

Prostate cancer: radioresistance molecular target-related markers and foreseeable modalities of radiosensitization

C. ALBERTI

L.D. of Surgical Semeiotics, Parma, Italy

Abstract. – OBJECTIVES: Though the external beam radiation therapy is a standard treatment option for both organ-confined and regionally advanced prostate cancer, unluckily, despite more and more effective advances in radiation delivery procedures, the prostate cancer radioresistance still occurs in a significant amount of patients undergone radiotherapy. This review aims to highlight the molecular aberrations of prostate cancer cell growth- and apoptosis signaling pathways that might induce, together with both prostate cancer cell/cancer stem cells gene- and surrounding microenvironment crucial implications, the tumor radioresistance.

METHODS: An up-dated review of several thorough studies on such matter.

RESULTS: The plenty of intriguing knowledge acquisitions about the prostate cancer radioresistance depending on cancer cell growth/apoptosis signaling pathway-related molecular aberrations together with prostate cancer cell/cancer stem cell abnormal gene profile, may be the premise leading – on the basis of preclinical research in animal models – to clinically overcome the tumor radioresistance.

CONCLUSIONS: Current developments of radiosensitizer agents focusly targeting prostate cancer cell radioresistance-associated specific molecular/gene aberrations are directed to improve, by implementing customized tumor radiosensitization modalities, the radiation therapy outcomes.

Key words:

Cell growth pathway, Apoptosis pathway, Cancer stem cells, Radiosensitizers.

Introduction

Though the general cancer incidence rates, during the five-year period, slightly declined in North American men, by 0.6% per year, with male cancer death rates lowering by 1.8% per

year, and were, instead, stable in women with a female cancer death decrease by 1.4% per year, the *prostate carcinoma* (PCa) remains a considerable health burden in developed countries, it representing, after the skin tumors, the most common cancer in men, about 1 man in 7 running into PCa onset during lifetime. According to American Cancer Society, PCa is the second leading cause of cancer death in North-American men, after the lung cancer¹. In European Union, the PCa incidence sets at fourth site among the most common malignancies including female breast-, colorectal-, lung tumors². Organ-confined PCa affects around 70% of cases whose the majority shows low-to-intermediate disease clinico-pathologic features³.

External beam radiation therapy (EBRT) is a standard treatment option for both organ-confined and regionally advanced PCa, while brachytherapy strategies, with either 125 I or 103 Pd low dose rate (LDR) permanent implant and 192 Ir high dose rate (HDR) temporary implant, may be applied to treat low-to-intermediate risk intraprostatic tumor.

Despite radiation delivery technological advances, the rate of biochemical/clinical relapse for a significant number of PCa patients undergone radiotherapy unfortunately remains high³⁻⁶. Given that gene expression profiles may predict, for PCa patients, an individual possible radioresistance, the identification of aberrant both cancer cell growth or death signaling pathway-related specific biomarkers can implement customized radiosensitization modalities by the resort to molecular targeted agents (7). Moreover, the more and more thorough characterization of prostate cancer stem cells (PCSCs) suggest that they may significantly be responsible for both radio- and chemoresistance, hence it following that PCSC radioresistant subpopulation-targeting strategies can improve PCa radiotherapy⁷⁻⁹.

Aberrant cell growth signaling pathway-related prostate cancer radioresistance

Radioresistant prostate cancer cells often exhibit high epidermal growth factor (EGF) receptor levels, hence resulting a considerable activation of phosphatidylinositol 3-kinase (PI3K)-Akt and mammalian target of rapamycin (mTOR) – PI3K-Akt/mTOR – pathway, besides mitogen-activated protein kinase (MAPK) and Janus tyrosine kinase (Jak)/signal transducer activator of transcription (STAT) – Jak-STAT – pathways¹⁰⁻¹². Otherwise, even the exposure of cells to ionizing radiations may activate such signaling pathways – among which particularly the MAPK/ERK (extracellular-regulated kinase)-mediated one – with various downstreams that result to be dependent on different expression of cell plasma-membrane growth factor receptors and influence of autocrine ligands, such as transforming growth factor-alpha or beta, or even triggered by Ras mutations¹⁰⁻¹⁴.

DOC2/DAB2 interactive protein (DAB2IP), that normally has a control over the PI3K/Akt pathway, is often down-regulated in aggressive PCa, from it resulting the onset of PCa cell radioresistance due to easier DNA double-strand break repair – so it maintaining the telomere stability – together with apoptosis evasion. Therefore, adjuvant treatment with *NU7441*, as a novel agent leading to ineffective DNA repair besides blocking overactivated PI3K-Akt/mTOR pathway, can overcome DAB2IP loss-induced PCa cell radioresistance^{15,16}. Moreover, radiosensitizing effects may be achieved by targeting PI3K-Akt/mTOR pathway by imidazol-quinoline derivative *NVP-BEZ 235*, which, in addition, has the potential to inhibit the expression of HIF-1alpha (hypoxia-inducible factor-1alpha), another intriguing target for tumor radiosensitization^{11,13,17,18}.

