expert hepatologists.

Key Words:

Cholestasis, Liver transplant, Ischemia reperfusion, Rejection, Small for size, Immunosuppression, Drug induced liver injury, DILI.

# Introduction

Cholestasis is a condition characterized by defective bile flow or formation<sup>1</sup>. It may result from an altered uptake, transfer and secretion of bile components, mainly caused by liver injury in the absence of mechanical obstruction.

The histological features of cholestasis are bile stasis in liver parenchyma and bile ducts plugs; bilirubin accumulation into hepatocytes, Kupffer cells, and canaliculi in zone 3 as well as ductular proliferation in zone 1, or bile acid retention ("cholate stasis") may be present. Blood examinations show the raise at various degrees of se-

rum bilirubin, alkaline phosphatase (ALP), and gamma-glutamyl transpeptidase (GGT), with or without increased transaminases levels.

In the liver transplant (LT) setting, cholestasis may be classified as extrahepatic, due to mechanical impairment of bile flow (e.g. anastomotic strictures, bile stones), or as intrahepatic, associated with impairment of liver cells or ductular dysfunction. Post LT cholestasis may develop early (within 6 months after LT) or late (more than 6 months after LT), with different etiology according to the time elapsed from LT (Table I).

In this review, the main causes of post LT intrahepatic cholestasis are discussed, focusing on clinical presentation and therapeutic approach.

# Early Post-Liver Transplant Intrahepatic Cholestasis

In the early post LT period intrahepatic cholestasis is common, usually subclinical and self-limiting. However, some patients may develop prolonged cholestasis and irreversible liver injury, leading to re-LT.

Initial poor graft function (IPGF) and primary graft non-function (PNF) may manifest with early post LT cholestasis. PNF is an irreversible graft dysfunction requiring emergency liver replacement within the first 10 days after LT<sup>2</sup>. It is characterized by rapidly rising transaminases, absence of bile production, severe coagulation deficit, hypoglycemia, high lactate levels, and hemodynamic instability. According to the united network of organ sharing (UNOS) criteria, PNF is defined by the presence of aspartate transaminase (AST) ≥ 5000 IU/L, international normalized ratio (INR)  $\geq$  3.0, and acidosis (pH  $\leq$  7.3 and/or lactate concentration  $\geq$  2× normal). Conversely, there is no agreement on the diagnostic criteria of IPGF, which is mainly characterized by elevation of serum transaminases<sup>3-5</sup>.

The incidence rates are variable, up to 11.8% for PNF and to 36% for IPGF, respectively; PNF is responsible for 81% of re-transplantations during the first week after surgery<sup>6</sup>.

The most frequent causes of early post LT cholestasis, which sometimes lead to PNF and IPGF, are mainly related to liver graft preservation and size match, infectious complications, acute rejection, and drugs.

# Ischemia/Reperfusion Injury

Ischemia reperfusion injury (IRI) is the consequence of the inflammatory response triggered by the procedures of organ procurement and preservation. Thus, IRI is a kind of "sterile inflammation", which develops in the absence of any microorganism<sup>7</sup>.

Two major types of IRI can be recognized. "Warm" IRI develops *in situ* during LT surgery when hepatic blood supply is temporarily interrupted, and is consequent to hepatocellular damage. "Cold" IRI, occurs *ex vivo* during liver graft preservation, is caused by hepatic sinusoidal endothelial cells damage and is coupled with warm IRI<sup>8,9</sup>. Other causes of IRI may be sepsis, shock, and trauma, which are pathological conditions eliciting systemic and liver hypoperfusion that may occur in the setting of LT.

In both warm and cold IRI, two stages of liver damage can be recognized. The first one is the ischemic injury phase, which initiates the process of cell death and is characterized by glycogen consumption, lack of oxygen, and ATP depletion. The subsequent reperfusion injury phase is the consequence of the metabolic derangement associated with inflammatory damage. Reperfusion injury phase can be further distinguished in early (or acute), occurring within the first 3 to 6 hours post-reperfusion and characterized by the activation of Kupffer cells, and late (or subacute), with massive neutrophil infiltration beginning at 18-24 hours post-reperfusion<sup>9-12</sup>.

IRI is caused by the innate immune response mediated by pattern recognition receptors (PRR)<sup>13-16</sup>, including Toll-like receptor (TLR) 4, TLR9 and the inflammasome<sup>17</sup>. Different cells of the immune system (T lymphocytes and natural killer lymphocytes, polymorphonuclear cells) are involved in the promotion of IRI, and recent data have reported a stimulation of adaptive immune response. The "no reflow" phenomenon observed after liver reperfusion is a common feature of IRI resulting from inflammation; in particular, blood flow mechanical obstruction is caused by adhe-

sion of inflammatory cells to the endothelium, increased interstitial fluid and endothelial vaso-constriction<sup>18</sup>. The activation of the immune system leads to a massive production of cytokines, chemokines, adhesion molecules, reactive oxygen species (ROS), to the activation of the complement system and to the promotion of autoimmune injury, as well as of mitochondrial dysfunction triggering cell death programs<sup>17</sup>. This sustains the pro-inflammatory process and produces liver damage.

#### **Treatment**

Liver susceptibility to IRI depends on organ preservation techniques and is increased by donor starvation, age, and graft steatosis<sup>19</sup>. Several measures can be adopted to reduce the risk of IRI in predisposed organs<sup>20</sup>. Surgical interventions, pharmacological agents, and gene therapy are the main treatment strategies, which have been extensively reviewed elsewhere<sup>19</sup>. On the whole, the use of a vasoprotectors, modulators of the renin-angiotensin system, β-blockers, antioxidants, growth factors, tyrosine kinase inhibitors, angiotensin II blockers, hydroxy-methylglutaryl (HMG)-Coenzyme A (CoA) reductase inhibitors, calcium channel blockers, peroxisome proliferator-activated receptors (PPAR-α) agonists are the most innovative approaches reported in literature, to be distinguished form surgical measures widely used in everyday practice (e.g. minimization of cold and warm ischemia time, intermittent clamping, ischemic preconditioning, the use of preservation solutions with specific additives).

# Small-For-Size Syndrome

Living donor LT (LDLT) is a common practice for Asian transplant Centers. It has been implemented in selected cases to increase the number of available organs in Western Countries and to reduce the waiting list period. In practice, donor's liver is splitted in two parts, one of them being used as transplantable graft. This procedure may be adopted for deceased donors too. To obtain an adequate equilibrium between the amount of transplanted (recipient) and residual (donor) liver tissue, in adult-to-adult LDLT graft size should cover 30-40% of the expected recipient's liver volume or 0.8-1.0% of the recipient's body weight<sup>21</sup>. Therefore, the right liver lobe is usually preferred. Living donor liver volumes are calculated based on three-dimensional CT scan; the most useful estimated parameters are graft volume to standard liver volume (SLV) ratio, and graft weight-to-recipient body weight ratio (GWBWR)<sup>22-25</sup>. A GWBWR of 0.8% is a widely accepted cut-off for minimizing the risk of graft failure<sup>26</sup>. In the case of deceased donors, the body surface area index (BSAi) is adopted<sup>27</sup>.

Small-for-size syndrome (SFSS) occurs when the partial liver graft fails to fulfill the functional demand of the recipient<sup>28</sup>. If the hepatocyte cellular mass is inadequate for the recipient size, the small vascular network of the graft causes a discrepancy between accelerated liver regeneration and inadequate supply of oxygen and growth factors. This leads to hyperafflux in the portal system (portal hyperperfusion), reduction in arterial perfusion (hepatic arterial buffer response) and sinusoidal microcirculatory disturbances causing liver damage and inflammation<sup>29-32</sup>.

SFSS encompasses various clinical presentations, ranging from mild hepatic dysfunction and isolated hyperbilirubinemia, to coagulopathy, ascites, portal hypertension, prolonged cholestasis, encephalopathy, and irreversible graft failure leading to death of the patient in the absence of an available organ for re-transplantation.

Liver biopsy shows diffuse ischemic damage, cellular ballooning, and features of cholestasis<sup>33</sup>.

#### **Treatment**

Treatment is mainly aimed at reducing portal hyperperfusion by mechanic procedures (splenic artery ligation, splenectomy, portosystemic shunts, extracorporeal continuous portal diversion [ECPD]), or medical therapy (splanchnic vasoconstrictors such as: terlipressin, somatostatin, octreotide)<sup>34</sup>.

#### Infections

Infectious complications are a common cause of morbidity and mortality among LT recipients. Bacterial infections are the most frequent (80%), followed by viral (20%) and fungal (8%) ones<sup>35</sup>-<sup>37</sup>. Both the reactivation of previous infections or the exposition to new infectious agents is possible. Concomitant factors may contribute to the onset and the evolution of the infective episode; they can be recipient-related (advanced age, MELD score >30 at LT, acute liver failure, malnutrition, > 48 hours stay in intensive care unit [ICU], prolonged hospital stay, previous infections, diabetes), donor-related (prolonged ICU stay, previous infections, marginal graft), surgery-related (choledochojejunostomy, prolonged surgery [> 12 h], re-operation or re-transplantation, transfusion of more than 15 blood units),

and related to other post-LT factors (mechanical ventilation, level and type of immunosuppression [e.g. monoclonal and polyclonal antibodies], PNF, vascular complications [e.g. hepatic artery thrombosis and portal vein thrombosis], biliary complications [e.g. ischaemic cholangitis, biliary strictures and fistulae])<sup>36-43</sup>.

The type of infection is associated with the time from LT. In the first month after surgery (early period) opportunistic, donor-derived and surgical site infections are usual, whereas at 2-12 months after LT (intermediate period) opportunistic (Mycobacterium tuberculosis, Pneumocystis, Listeria, Cryptococcus, Toxoplasma, Rhodococcus, Nocardia, Legionella, etc.) and community acquired infections (flu, urinary tract infections) are more frequent. After the first year post LT (late period) community acquired infections are more likely to occur, whereas opportunistic infections are generally rare<sup>44</sup>.

Cytomegalovirus (CMV) infection is one of the most common viral complications of the early-intermediate post LT period, involving about 30-50% of LT recipients although a delayed onset can also be observed<sup>44</sup>. Either new or reactivated infection may occur. It is characterized by virus replication in blood, which can be defined as "CMV disease" in the presence of the following associated manifestations: fever >38°C, for at least 2 days within a 4-day period, neutropenia or thrombocytopenia, and tissue invasion with organ dysfunction, including lung, liver, kidney and central nervous system<sup>45</sup>. According to donor and recipient previous contact with CMV, the infection can be defined as primary (donor positive/recipient negative) or as superinfection (donor positive/recipient negative). In the case of primary infection, in the absence of prophylaxis, over 90% of the recipients develop CMV infection compared to about 25% of the recipients in the case of superinfection; in the case of reactivation of previous infection (donor positive/ recipient negative), around 15% of the recipients become ill.

The liver is a regulatory organ in the host defense system, acting as a firewall against systemic diffusion of bacteria and pathogens; during infections, several proinflammatory cytokines (tumor necrosis factor alpha [TNF- $\alpha$ ], interleukin [IL]-1, IL-6, and IL-8) are released, altering bile acid transport at the sinusoidal and canalicular membrane domains<sup>46-49</sup>.

Cholestasis may precede the development of septic complications; bacterial infections usually present with high fever, cholestasis, and positive blood cultures. Liver biopsy specimens show biliary tract inflammation with neutrophil infiltrates, bile duct proliferation, and bile plugs<sup>50</sup>.

CMV hepatitis usually presents with fever, jaundice and increased cholestasis blood test. Liver histology shows cholestatic features, microabscesses and typical intranuclear "owl's eye" inclusions<sup>51</sup>. Persistence of CMV has been demonstrated in liver grafts developing vanishing bile duct syndrome and chronic rejection<sup>52</sup>. Although CMV infection may increase alloantigens expression, making bile ducts more vulnerable to immunologic damage<sup>53</sup>, large studies have failed to demonstrate any significant association between CMV infection and the development of graft cholangiopathy<sup>54</sup>.

#### **Treatment**

The treatment of the infectious complication usually resolves cholestasis.

## Acute Cellular Rejection

Although the prevalence of acute cellular rejection (ACR) is declining, the current incidence of clinically significant acute rejection is 10-40%<sup>55</sup>.

ACR may be early, usually occurring within 3 months post LT, or late, occurring after 3 months post LT<sup>56</sup>. In any case, it is triggered by a T cells-driven immune response against major histocompatibility complex (MHC) alloantigens in the liver graft. CD4+ and CD8+ T cells but also myeloid cells and innate lymphoid cells are initially involved, but hepatic inflammation leads to further recruitment of leukocytes from circulation<sup>55,57</sup>. Inflammation results in hepatocytes, endothelial cells, and bile ducts damage<sup>58-62</sup>.

The association between antibody-mediated rejection and ACR has also been a controversial issue in the LT setting. Antibody-mediated rejection is a hyperacute rejection characterized by graft endothelial damage by complement activation, with direct injury to the capillary endothelium and activation of inflammatory cells, such as natural killer (NK) lymphocytes, macrophages, and neutrophils<sup>63</sup>.

ACR should be distinguished from hyperactue rejection. Donor-specific human alloantibodies (DSAs) against leukocyte antigen (HLA) and ABO antibodies are involved in hyperacute rejection. DSAs are a well-known risk factor for decreased graft survival after kidney and heart transplantation. A positive "cross-match", intended as the detection of antibodies in the recipient's serum binding to the surface of donor's lymphocytes, is associated with an increased risk of hyperacute rejection of the kidney allograft<sup>64</sup>.

The liver is relatively resistant to antibodies mediated injury due to its wide sinusoidal endothelial surface allowing the absorption of circulating antibodies, to its intrinsic ability to regenerate and also for the secretion of soluble HLA I that binds and inactivate circulating antibodies<sup>65-68</sup>. These mechanisms of protection allow the clearance of preexisting DSAs in 90% of presensitized LT-recipients<sup>69</sup>. However, in patients with rejection or inflammatory conditions involving the liver the exposure of alloantigen secondary to tissue injury may trigger the production of "de novo" DSAs, similary to what reported for kidney transplant recipients<sup>70,71</sup>.

The association between a positive cross-matching and ACR in LT recipients is not constant among published studies and is, therefore, difficult to draw firm conclusions<sup>69,72-77</sup>. Although the presence of inflammation in biopsy specimens from patients with a positive cross-match has been reported<sup>78</sup>, no significant difference in transaminases and bilirubin levels, as well as in graft and recipient's survival, was proven compared to cross-match negative patients<sup>78-81</sup>.

Histologically, ACR is classified according to Banff criteria<sup>82,83</sup>. Portal, bile duct and venous endothelial inflammation are the three main features of ACR, and at least two of them are required for the diagnosis. Inflammatory cells in the portal spaces are represented by a mixed population of lymphocytes (T cells), blast cells, macrophages, neutrophils, and eosinophils. Bile ducts appear inflamed, with degenerative changes or focal luminal disruption. Centrilobular perivenulitis consists in hepatic venous and perivenular inflammation with perivenular hepatocyte loss. In late ACR this "typical" picture is less prominent, but central perivenulitis is more marked<sup>56,84,85</sup>.

ACR is as always asymptomatic, but jaundice can be present. The diagnosis is supposed based on the increase in transaminases, bilirubin and cholestasis enzymes levels and is confirmed by liver biopsy findings.

#### **Treatment**

Advances in immunosuppressive therapy have significantly increased the success of LT, minimizing the risk of ACR. Histological evidence of ACR without any biochemical alteration does not

require treatment; on the other hand, when ACR is suspected liver biopsy is mandatory to grade its severity and to exclude other causes of liver damage (e.g. drugs, IRI, infections, recurrent disease) before treatment. The increase of immunosuppression alone is usually effective to treat mild ACR; the maintenance of increased levels of immunosuppressive drugs associated with corticosteroid boluses is recommended in moderate-severe ACR<sup>86</sup>.

# Drug Induced Liver Injury

Despite data on the general population are scarce, the prevalence of hepatotoxicity among patients hospitalized for jaundice has been estimated to range between 2% and 10%<sup>87-91</sup> and an incidence as high as 2% has been reported in LT recipients<sup>92</sup>.

Drug induced liver injury (DILI) is an uncommon cause of cholestasis, even if large studies confirm that among patients with DILI 20-40% may have a cholestatic histological pattern and 12-20% a mixed hepatocellular/cholestatic pattern<sup>93-96</sup>

Several drugs necessary for LT recipients may have hepatotoxic effects and may lead to cholestasis. Therefore, drug induced liver injury (DILI) may be a cause of cholestatic damage in both the early and the late post LT period.

The most common clinical presentation of cholestatic DILI is the increase in ALP with or without jaundice and pruritus. Fever and abdominal pain may be present. Diagnosis of DILI is often difficult after LT, especially in the early period, due to the coexistence of other causes of liver damage.

Histology is helpful in the differential diagnostic process and in the classification of cholestatic DILI, which can be distinguished as follows<sup>93-95</sup>:

Acute pure cholestasis, presenting with hepatocyte cholestasis, canalicular dilation, and bile plugs in the absence of relevant inflammation;

- Acute cholestatic hepatitis, characterized by the association of cholestasis, inflammation and sometimes hepatocellular necrosis;
- Cholestasis with bile duct injury, when ductular, cholangiolar, or cholangiolytic damage is prevalent and hepatocellular injury is minimal;
- Vanishing bile duct syndrome.

Cholestatic DILI pattern is defined as an increase in ALP >2 × the upper limit of normal and/or with an alanine aminotransferase (ALT)/ ALP ratio <2, whereas a mixed (citonecrotic and cholestatic) DILI pattern as an ALT/alkaline phosphatase ratio greater than 2 and less than 5. However, these features are common in the early post LT period<sup>97,98</sup>. The Roussel Uclaf Assessment model (RUCAM) may be useful to investigate the likelihood of DILI97,98. Furthermore, some drugs have a "signature pattern," as a typical pattern of liver injury may be recognized after a similar duration of drug intake. In the diagnostic workup, online archives and web sites are useful tools to investigate drugs hepatoxicity99.