Regarding the Jak-STAT persistent activation-promoted PCa cell survival against ionizing radiation, the radiosensitization could be reached by block such pathway by resorting to *AG 490*, a STAT 3 inhibitor⁸.

As for cell cycle modulation and DNA repair, P53 gene, through its own codified p53 transcriptional factor protein, plays a central role by regulating the expression of different genes concerning either cell growth or apoptosis, that's why P53 deletion/mutation-induced loss of p53 functions may promote dramatic effects, among which the tumorigenesis with development of cancer cell radioresistance. What's particularly due to increased interactions between p53 and

MDM2 (mouse double minute 2) protein, that, when aberrantly overexpressed as it sometimes occurs under the radiation stress, may induce the p53 inactivation, through its ubiquitination and intraproteasome degradation, with subsequent lack of p53-dependent normal functions. It follows that the use of drugs preventing the p53-MDM2 interactions – such as the anticancer *Nutlins*, cis-imidazole analogs – can lead, by allowing the p53 accumulation/activation, to the p53-mediated efficient block of cancer cells growth meanwhile with activation of their apoptosis mechanisms¹⁹⁻²². Otherwise, the P53 gene mutations can themselves make radioresistant the cancer cells, that's why the restoration of wild-P53 can induce their radiosensitization as it has been shown in human PCa xenograft animal models⁵⁻¹². Moreover, given that Reprimo (RPRM) tumor suppressor gene is normally involved in p53-mediated radiation-due G2/M phase cell cycle arrest, an aberrant epigenetic methylation of DNA-RPRM promoter region, can facilitate both tumor progression and apoptosis evasion together with development of cancer cell radioresistance. In this regard, the nucleotide analog *5-azacytidine*, as methyl-transferase inhibitor, can avoid the aberrant DNA-RPRM silencing, thus it radiosensitizing the cancer cells^{23,24}.

Aberrant cell death signaling pathway-related prostate cancer radioresistance

For the androgen-sensitive prostate carcinoma, the androgen deprivation therapy (ADT) leads to apoptosis of hormone-dependent cancer cells meanwhile unfortunately selecting out, after an average of 18 to 22 months, hormone-refractory cell clones, thus the cancer cell apoptotic evasion contributing, together with abnormal cancer cell proliferation, to tumor progression. Ionizing radiation can not only act on the nucleus-leaving signalling pathways, by inducing the DNA damage, but also by activating, even at the plasma membrane level, the apoptotic process-driving one^{25,26}.

The apoptotic mechanism encompasses crucial molecular events particularly involving two distinct routes, where various caspases (interleukin converting enzyme, ICE-like cysteine proteases) have an important role: the one – intrinsic/cell surface receptor-independent, mitochondrial activity involving, pathway – including translocation of Bax (Bcl 2-associated x-protein) from cytosol into mitochondria, hence activation of mitochondrial protein Bak (Bcl 2 antagonist killer) with subsequent release of several mito-

chondrial pro-apoptotic proteins such as cytochrome C and SMAC (second mitochondria-derived activator of caspases), that lead to activate the procaspase 9 – caspase 9 – caspase 3 sequence, while the other – extrinsic/cell surface receptor-dependent pathway – resulting from stress/cytotoxic agent (among which ionizing radiation)-induced activation of transmembrane “death receptors”, including Fas/APO 1 (apoptosis inducing protein 1), also known TNFRSF6 (tumor necrosis factor-receptor super-family 6) as crosslinked by tumor necrosis factor-alpha (TNF alpha). The apoptotic signal down-stream emerging from such plasma membrane receptor activation, sequentially includes procaspase 8 – caspase 8, down to caspase 3, that, as a crucial key protease mediator of both intrinsic and extrinsic pathways, promotes the proteolytic cleavage of poly(ADP-ribose)polymerase-1(PARP-1), thus allowing the apoptotic process closing endonuclease-mediated DNA fragmentation²⁷⁻²⁹.

The anti-apoptotic gene Bcl 2(B-cell lymphoma) overexpression may promote an aggressive behaviour of PCa cells with their heavy radio/chemo-resistance, that's why a recently identified Bcl 2 inhibitor *HA14-1* can improve the cancer cells apoptosis meanwhile enhancing their radiosensitivity(30). Otherwise, the clusterin, a glycoprotein overexpressed in various malignancies, protects, by interfering with Bax proapoptotic activity, the cancer cells from TGF beta(transforming growth factor-beta)-promoted apoptotic mechanisms, it following that, by down-regulating its expression through specific antisense nucleotide *OGX-011*, the apoptosis might be restored together with cancer cell radio- and chemo-sensitivity^{15,31-33}. Furthermore, also the survivin can facilitate the cancer cell survival – though upon cell death stimuli such as ionizing radiation – by interfering with the caspase activity, hence it resulting that the survivin inhibitor *YM155* can sensitize PCa cells to radiation⁹. Otherwise, the over-expression of cell surface membrane integrin”alpha v beta 3” significantly prevents the radiation-induced downregulation of survivin, that's why *cRGDfV*, as an antagonists of such integrin and survivin-mediated anti-apoptotic signaling, may allow the achievement of radiation-promoted PCa cell apoptosis³⁴.

Given that the above-mentioned PARP-1 may play an antiapoptotic role, as preventing caspase/endonuclease-induced DNA fragmentation, its block by specific inhibitors – such as *veliparib*, *rucaparib*, *niraparib* and particularly *olaparib* –

can maximize the DNA damage-related cancer cell death, so it reaching an effective PCa cell radiosensitization in both EBRT and alpha-emitter Ra 223 radiopharmaceutical treatment^{35,36}.

Among various molecular factors involved in the apoptotic process, the *ceramide* plays a crucial role just regarding its changeable conditions under radiation treatment. Indeed, the radiation stress promotes the membrane-associated sphingomyelin hydrolysis with generation of ceramide that acts as a potent proapoptotic mediator by inhibiting Bcl 2 protein-induced mitochondrial depolarization, therefore the ceramide production during radiotherapy predicting a treatment favourable outcome^{37,38}. Unfortunately, the surrounding tumor microenvironment ceramide accumulation, induces, by feed-back, the acid ceramidase (N-acylsphingosine amidohydrolase) gene up-regulation, that, in turn, leads to production of ceramide catabolite sphingosine and its mitogenic phosphorylated derivative sphingosine-1-phosphate which, besides its ineffectiveness to maintain the ceramide's role in the apoptosis, may activate the Akt pathway, hence it enhancing cancer cells proliferation together with supporting their radioresistance. It follows that acid ceramidase proteolytic degradation promoters, such as lysosomotropic agents *LCL 521* and *LCL 385*, could maintain the ceramide-associated apoptotic process meanwhile radiosensitizing PCa cells^{39,40}. In addition, even the *toremifene*, a structurally tamoxifen-like antiestrogen, has been recognized to be an efficacious inhibitor of the acid ceramidase activity, thus its use allowing to restore both the cancer cells death and PCa radiosensitization⁴¹.

Unlike cancer cell apoptosis, the autophagy – as lysosomal machinery driving to ubiquitin-mediated degradation of cell's own components and cytoplasm sequestration into autophagosomes with vacuolated appearance – maintains cells alive in response to different stress stimuli, including the ionizing radiation, what contributing to cancer cell radioresistance. Therefore, autophagy blockers, such as a lysosomotropic anti-malarian chloroquine-like *bafilomycin*, may act as cancer cell radiosensitizers^{8,9}.

Cancer stem cells and radioresistance

The prostate cancer growth, as well as the any tumor one, is driven by the subpopulation of prostate cancer stem cells (PCSCs) – also called tumor-initiating cells – that, in addition, provide a cellular reservoir to promote tumor recurrence after therapy^{18,42}.

Today's more and more reliable characterization of PCSCs – by taking advantage from stem cell sphere-forming assay – particularly with identification of their specific markers expression, suggests that they are highly responsible for both radio- and chemoresistance, by enhancing the cancer cell proliferation and supporting the apoptosis evasion^{8,9,43}. PCSCs, indeed, by their ATR (Ataxia-telangiectasia mutated/Rad-related kinase protein)-mediated DNA repair mechanisms, make sure their own survival, in niche-guarded quiescent state (late “S” cell cycle phase), against ionizing radiation, compared, instead, with the high radio- and chemosensitivity of proliferating cells in G2/M phase^{8,42,44}. Otherwise, the specific gene mutation-dependent over-activation of stem cell specific pathways – such as Wnt/beta catenin-, Hedgehog- and Notch pathways – play an important role in facilitating both PCSCs self-renewal and radioresistance. As for the beta-catenin, its abnormal accumulation, resulting from over-activated Wnt signaling, can increase the PCa progression, as well as of other tumors, by enhancing the specific cell growth-dependent transcriptional activity meanwhile supporting the production of survivin that, in turn, can interfere, as it has been above mentioned, with the apoptosis-associated caspase protein family; that's why the survivin inhibitor *YMI55* may induce anticancer-radiosensitizer effects⁹. Furthermore, the *perifosine*, besides blocking the Akt pathway, can also inhibit the Wnt signaling with following restoration of tumor radiation sensitivity⁸. On the other hand, the aberrant activation of Hedgehog pathway seems to lead to development of various malignancies, including PCa, through the transformation of adult stem cells into PCSCs, which, because of own inside decrease in E-cadherin (E-calcium-dependent adhesion glycoprotein) and, instead, increase in pro-cell growth cyclins, give rise to a tumor onset with radioresistance. Regarding the Notch pathway – as a family of cell transmembrane protein-receptors – its dysregulation, apart from maintaining the cancer cell self-renewal stemness, can facilitate the tumor radioresistance onset together with metastasis^{45,46}.