It is usually difficult to recognize DILI in the LT setting due to the multiple factors potentially responsible for liver damage. However, in presence of unexplained cholestasis, after the exclusion of common causes of liver injury and after extensive testing including liver biopsy, DILI should be considered as a possible cause.

In the same way as in the general population, histological assessment of DILI is challenging in LT recipients and may be confounding, due to the influence of several other factors, sometimes delaying DILI diagnosis. Moreover, the investigation of medication history, which is mandatory to estimate the time to DILI onset, may be difficult due to variations in treatment doses and duration as well as interruptions/re-start. Conversely, the strict biochemical monitoring to which LT recipients are subjected is a potential advantage in the early detection of DILI.

Table I. Main causes of post LT intrahepatic cholestasis according to post LT period

Early post LT period (<6 months)	Late post LT period (≥6 months)
Primary non function Ischemia/reperfusion injury Small for size graft Infections Acute rejecton Antibodies-mediated rejection Drug induced liver injury	Chronic rejection Infections Drug induced liver injury Recurrence of the original disease

**Table II.** Classification of medications associated with drug induced liver injury according to the number of reports and to the likelihood of use in liver transplant recipients 98,123.

Drugs category A (≥ 50 Cases)	Drugs category B (18-49 Cases)	Drugs category C (4-11 Cases)	Drugs category D (1-3 Cases)
Frequently used in LT:	Frequently used in LT:	Frequently used in LT:	Frequently used in LT:
Illopurinol*	Celecoxib*	Amlodipine*	Acyclovir
torvastatin*	Clopidogrel*	Candesartan*	Atenolol*
Diclofenac	Enalapril*	Gemfibrozil*	Carvedilol*
buprofen*	Esomeprazole	Gilipizide*	Clofibrate*
iclopidine*	Fenofibrate*	Glimepiride*	Enoxaparin
	Fluvastatin*	Irbesartan*	Fondaparinux
Incommonly/occasionally	Glibenclamide*	Ketoprofen*	Hydrochlorotiazide*
sed in LT:	Lisinopril	Losartan*	Glicazide*
miodarone*	Lovastatin*	Lansoprazole*	Propofol
moxicillin+calvulanate*	Metformin*	Metronidazole	Rabeprazole*
	Naproxen*	Pravastatin*	Raltegravir
arely used in LT:	Nifedipine	Pantoprazole	Repaglinide
usulfan*	Omeprazole	Ramipril*	Spironolactone*
arbamazepin*	Ranitidine*	Kumpru	Tamsulosin
antrolene	Rosuvastatin*	Uncommonly/occasionally	Valsartan*
favirenz*	2.00 m r mo mm m m	used in LT:	raisui iuii
rythromycin*	Uncommonly/occasionally	Ampicillin*	Uncommonly/Occasionally
loxuridine*	used in LT:	Alfuzosin*	used in LT:
alothane	Amoxicillin*	Diltiazem*	Cefaclor
ıfliximab*	Azithromycin*	Levocetirizine*	Cefadroxi
etoconazole*	Captopril*	Vancomycin	Cefnidir
evirapine*	Captoprii Cefazolin	Verapamil*	Cefepime
imesulide*	Ceftriaxone*		
henytoin*	Ciprofloxacin*	Warfarin*	Cefoperazone
nenytotn Juinidine*	Clarithromycin*	Rarely used in LT:	Cefotaxime
	Fluconazole		Cefprozil
ifampin* imvastatin*		Amphotericin B*	Ceftazidime
	Levofloxacin*	Albendazole* Bosentan*	Cefuroxime
ulfasalazine*	Propafenone*		Flecainide
ulfonamides*	Danala and in LT.	Cephalexin	Fosfomycin*
ulindac*	Rarely used in LT:	Citalopram*	Imipenem*
hioguanine*	Clindamycin	Cytarabine*	Meropenem*
alproic Acid*	Clozapine*	Daptomycin	Sitagliptin
	Doxorubicin	Doxycycline*	Valacyclovir*
	Duloxetine*	Escitalopram*	n 1 1: rm
	Etanercept*	Fluoxetine*	Rarely used in LT:
	Heparin	Erlotinib	Acetazolamide*
	Imatinib*	Famotidine	Aliskiren*
	Irinotecan	Flavocoxid*	Alosetron
	Itraconazole*	Fluorouracil*	Alprazolam*
	Moxifloxacin*	Gabapentin*	Anastrozole
	Olanzapine*	Gemcitabine*	Atazanavir*
	Ofloxacin*	Hydroxyurea	Bortezomib*
	Paroxetine*	Indomethacin*	Carbenicillin*
	Phenobarbital*	Isotretinoin	Carboplatin
	Quinine*	Labetalol	Chlorambucil*
	Rivaroxaban	Levetiracetam	Cisplatin
	Sertraline*	Linezolid*	Clomipramine*
	Tamoxifen*	Mesalamines*	Clonazepam*
	Thiabendazole*	Mirtazapine*	Dabigatran
	Venlafaxine*	Mitomycin	Dalteparin
	Voriconazole*	Montelukast*	Darbepoetin alfa
		Nafcillin	Dasatinib
		Natalizumab*	Deferoxamine
		Nefazodone*	Donepezil
		Norfloxacin*	Entacapone*
		Orlistat	Ethambutol*
		Penicillin G*	
		Pioglitazone*	
		Pregabalin*	
		Procainamide*	

Table continued

Table II. Continued. Classification of medications associated with drug induced liver injury according to the number of reports
and to the likelihood of use in liver transplant recipients <sup>98,123</sup> .

Drugs category A	Drugs category B	Drugs category C	Drugs category D
(≥ 50 Cases)	(18-49 Cases)	(4-11 Cases)	(1-3 Cases)
	Quetiapine* Risperidone* Ritonavir* Rosiglitazone* Sorafenib Thalidomide* Tolcapone* Topiramate Trazodone* Vincristine Zafirlukast	Hydroxychloroquine Lopinavir* Mebendazole Mefloquine* Metoprolol Micafungin Nelfinavir Ondasetron Oxcarbazepine* Rifabutin* Rilpivirine Rivastigmine Saquinavir* Sildenafil* Terbutaline Thyroxine Tobramycin Tolvaptan* Triamterene* Zileuton	

<sup>\*</sup>mixed or cholestatic pattern of drug induced liver injury.

The optimization of tailored immunosuppressive drugs regimens have significantly minimized the incidence of DILI in LT recipients, but the risk is not completely absent and increases when complications or other concomitant medical conditions require treatment.

Cyclosporine may have cholestatic effects interfering with bile formation and leading to hyperbilirubinemia and to the formation of biliary sludge<sup>100-108</sup>. The inhibition of ATP-dependent export carriers and of the bile salts export pumps in the canalicular membrane of hepatocytes, together with impairment of biliary secretion of glutathione, are involved in cyclosporine-related impairment of canalicular bile flow 105-107. Similarly to cyclosporine but at a lesser extent, tacrolimus has been reported to inhibit canalicular bile acids transport and glutathione secretion<sup>105,109,110</sup>. However, Ericzon et al. reported that tacrolimus was able to recover bile acid secretion after LT more rapidly than cyclosporine<sup>111,112</sup>. As regards other immunosuppressive agents, two cases of cholestatic liver damage have been reported in two renal transplant recipients receiving mycophenolate and sirolimus, respectively<sup>112,113</sup>.

Azathioprine, a purine analogue rarely used as immunsosuppressant in the LT setting today, may cause damage to the hepatic sinusoidal and venular endothelial cells, resulting in a variety of clinical, biochemical, and histologic manifestations, including cholestasis<sup>115-123</sup>.

Among other drugs that may cause a cholestatic syndrome in LT recipients, antifungal agents as well as antibiotics have been reported as the most common agents of DILI in LT recipients<sup>92</sup>.

Pharmacological therapies for cardiovascular diseases are frequently employed in LT recipients as well as analgesics and antinflammatory antidiabetic neurologic, anticoagulant, antiaggregant, gastroenterologic and psychotropic medications. The main drugs potentially causing DILI, including the cholestatic type, are reported in Table II, and are stratified according to the frequency of hepatic adverse effects<sup>124</sup>. Medical therapies should not be avoided only for concerns regarding their potential hepatotoxicity, but thorough patient evaluation is mandatory to avoid underrecognized cases of DILI. Particular attention should be paid to the use of herbal and dietary supplements too, which may be hepatotoxic by itself or due to the presence of other ingredients and adulterants<sup>125,126</sup>.

Only 6% of patients with DILI develop chronic liver injury<sup>127</sup>. However, patients with cholestatic DILI seem to be more prone to experience chronic liver disease; in a minor part of them, ductopenia and vanishing bile duct syndrome with the progression to biliary cirrhosis may also occur, especially in association with specific drugs<sup>128-134</sup>. Reversal of drug-related vanishing bile duct syndrome has also been reported<sup>135,136</sup>.

DILI-related mortality ranges between 5% and 14%<sup>93-95</sup>. However, in a recent study focused only on LT recipients, DILI did not affect the post LT outcome as patients did not require re-transplantation and no death was reported<sup>92</sup>.

#### **Treatment**

Immediate withdrawal of the suspected drug is the core principle of DILI management. In the case of immunosuppressive drugs, dose reduction or conversion to other regimens may be considered. Specific therapies may be adopted for few toxic agents, such as N-acetylcysteine for acetaminophen overdose<sup>137</sup>. N-acetylcysteine has been empirically administered to adults with DILI due to other agents too, reporting favorable results. Corticosteroids may be used in case of tyrosine kinase inhibitors-related DILI or in case of drug-induced autoimmune-like hepatitis, especially when liver injury does not resolve after withdrawal of the offending agent. Ursodeoxycholic acid administration has obtained beneficial results in the setting of cholestatic DILI, as well as cholestyramin alone or in association with antihistamines as regards pruritus relief. Silibinin use in the setting of DILI has obtained contrasting results, whereas few data on methionine and glutathione are available.

Gastric decontamination (aspiration by lavage, charcoal) should be considered in selected cases of DILI, such as those related to acteminophen overdose, toxic mushrooms, salicylate or to the ingestion of other toxic substances.

# Late Post-Liver Transplant Intrahepatic Cholestasis

The main causes of intrahepatic cholestasis in the late post LT period are chronic rejection (CR), infections and DILI, which have been extensively discussed above.

Exclusion of the recurrence of the original liver disease (e.g. hepatitis C virus infection, primary sclerosing cholangitis, primary biliary cholangitis) and of extrahepatic conditions inducing cholestasis, such as hepatic artery thrombosis and mechanical obstruction, is mandatory to guide treatment.

## Chronic Rejection

The prevalence of CR (otherwise called "ductopenic rejection") is 2% in LT recipients, although probably underestimated for the lack of protocols

for routine histological assessment<sup>138,139</sup>. It tipically occurs during the first 12 months after LT but a delayed diagnosis is possible, since CR may have an indolent course over years<sup>140</sup>. CR may occur in patients with previous episodes of ACR, even if this is not a strict requirement<sup>141</sup>. As like as for ACR, a positive cross-matching has been demonstrated in patients with CR<sup>80,142-144</sup>. Notably, even if data from published studies are contrasting, circulating DSAs have been associated with an increased risk of developing rejection in patients who have weaned immunosuppression.

Loss of bile ducts (vanishing bile duct syndrome) at liver histology and obliteration of large and medium size arteries are the mainstay of CR. These pathological alterations are due to the infiltration of macrophages (foam cells) and inflammatory cells into the vessel wall, leading to fibrosis and, finally to the obliteration of arterial lumen with consequent ischemic damage.

Histologically, ductopenia should be present in more than 50% of portal tracts to make a reliable diagnosis of CR145-147. Bile ducts inflammation can be present in the early stages, subsiding during time, and bile ducts degeneration leads to dysplastic and athrophic features. In the centrilobular zone, bilirubinostasis, hepatocyte ballooning and loss may be present, evolving in centrilobular fibrosis and cirrhosis during time. Fibrosis pattern may be veno-centric, which is associated with the obliteration of hepatic and portal vein branches, periportal/biliary, when bile duct inflammation and loss is predominant, or centrilobular, which is associated with central perivenulitis. Bile duct proliferation and periportal fibrosis are more frequent in cases of late presentation, and are suggestive of a prolonged course of CR.

Microscopic vascular abnormalities can be present and the small portal tracts may show a reduced number of small arterial branches and of other microvascular channels<sup>145-147</sup>.

CR may have a long asymptomatic course; progressive jaundice, with or without ascites, is the typical clinical manifestation.

#### **Treatment**

Switch of immunosuppressive therapy or its potentiation (combination of two or 3 drugs) are the most commonly used approaches. Although chronic rejection is sometimes irreversible, retransplantation is necessary in only 5% of cases<sup>138,139</sup>.

#### Conclusions

Intrahepatic cholestasis in LT recipients is a challenge for clinicians, as multiple conditions may alter the clinical picture. In the early post LT period, cholestasis may be a manifestation of graft dyfunction, and a rapid assessment of the possible cause is mandatory to start the correct treatment rapidly and to avoid the loss of liver graft or its irreversible damage. The improvement in surgical techniques and organ preservation methods as well as the progressive shortening of both donor and recipient surgical procedures times, have significantly reduced the incidence of early post-LT cholestasis. Likewise, the optimization of immunosuppressive regimens has minimized the incidence of rejection, of early and late infections and of immunosuppressive drugs-related cholestasis.

In this scenario, DILI remains the most common cause of post LT cholestasis, especially in the late period. Thorough investigation of medical history is mandatory to exclude a possible DILI; while the identification of a drug responsible for cholestasis may be very difficult in the early postoperative period, it can be easier in the late phase, when pharmacological therapy is almost stable. However, most of the medications are necessary and treatment modifications should be done considering potential drug-to-drug interactions and under close clinical monitoring.

Liver biopsy maintains a crucial role in the differential diagnosis of cholestasis in the LT setting. However, as protocol biopsies in asymptomatic recipients with normal or near-normal liver tests after years from LT are not universally performed, liver histology is frequently obtained only when ematochemical parameters become abnormal.

For these reasons, the management of cholestasis in the post LT setting is a complex medical issue, and should be performed in collaboration with the LT center and devolved to expert hepatologists.

#### **Conflict of interest**

The authors declare no conflicts of interest.

## References

- SHERLOCK S, DOOLEY J. Diseases of the liver and biliary system. Blackwell Science, 2002.
- 2) UEMURA T, RANDALL HB, SANCHEZ EQ, IKEGAMI T, NARA-SIMHAN G, MCKENNA GJ, CHINNAKOTLA S, LEVY MF, GOLD-

- STEIN RM, KLINTMALM GB. Liver retransplantation for primary nonfunction: analysis of a 20-year single-center experience. Liver Transpl 2007; 13: 227-233.
- PLOEG RJ, D'ALESSANDRO AM, KNECHTLE SJ, STEGALL MD, PIRSCH JD, HOFFMANN RM, SASAKI T, SOLLINGER HW, BELZER FO, KALAYOGLU M. Risk factors for primary dysfunction after liver transplantation--a multivariate analysis. Transplantation 1993; 55: 807-813.
- GONZALEZ FX, RIMOLA A, GRANDE L, ANTOLIN M, GAR-CIA-VALDECASAS JC, FUSTER J, LACY AM, CUGAT E, VISA J, RODES J. Predictive factors of early postoperative graft function in human liver transplantation. Hepatology 1994; 20: 565-573.
- NANASHIMA A, PILLAY P, VERRAN DJ, PAINTER D, NAKASUJI M, CRAWFORD M, SHI L, Ross AG. Analysis of initial poor graft function after orthotopic liver transplantation: experience of an australian single liver transplantation center. Transplant Proc 2002; 34: 1231-1235.
- 6) CHEN HAO XJ, SHEN BAIYONG, DENG XIAXING, TAO RAN, PENG CHENGHONG, LI HONGWEI. Initial Poor Graft Dysfunction and Primary Graft Non-Function After Orthotopic Liver Transplantation, Liver Biopsy in Modern Medicine. In: D. Y. Mizuguchi, ed., 2011.
- CHEN GY, NUNEZ G. Sterile inflammation: sensing and reacting to damage. Nat Rev Immunol 2010; 10: 826-837.
- ZHAI Y, BUSUTTIL RW, KUPIEC-WEGLINSKI JW. Liver ischemia and reperfusion injury: new insights into mechanisms of innate-adaptive immune-mediated tissue inflammation. Am J Transplant 2011; 11: 1563-1569.
- IKEDA T, YANAGA K, KISHIKAWA K, KAKIZOE S, SHIMADA M, SUGIMACHI K. Ischemic injury in liver transplantation: difference in injury sites between warm and cold ischemia in rats. Hepatology 1992; 16: 454-461.
- JAESCHKE H, FARHOOD A. Neutrophil and Kupffer cell-induced oxidant stress and ischemia-reperfusion injury in rat liver. Am J Physiol 1991; 260: G355-362.
- FONDEVILA C, BUSUTTIL RW, KUPIEC-WEGLINSKI JW. Hepatic ischemia/reperfusion injury--a fresh look. Exp Mol Pathol 2003; 74: 86-93.
- 12) ARDIZZONE G, STRATTA C, VALZAN S, CRUCITTI M, GALLO M, CERUTTI E. Acute blood leukocyte reduction after liver reperfusion: a marker of ischemic injury. Transplant Proc 2006; 38: 1076-1077.
- 13) TAKEUCHI O, AKIRA S. Pattern recognition receptors and inflammation. Cell 2010; 140: 805-820.
- AKIRA S, TAKEDA K. Toll-like receptor signalling. Nat Rev Immunol 2004; 4: 499-511.
- TRINCHIERI G, SHER A. Cooperation of Toll-like receptor signals in innate immune defence. Nat Rev Immunol 2007; 7: 179-190.
- 16) BACCALA R, GONZALEZ-QUINTIAL R, LAWSON BR, STERN ME, KONO DH, BEUTLER B, THEOFILOPOULOS AN. Sensors of the innate immune system: their mode of action. Nat Rev Rheumatol 2009; 5: 448-556.

- 17) Zhai Y, Petrowsky H, Hong JC, Busuttil RW, Kupiec-Weglinski JW. Ischaemia-reperfusion injury in liver transplantation--from bench to bedside. Nat Rev Gastroenterol Hepatol 2013; 10: 79-89.
- 18) ELTZSCHIG HK, ECKLE T. Ischemia and reperfusion-from mechanism to translation. Nat Med 2011; 17: 1391-1401.
- Peralta C, Jimenez-Castro MB, Gracia-Sancho J. Hepatic ischemia and reperfusion injury: effects on the liver sinusoidal milieu. J Hepatol 2013; 59: 1094-1106.
- Li J, Li RJ, Lv GY, Liu HQ. The mechanisms and strategies to protect from hepatic ischemia-reperfusion injury. Eur Rev Med Pharmacol Sci 2015; 19: 2036-2047.
- KAWASAKI S, MAKUUCHI M, MATSUNAMI H, HASHIKURA Y, IKEGAMI T, NAKAZAWA Y, CHISUWA H, TERADA M, MIYA-GAWA S. Living related liver transplantation in adults. Ann Surg 1998; 227: 269-274.
- 22) URATA K, KAWASAKI S, MATSUNAMI H, HASHIKURA Y, IKEGAMI T, ISHIZONE S, MOMOSE Y, KOMIYAMA A, MAKUUCHI M. Calculation of child and adult standard liver volume for liver transplantation. Hepatology 1995; 21: 1317-1321.
- 23) KIUCHI T, KASAHARA M, URYUHARA K, INOMATA Y, UEMOTO S, ASONUMA K, EGAWA H, FUJITA S, HAYASHI M, TANAKA K. Impact of graft size mismatching on graft prognosis in liver transplantation from living donors. Transplantation 1999; 67: 321-327.
- 24) LEMKE AJ, BRINKMANN MJ, PASCHER A, STEINMULLER T, SETTMACHER U, NEUHAUS P, FELIX R. [Accuracy of the CT-estimated weight of the right hepatic lobe prior to living related liver donation (LRLD) for predicting the intraoperatively measured weight of the graft]. Rofo 2003; 175:1232-8.
- 25) Yoshizumi T, Gondolesi GE, Bodian CA, Jeon H, Schwartz ME, Fishbein TM, Miller CM, Emre S. A simple new formula to assess liver weight. Transplant Proc 2003; 35: 1415-1420.
- 26) Breitenstein S, Apestegui C, Petrowsky H, Clavien PA. "State of the art" in liver resection and living donor liver transplantation: a worldwide survey of 100 liver centers. World J Surg 2009; 33: 797-803.
- 27) FUKAZAWA K, NISHIDA S, VOLSKY A, TZAKIS AG, PRETTO EA, JR. Body surface area index predicts outcome in orthotopic liver transplantation. J Hepatobiliary Pancreat Sci 2011; 18: 216-225.
- 28) DAHM F, GEORGIEV P, CLAVIEN PA. Small-for-size syndrome after partial liver transplantation: definition, mechanisms of disease and clinical implications. Am J Transplant 2005; 5: 2605-2610.
- FUKAZAWA K, NISHIDA S. Size mismatch in liver transplantation. J Hepatobiliary Pancreat Sci 2016; 23: 457-466.
- YANG ZF, Ho DW, CHU AC, WANG YO, FAN ST. Linking inflammation to acute rejection in small-forsize liver allografts: the potential role of early macrophage activation. Am J Transplant 2004; 4: 196-209.
- 31) YANG ZF, POON RT, LUO Y, CHEUNG CK, HO DW, LO CM, FAN ST. Up-regulation of vascular endothelial

- growth factor (VEGF) in small-for-size liver grafts enhances macrophage activities through VEGF receptor 2-dependent pathway. J Immunol 2004; 173: 2507-2515.
- 32) Oura T, Taniguchi M, Shimamura T, Suzuki T, Yamashita K, Uno M, Goto R, Watanabe M, Kamiyama T, Matsushita M, Furukawa H, Todo S. Does the permanent portacaval shunt for a small-for-size graft in a living donor liver transplantation do more harm than good? Am J Transplant 2008; 8: 250-252.
- 33) DEMETRIS AJ, KELLY DM, EGHTESAD B, FONTES P, WALLIS MARSH J, TOM K, TAN HP, SHAW-STIFFEL T, BOIG L, NO-VELLI P, PLANINSIC R, FUNG JJ, MARCOS A. Pathophysiologic observations and histopathologic recognition of the portal hyperperfusion or small-for-size syndrome. Am J Surg Pathol 2006; 30: 986-993.
- 34) Taniguchi M, Shimamura T, Todo S, Furukawa H. Small-for-size syndrome in living-donor liver transplantation using a left lobe graft. Surg Today 2015: 45: 663-671.
- 35) VERA A, CONTRERAS F, GUEVARA F. Incidence and risk factors for infections after liver transplant: single-center experience at the University Hospital Fundacion Santa Fe de Bogota, Colombia. Transpl Infect Dis 2011; 13: 608-615.
- 36) ROMERO FA, RAZONABLE RR. Infections in liver transplant recipients. World J Hepatol 2011; 3: 83-92.
- FISHMAN JA. Infection in solid-organ transplant recipients. N Engl J Med 2007; 357: 2601-2614.
- 38) Huprikar S. Update in infectious diseases in liver transplant recipients. Clin Liver Dis 2007; 11: 337-354.
- SNYDMAN DR. Posttransplant microbiological surveillance. Clin Infect Dis 2001; 33 Suppl 1: S22-25
- Sun HY, Cacciarelli TV, Singh N. Identifying a targeted population at high risk for infections after liver transplantation in the MELD era. Clin Transplant 2011; 25: 420-425.
- 41) RUSSELL DL, FLOOD A, ZARODA TE, ACOSTA C, RILEY MM, BUSUTTIL RW, PEGUES DA. Outcomes of colonization with MRSA and VRE among liver transplant candidates and recipients. Am J Transplant 2008; 8: 1737-1743.
- 42) VAN HOEK B, DE ROOU BJ, VERSPAGET HW. Risk factors for infection after liver transplantation. Best Pract Res Clin Gastroenterol 2012; 26: 61-72.
- FISHMAN JA, ISSA NC. Infection in organ transplantation: risk factors and evolving patterns of infection. Infect Dis Clin North Am 2010; 24: 273-283.
- 44) FAGIUOLI S, COLLI A, BRUNO R, CRAXI A, GAETA GB, GROSSI P, MONDELLI MU, PUOTI M, SAGNELLI E, STEFANI S, TONIUTTO P, BURRA P, GROUP AST. Management of infections pre- and post-liver transplantation: report of an AISF consensus conference. J Hepatol. 2014; 60: 1075-1089.
- 45) LAUTENSCHLAGER I. CMV infection, diagnosis and antiviral strategies after liver transplantation. Transpl Int 2009; 22: 1031-1040.
- 46) Moseley RH, Wang W, Takeda H, Lown K, Shick L, Ananthanarayanan M, Suchy FJ. Effect of endotoxin

- on bile acid transport in rat liver: a potential model for sepsis-associated cholestasis. Am J Physiol 1996; 271: G137-146.
- 47) BALMER ML, SLACK E, DE GOTTARDI A, LAWSON MA, HAPFELMEIER S, MIELE L, GRIECO A, VAN VLIERBERGHE H, FAHRNER R, PATUTO N, BERNSMEIER C, RONCHI F, WYSS M, STROKA D, DICKGREBER N, HEIM MH, McCOY KD, MACPHERSON AJ. The liver may act as a firewall mediating mutualism between the host and its gut commensal microbiota. Sci Transl Med 2014; 6: 237ra66.
- 48) Bolder U, Ton-Nu HT, Schteingart CD, Frick E, Hofmann AF. Hepatocyte transport of bile acids and organic anions in endotoxemic rats: impaired uptake and secretion. Gastroenterology 1997; 112: 214-225.
- 49) CHANG FY, SINGH N, GAYOWSKI T, WAGENER MM, MARINO IR. Fever in liver transplant recipients: changing spectrum of etiologic agents. Clin Infect Dis 1998; 26: 59-65.
- 50) Lefkowitch JH. Bile ductular cholestasis: an ominous histopathologic sign related to sepsis and "cholangitis lenta". Hum Pathol 1982; 13: 19-24.
- 51) LAMPS LW, PINSON CW, RAIFORD DS, SHYR Y, SCOTT MA, WASHINGTON MK. The significance of microabscesses in liver transplant biopsies: a clinicopathological study. Hepatology 1998; 28: 1532-1537.
- 52) ARNOLD JC, PORTMANN BC, O'GRADY JG, NAOUMOV NV, ALEXANDER GJ, WILLIAMS R. Cytomegalovirus infection persists in the liver graft in the vanishing bile duct syndrome. Hepatology 1992; 16: 285-292.
- 53) WALDMAN WJ, KNIGHT DA, ADAMS PW, OROSZ CG, SED-MAK DD. In vitro induction of endothelial HLA class II antigen expression by cytomegalovirus-activated CD4+ T cells. Transplantation 1993; 56: 1504-1512.
- 54) Heidenhain C, Pratschke J, Puhl G, Neumann U, Pascher A, Veltzke-Schlieker W, Neuhaus P. Incidence of and risk factors for ischemic-type biliary lesions following orthotopic liver transplantation. Transpl Int 2010; 23: 14-22.
- 55) Shaked A, Ghobrial RM, Merion RM, Shearon TH, Emond JC, Fair JH, Fisher RA, Kulik LM, Pruett TL, Terrault NA, Group AAS. Incidence and severity of acute cellular rejection in recipients undergoing adult living donor or deceased donor liver transplantation. Am J Transplant 2009; 9: 301-308.
- 56) BANFF WORKING G, DEMETRIS AJ, ADEYI O, BELLAMY CO, CLOUSTON A, CHARLOTTE F, CZAJA A, DASKAL I, EL-MONAYERI MS, FONTES P, FUNG J, GRIDELLI B, GUIDO M, HAGA H, HART J, HONSOVA E, HUBSCHER S, ITOH T, JHALA N, JUNGMANN P, KHETTRY U, LASSMAN C, LIGATO S, LUNZ JG, 3RD, MARCOS A, MINERVINI MI, MOLNE J, NALESNIK M, NASSER I, NEIL D, OCHOA E, PAPPO O, RANDHAWA P, REINHOLT FP, RUIZ P, SEBAGH M, SPADA M, SONZOGNI A, TSAMANDAS AC, WERNERSON A, WU T, YILMAZ F. LIVER biopsy interpretation for causes of late liver allograft dysfunction. Hepatology 2006; 44: 489-501.
- 57) Oo YH, ADAMS DH. The role of chemokines in the recruitment of lymphocytes to the liver. J Autoimmun 2010; 34: 45-54.

- 58) KAHRAMAN A, BARREYRO FJ, BRONK SF, WERNEBURG NW, MOTT JL, AKAZAWA Y, MASUOKA HC, HOWE CL, GORES GJ. TRAIL mediates liver injury by the innate immune system in the bile duct-ligated mouse. Hepatology 2008; 47: 1317-1330.
- 59) EKSTEEN B, AFFORD SC, WIGMORE SJ, HOLT AP, ADAMS DH. Immune-mediated liver injury. Semin Liver Dis 2007; 27: 351-366.
- 60) BRAIN JG, ROBERTSON H, THOMPSON E, HUMPHREYS EH, GARDNER A, BOOTH TA, JONES DE, AFFORD SC, VON ZGLINICKI T, BURT AD, KIRBY JA. Biliary epithelial senescence and plasticity in acute cellular rejection. Am J Transplant 2013; 13: 1688-1702.
- 61) Lunz JG 3<sup>®</sup>, Contrucci S, Ruppert K, Murase N, Fung JJ, Starzl TE, Demetris AJ. Replicative senescence of biliary epithelial cells precedes bile duct loss in chronic liver allograft rejection: increased expression of p21(WAF1/Cip1) as a disease marker and the influence of immunosuppressive drugs. Am J Pathol 2001; 158: 1379-1390.
- 62) VAN DEN HEUVEL MC, DE JONG KP, BOOT M, SLOOFF MJ, POPPEMA S, GOUW AS. Preservation of bile ductules mitigates bile duct loss. Am J Transplant 2006; 6: 2660-2671.
- 63) FARKASH EA, COLVIN RB. Diagnostic challenges in chronic antibody-mediated rejection. Nat Rev Nephrol 2012; 8: 255-257.
- 64) PATEL R, TERASAKI PI. Significance of the positive crossmatch test in kidney transplantation. N Engl J Med 1969; 280: 735-739.
- 65) GUGENHEIM J, AMOROSA L, GIGOU M, FABIANI B, ROUGER P, GANE P, REYNES M, BISMUTH H. Specific absorption of lymphocytotoxic alloantibodies by the liver in inbred rats. Transplantation 1990; 50: 309-313.
- 66) ASTARCIOGLU I, CURSIO R, REYNES M, GUGENHEIM J. Increased risk of antibody-mediated rejection of reduced-size liver allografts. J Surg Res 1999; 87: 258-262.
- 67) MATHEW JM, SHENOY S, PHELAN D, LOWELL J, HOWARD T, MOHANAKUMAR T. Biochemical and immunological evaluation of donor-specific soluble HLA in the circulation of liver transplant recipients. Transplantation 1996; 62: 217-223.
- 68) DAR W, AGARWAL A, WATKINS C, GEBEL HM, BRAY RA, KOKKO KE, PEARSON TC, KNECHTLE SJ. Donor-directed MHC class I antibody is preferentially cleared from sensitized recipients of combined liver/kidney transplants. Am J Transplant 2011; 11: 841-847.
- 69) TANER T, GANDHI MJ, SANDERSON SO, POTERUCHA CR, DE GOEY SR, STEGALL MD, HEIMBACH JK. Prevalence, course and impact of HLA donor-specific antibodies in liver transplantation in the first year. Am J Transplant 2012; 12: 1504-1510.
- 70) WIEBE C, GIBSON IW, BLYDT-HANSEN TD, KARPINSKI M, Ho J, STORSLEY LJ, GOLDBERG A, BIRK PE, RUSH DN, NI-CKERSON PW. Evolution and clinical pathologic correlations of de novo donor-specific HLA antibody post kidney transplant. Am J Transplant 2012; 12: 1157-1167.
- 71) REBELLATO LM, EVERLY MJ, HAISCH CE, OZAWA M, BRILEY KP, PARKER K, CATROU PG, BOLIN P, KENDRICK WT, KENDRICK SA, HARLAND RC. A report of the epidemiolo-

- gy of de novo donor-specific anti-HLA antibodies (DSA) in "low-risk" renal transplant recipients. Clin Transpl 2011: 337-340.
- 72) O'LEARY JG, KANEKU H, JENNINGS LW, BANUELOS N, SUSSKIND BM, TERASAKI PI, KLINTMALM GB. Preformed class II donor-specific antibodies are associated with an increased risk of early rejection after liver transplantation. Liver Transpl 2013; 19: 973-980.
- 73) Kasahara M, Kiuchi T, Takakura K, Uryuhara K, Egawa H, Asonuma K, Uemoto S, Inomata Y, Ohwada S, Morishita Y, Tanaka K. Postoperative flow cytometry crossmatch in living donor liver transplantation: clinical significance of humoral immunity in acute rejection. Transplantation 1999; 67: 568-575.
- 74) SCORNIK JC, SOLDEVILLA-PICO C, VAN DER WERF WJ, HEMMING AW, REED AI, LANGHAM MR JR, HOWARD RJ. Susceptibility of liver allografts to high or low concentrations of preformed antibodies as measured by flow cytometry. Am J Transplant 2001; 1: 152-156.
- 75) BISHARA A, BRAUTBAR C, EID A, SCHERMAN L, ILAN Y, SAFADI R. Is presensitization relevant to liver transplantation outcome? Hum Immunol 2002; 63: 742-750.
- 76) MATINLAURI IH, HOCKERSTEDT KA, ISONIEMI HM. Equal overall rejection rate in pre-transplant flow-cytometric cross-match negative and positive adult recipients in liver transplantation. Clin Transplant 2005; 19: 626-631.
- 77) CASTILLO-RAMA M, CASTRO MJ, BERNARDO I, MENEU-DIAZ JC, ELOLA-OLASO AM, CALLEJA-ANTOLIN SM, ROMO E, MORALES P, MORENO E, PAZ-ARTAL E. Preformed antibodies detected by cytotoxic assay or multibead array decrease liver allograft survival: role of human leukocyte antigen compatibility. Liver Transpl 2008; 14: 554-562.
- 78) Lunz J, Ruppert KM, Cajaiba MM, Isse K, Bentlejewski CA, Minervini M, Nalesnik MA, Randhawa P, Rubin E, Sasatomi E, de Vera ME, Fontes P, Humar A, Zeevi A, Demetris AJ. Re-examination of the lymphocytotoxic crossmatch in liver transplantation: can C4d stains help in monitoring? Am J Transplant 2012; 12: 171-182.
- 79) GOH A, SCALAMOGNA M, DE FEO T, POLI F, TERASAKI PI. Human leukocyte antigen crossmatch testing is important for liver retransplantation. Liver Transpl 2010; 16: 308-313.
- 80) Ruiz R, Tomiyama K, Campsen J, Goldstein RM, Levy MF, McKenna GJ, Onaca N, Susskind B, Tillery GW, Klintmalm GB. Implications of a positive crossmatch in liver transplantation: a 20-year review. Liver Transpl 2012; 18: 455-460.
- 81) Shin M, Moon HH, Kim JM, Park JB, Kwon CH, Kim SJ, Lee SK, Joh JW. Significance of true-positive and false-positive pretransplantation lymphocytotoxic crossmatch in primary liver allograft outcomes. Transplantation 2013; 95: 1410-1417.
- 82) Banff schema for grading liver allograft rejection: an international consensus document. Hepatology 1997; 25: 658-663.
- 83) Demetris AJ, Bellamy C, Hubscher SG, O'Leary J, Randhawa PS, Feng S, Neil D, Colvin RB, McCau-

- GHAN G, FUNG JJ, DEL BELLO A, REINHOLT FP, HAGA H, Adeyi O, Czaja AJ, Schiano T, Fiel MI, Smith ML, Sebagh M, Tanigawa RY, Yilmaz F, Alexander G, BAIOCCHI L, BALASUBRAMANIAN M, BATAL I, BHAN AK, BUCUVALAS J, CERSKI CT, CHARLOTTE F, DE VERA ME, ElMonayeri M, Fontes P, Furth EE, Gouw AS, Ha-FEZI-BAKHTIARI S, HART J, HONSOVA E, ISMAIL W, ITOH T, JHALA NC, KHETTRY U, KLINTMALM GB, KNECHTLE S, Koshiba T, Kozlowski T, Lassman CR, Lerut J, Le-VITSKY J, LICINI L, LIOTTA R, MAZARIEGOS G, MINERVINI MI, Misdraji J, Mohanakumar T, Molne J, Nasser I, NEUBERGER J, O'NEIL M, PAPPO O, PETROVIC L, RUIZ P, SAGOL O, SANCHEZ FUEYO A, SASATOMI E, SHAKED A, SHILLER M, SHIMIZU T, SIS B, SONZOGNI A, STEVENSON HL, Thung SN, Tisone G, Tsamandas AC, Wernerson A, Wu T, Zeevi A, Zen Y. 2016 Comprehensive Update of the Banff Working Group on Liver Allograft Pathology: Introduction of Antibody-Mediated Rejection. Am J Transplant 2016 Jun 7. doi: 10.1111/ajt.13909. [Epub ahead of print].
- 84) ABRAHAM SC, FREESE DK, ISHITANI MB, KRASINSKAS AM, Wu TT. Significance of central perivenulitis in pediatric liver transplantation. Am J Surg Pathol 2008; 32: 1479-1488.
- 85) Krasinskas AM, Demetris AJ, Poterucha JJ, Abraham SC. The prevalence and natural history of untreated isolated central perivenulitis in adult allograft livers. Liver Transpl 2008; 14: 625-632.
- ADAMS DH, SANCHEZ-FUEYO A, SAMUEL D. From immunosuppression to tolerance. J Hepatol 2015; 62: S170-185.
- 87) BJORNEBOE M, IVERSEN O, OLSEN S. Infective hepatitis and toxic jaundice in a municipal hospital during a five-year period. Incidence and prognosis. Acta Med Scand 1967; 182: 491-501.
- 88) MALCHOW-MOLLER A, MATZEN P, BJERREGAARD B, HILDEN J, HOLST-CHRISTENSEN J, STAEHR JOHANSEN T, ALTMAN L, THOMSEN C, JUHL E. Causes and characteristics of 500 consecutive cases of jaundice. Scand J Gastroenterol 1981; 16: 1-6.
- 89) WHITEHEAD MW, HAINSWORTH I, KINGHAM JG. The causes of obvious jaundice in South West Wales: perceptions versus reality. Gut 2001; 48: 409-413.
- 90) BJORNSSON E, ISMAEL S, NEJDET S, KILANDER A. Severe jaundice in Sweden in the new millennium: causes, investigations, treatment and prognosis. Scand J Gastroenterol 2003; 38: 86-94.
- VUPPALANCHI R, LIANGPUNSAKUL S, CHALASANI N. Etiology of new-onset jaundice: how often is it caused by idiosyncratic drug-induced liver injury in the United States? Am J Gastroenterol 2007; 102: 558-562; quiz 693.
- 92) SEMBERA S, LAMMERT C, TALWALKAR JA, SANDERSON SO, POTERUCHA JJ, HAY JE, WIESNER RH, GORES GJ, ROSEN CB, HEIMBACH JK, CHARLTON MR. Frequency, clinical presentation, and outcomes of drug-induced liver injury after liver transplantation. Liver Transpl 2012; 18: 803-810.
- BJORNSSON E, OLSSON R. Outcome and prognostic markers in severe drug-induced liver disease. Hepatology 2005; 42: 481-489.

- 94) ANDRADE RJ, LUCENA MI, FERNANDEZ MC, PELAEZ G, PACHKORIA K, GARCIA-RUIZ E, GARCIA-MUNOZ B, GONZALEZ-GRANDE R, PIZARRO A, DURAN JA, JIMENEZ M, RODRIGO L, ROMERO-GOMEZ M, NAVARRO JM, PLANAS R, COSTA J, BORRAS A, SOLER A, SALMERON J, MARTIN-VIVALDI R, SPANISH GROUP FOR THE STUDY OF DRUG-INDUCED LIVER D. Drug-induced liver injury: an analysis of 461 incidences submitted to the Spanish registry over a 10-year period. Gastroenterology 2005: 129: 512-521.
- 95) CHALASANI N, FONTANA RJ, BONKOVSKY HL, WATKINS PB, DAVERN T, SERRANO J, YANG H, ROCHON J, DRUG INDUCED LIVER INJURY N. Causes, clinical features, and outcomes from a prospective study of drug-induced liver injury in the United States. Gastroenterology 2008; 135: 1924-1934, 1934 e1-4.
- 96) DE VALLE MB, AV KLINTEBERG V, ALEM N, OLSSON R, BJORNSSON E. Drug-induced liver injury in a Swedish University hospital out-patient hepatology clinic. Aliment Pharmacol Ther 2006; 24: 1187-1195.
- 97) Benichou C, Danan G, Flahault A. Causality assessment of adverse reactions to drugs--II. An original model for validation of drug causality assessment methods: case reports with positive rechallenge. J Clin Epidemiol 1993; 46: 1331-1336.
- 98) Danan G, Benichou C. Causality assessment of adverse reactions to drugs--I. A novel method based on the conclusions of international consensus meetings: application to drug-induced liver injuries. J Clin Epidemiol 1993; 46: 1323-1330.
- 99) http://livertox.nlm.nih.gov.
- 100) Kowdley KV, Keeffe EB. Hepatotoxicity of transplant immunosuppressive agents. Gastroenterol Clin North Am 1995; 24: 991-1001.
- 101) KLINTMALM GB, IWATSUKI S, STARZL TE. Cyclosporin A hepatotoxicity in 66 renal allograft recipients. Transplantation 1981; 32: 488-489.
- 102) LORBER MI, VAN BUREN CT, FLECHNER SM, WILLIAMS C, KAHAN BD. Hepatobiliary and pancreatic complications of cyclosporine therapy in 466 renal transplant recipients. Transplantation 1987; 43: 35-40.
- 103) LAUPACIS A, KEOWN PA, ULAN RA, SINCLAIR NR, STIL-LER CR. Hyperbilirubinaemia and cyclosporin A levels. Lancet 1981; 2: 1426-1427.
- 104) CHAN FK, SHAFFER EA. Cholestatic effects of cyclosporine in the rat. Transplantation 1997; 63: 1574-1578.
- 105) KADMON M, KLUNEMANN C, BOHME M, ISHIKAWA T, GORGAS K, OTTO G, HERFARTH C, KEPPLER D. Inhibition by cyclosporin A of adenosine triphosphate-dependent transport from the hepatocyte into bile. Gastroenterology 1993; 104: 1507-1514.
- 106) MORAN D, DE BUITRAGO JM, FERNANDEZ E, GALAN AI, MUNOZ ME, JIMENEZ R. Inhibition of biliary glutathione secretion by cyclosporine A in the rat: possible mechanisms and role in the cholestasis induced by the drug. J Hepatol 1998; 29: 68-77.

- 107) STIEGER B, FATTINGER K, MADON J, KULLAK-UBLICK GA, MEIER PJ. Drug- and estrogen-induced cholestasis through inhibition of the hepatocellular bile salt export pump (Bsep) of rat liver. Gastroenterology 2000; 118: 422-430.
- 108) CADRANEL JF, ERLINGER S, DESRUENNE M, LUCIANI J, LU-NEL F, GRIPPON P, CABROL A, OPOLON P. Chronic administration of cyclosporin A induces a decrease in hepatic excretory function in man. Dig Dis Sci 1992; 37: 1473-1476.
- 109) SANCHEZ-CAMPOS S, LOPEZ-ACEBO R, GONZALEZ P, CU-LEBRAS JM, TUNON MJ, GONZALEZ-GALLEGO J. Cholestasis and alterations of glutathione metabolism induced by tacrolimus (FK506) in the rat. Transplantation 1998; 66: 84-88.
- 110) TANIAI N, AKIMARU K, ISHIKAWA Y, KANADA T, KAKINU-MA D, MIZUGUCHI Y, MAMADA Y, YOSHIDA H, TAJIRI T. Hepatotoxicity caused by both tacrolimus and cyclosporine after living donor liver transplantation. J Nippon Med Sch 2008; 75: 187-191.
- 111) ERICZON BG, EUSUFZAI S, SODERDAHL G, DURAJ F, EI-NARSSON K, ANGELIN B. Secretion and composition of bile after human liver transplantation: studies on the effects of cyclosporine and tacrolimus. Transplantation 1997; 63:74-80.
- 112) NAVARRO VJ, SENIOR JR. Drug-related hepatotoxicity. N Engl J Med 2006; 354: 731-739.
- 113) LOUPY A, ANGLICHEAU D, MAMZER-BRUNEEL MF, MARTINEZ F, THERVET E, LEGENDRE C, SERPAGGI J, POL S. Mycophenolate sodium-induced hepatotoxicity: first report. Transplantation 2006; 82: 581.
- 114) JACQUES J, DICKSON Z, CARRIER P, ESSIG M, GUILLAUDE-AU A, LACOUR C, BOCQUENTIN F, ALDIGIER JC, REPOLLE JP. Severe sirolimus-induced acute hepatitis in a renal transplant recipient. Transpl Int 2010; 23: 967-970.
- 115) SPARBERG M, SIMON N, DEL GRECO F. Intrahepatic cholestasis due to azathioprine. Gastroenterology 1969; 57: 439-441.
- 116) DEPINHO RA, GOLDBERG CS, LEFKOWITCH JH. Azathioprine and the liver. Evidence favoring idiosyncratic, mixed cholestatic-hepatocellular injury in humans. Gastroenterology 1984; 86: 162-165.
- 117) STERNECK M, WIESNER R, ASCHER N, ROBERTS J, FERRELL L, LUDWIG J, LAKE J. Azathioprine hepatotoxicity after liver transplantation. Hepatology 1991; 14: 806-810.
- 118) GANE E, PORTMANN B, SAXENA R, WONG P, RAMAGE J, WILLIAMS R. Nodular regenerative hyperplasia of the liver graft after liver transplantation. Hepatology 1994; 20: 88-94.
- 119) Kowdley KV, Keeffe EB, Fawaz KA. Prolonged cholestasis due to trimethoprim sulfamethoxazole. Gastroenterology 1992; 102: 2148-2150.
- 120) Romagnuolo J, Sadowski DC, Lalor E, Jewell L, Thomson AB. Cholestatic hepatocellular injury with azathioprine: a case report and review of the mechanisms of hepatotoxicity. Can J Gastroenterol 1998; 12: 479-483.
- (21) BEN SALEM C, BEN SALAH L, BELAJOUZA C, BOURAOUI K. Azathioprine-induced severe cholestatic hepati-

- tis in patient carrying TPMT\*3C polymorphism. Pharm World Sci 2010; 32: 701-703.
- 122) Roda G, Caponi A, Belluzzi A, Roda E. Severe cholestatic acute hepatitis following azathioprine therapy in a patient with ulcerative pancolitis. Dig Liver Dis 2009; 41: 914-915.
- 123) Bastida G, Nos P, Aguas M, Beltran B, Rubin A, Dasi F, Ponce J. Incidence, risk factors and clinical course of thiopurine-induced liver injury in patients with inflammatory bowel disease. Aliment Pharmacol Ther 2005; 22: 775-782.
- 124) BJORNSSON ES, HOOFNAGLE JH. Categorization of drugs implicated in causing liver injury: Critical assessment based on published case reports. Hepatology 2016; 63: 590-603.
- 125) NAVARRO VJ, KHAN I, BJORNSSON E, SEEFF LB, SER-RANO J, HOOFNAGLE JH. Liver injury from herbal and dietary supplements. Hepatology 2017; 65: 363-373.
- 126) COHEN PA. Hazards of hindsight--monitoring the safety of nutritional supplements. N Engl J Med 2014; 370: 1277-1280.
- 127) Andrade RJ, Lucena MI, Kaplowitz N, Garcia-Munoz B, Borraz Y, Pachkoria K, Garcia-Cortes M, Fernandez MC, Pelaez G, Rodrigo L, Duran JA, Costa J, Planas R, Barriocanal A, Guarner C, Romero-Gomez M, Munoz-Yague T, Salmeron J, Hidalgo R. Outcome of acute idiosyncratic drug-induced liver injury: Long-term follow-up in a hepatotoxicity registry. Hepatology 2006; 44: 1581-1588.
- 128) MORADPOUR D, ALTORFER J, FLURY R, GREMINGER P, MEYEN-BERGER C, JOST R, SCHMID M. Chlorpromazine-induced vanishing bile duct syndrome leading to biliary cirrhosis. Hepatology 1994; 20: 1437-1441.
- 129) OLSSON R, WIHOLM BE, SAND C, ZETTERGREN L, HULTCRANTZ R, MYRHED M. Liver damage from flucloxacillin, cloxacillin and dicloxacillin. J Hepatol 1992; 15: 154-161.
- 130) GREGORY DH, ZAKI GF, SARCOSI GA, CAREY JB. Chronic cholestasis following prolonged tolbutamide administration. Arch Pathol 1967; 84: 194-201.
- 131) GLOBER GA, WILKERSON JA. Biliary cirrhosis following the administration of methyltestosterone. JAMA 1968; 204: 170-173.
- 132) Ishii M, Miyazaki Y, Yamamoto T, Miura M, Ueno Y, Takahashi T, Toyota T. A case of drug-induced ductopenia resulting in fatal biliary cirrhosis. Liver 1993; 13: 227-231.
- 133) Eckstein RP, Dowsett JF, Lunzer MR. Flucloxacillin induced liver disease: histopathological findings at biopsy and autopsy. Pathology 1993; 25: 223-228.
- 134) BJORNSSON E, DAVIDSDOTTIR L. The long-term follow-up after idiosyncratic drug-induced liver injury with jaundice. J Hepatol 2009; 50: 511-517.
- 135) RAMOS AM, GAYOTTO LC, CLEMENTE CM, MELLO ES, LUZ KG, FREITAS ML. Reversible vanishing bile duct syndrome induced by carbamazepine. Eur J Gastroenterol Hepatol 2002; 14: 1019-1022.

- 136) VUPPALANCHI R, CHALASANI N, SAXENA R. Restoration of bile ducts in drug-induced vanishing bile duct syndrome due to zonisamide. Am J Surg Pathol 2006; 30: 1619-1623.
- 137) STINE JG, LEWIS JH. Current and future directions in the treatment and prevention of drug-induced liver injury: a systematic review. Expert Rev Gastroenterol Hepatol 2016; 10: 517-536.
- 138) SEBAGH M, SAMUEL D, ANTONINI TM, COILLY A, DEGLI ESPOSTI D, ROCHE B, KARAM V, DOS SANTOS A, DUCLOS-VALLEE JC, ROQUE-AFONSO AM, BALLOT E, GUETTIER C, BLANDIN F, SALIBA F, AZOULAY D. Twenty-year protocol liver biopsies: invasive but useful for the management of liver recipients. J Hepatol 2012; 56:840-847.
- 139) HUBSCHER SG. What is the long-term outcome of the liver allograft? J Hepatol 2011; 55: 702-717.
- 140) SEBAGH M, RIFAI K, FERAY C, YILMAZ F, FALISSARD B, ROCHE B, BISMUTH H, SAMUEL D, REYNES M. All liver recipients benefit from the protocol 10-year liver biopsies. Hepatology. 2003; 37: 1293-1301.
- 141) Hubscher SG. Transplantation pathology. Semin Liver Dis 2009; 29: 74-90.
- 142) Muro M, Marin L, Miras M, Moya-Quiles R, Mingue-La A, Sanchez-Bueno F, Bermejo J, Robles R, Ramirez P, Garcia-Alonso A, Parrilla P, Alvarez-Lopez MR. Liver recipients harbouring anti-donor preformed lymphocytotoxic antibodies exhibit a poor allograft survival at the first year after transplantation: experience of one centre. Transpl Immunol 2005; 14: 91-97.
- 143) O'LEARY JG, KANEKU H, SUSSKIND BM, JENNINGS LW, NERI MA, DAVIS GL, KLINTMALM GB, TERASAKI PI. High mean fluorescence intensity donor-specific anti-HLA antibodies associated with chronic rejection Postliver transplant. Am J Transplant 2011; 11: 1868-1876.
- 144) KANEKU H, O'LEARY JG, TANIGUCHI M, SUSSKIND BM, TERASAKI PI, KLINTMALM GB. Donor-specific human leukocyte antigen antibodies of the immunoglobulin G3 subclass are associated with chronic rejection and graft loss after liver transplantation. Liver Transpl 2012; 18: 984-992.
- 145) Neil DA, Hubscher SG. Current views on rejection pathology in liver transplantation. Transpl Int 2010; 23: 971-983.
- 146) WYATT JI. Liver transplant pathology-messages for the non-specialist. Histopathology 2010; 57: 333-341.
- 147) DEMETRIS A, ADAMS D, BELLAMY C, BLAKOLMER K, CLOUSTON A, DHILLON AP, FUNG J, GOUW A, GUSTAFSSON B, HAGA H, HARRISON D, HART J, HUBSCHER S, JAFFE R, KHETTRY U, LASSMAN C, LEWIN K, MARTINEZ O, NAKAZAWA Y, NEIL D, PAPPO O, PARIZHSKAYA M, RANDHAWA P, RASOUL-ROCKENSCHAUB S, REINHOLT F, REYNES M, ROBERT M, TSAMANDAS A, WANLESS I, WIESNER R, WERNERSON A, WRBA F, WYATT J, YAMABE H. Update of the International Banff Schema for Liver Allograft Rejection: working recommendations for the histopathologic staging and reporting of chronic rejection. An International Panel. Hepatology 2000; 31: 792-799.