Recent intriguing studies have allowed to identify some PCSC radioresistance-related genes, such as PCSC-1 and PSCS-3 RAN (Ras-associated nuclear protein)-signaling genes, that are involved in the DNA synthesis and in cell cycle promotion. It follows that the PCSC-1 and

PCSC-3 RAN signaling genes might be an important target to hopefully reach the PCa radiosensitization⁹. In addition, CXCR4 (chemokine CXC motif receptor 4) has been recently recognized as a biomarker for both drug- and radioresistant cancer stem cells, the interaction of such receptor with its ligand (CXCL12) playing a crucial role in protecting them from anticancer agents and radiation. Therefore, a feasible therapeutic block of the CXCR4/CXCL12 signaling pathway should represent a promising opportunity to refine the PCa radiation therapy⁴⁷.

Surrounding microenvironment-dependent tumor radioresistance

Different both cell components of tumor surrounding microenvironment – such as fibroblasts, endothelial cells and vascular network, infiltrating immune cells – and bioactive factors, including growth factors, cytokines, hormones, variable level of oxygen/ROS (reactive oxygen species, generated by radiation-induced ionization of water molecules), can significantly influence the cancer growth, particularly with the involvement of CSC behaviour. In this regard, it has been highlighted that fibroblast- and endothelial cell-made niches of CSCs, protect them from radiation stress with following up-regulation of CSC-associated specific signaling pathways – such as Wnt/beta catenin-, Hedgehog- and Notch pathways – that accelerate the CSC self-renewal^{8,9}.

Given that oxygen is a potent ROS-mediated tumor radiosensitizer, a hypoxic microenvironment, with subsequent low ROS levels, prevents cancer cells from radiation-induced oxidative DNA damage, so driving them to radioresistance, moreover their survival advantage also resulting from HIF 1alpha expression-mediated inhibition of apoptotic process⁴⁸⁻⁵⁰. The significance of antitumor DNA oxidative damage-mediated effects – at least partly – of radiation and also of some anticancer agents, appears to be understandable considering that their activity results to be often made ineffective by ROS-scavengers – such as N-acetylcysteine or glutathione or even dihydrolipoic acid – what recently has been shown by the evaluation of Altrenol anticancer activity in several PCa cell lines⁵¹. On the other hand, the microenvironmental ROS-scavenger glutathione depletion by *buthionine-sulfoximine* can reverse the radioresistant cancer cell condition⁸.

To increase the radiation effects in hypoxic cancer cells, may be useful the resort to administration of the imidazol-quinoline derivative *NVP-BEZ 235*, which, a part from the block on PI3K/mTOR pathway, as it has been above mentioned, can also inhibit the HIF1alpha in hypoxic cancer cells, so improving the radiotherapy outcomes¹⁷.

Some significant biomarkers of prostate cancer radioresistance

The identification of cancer cell radioresistance-related predictive biomarkers allows the potential of either select alternative treatment modalities or, at least, plan a radiation therapy in combination with specific radiosensitizer agents. Besides the above mentioned cell growth/cell death signaling pathway-related molecular aberrations, recent research results suggest that acquisition of EMT (epithelial-mesenchymal transition) from cancer cells is highly indicative of their radioresistance. Such cancer cell occurrence is characterized, indeed, by a down-expression of E-cadherin compared with the mesenchyme-peculiar expression of vimentin¹⁸.

A significant predictor of PCa cell- and, particularly, of PCSC-radioresistance is represented by their CD44-v6 marker, that appears to be positively significant when microenvironmental basal or radiation-induced ROS levels are relatively low⁽⁸⁾. Knock down of CD44-v6 in PCa cell lines can suppress, by following down-regulation of both Wnt/beta catenin- and PI3K-Akt/mTOR signaling pathways together with drop in EMT, tumor cell proliferation meanwhile enhancing PCa cell line radio- and chemosensitivity⁵².

Besides the CD44-v6, other biomarkers, particularly the PCSC-associated ones – such as CD133, ABCG2 – may be a crucial factor for a radio-curative option⁴³.

The discovery of PCSC-peculiar radioresistance genes, among which the PCSC 1- and PCSC 2 RAN signaling ones, provides a further explanation of post-radiotherapy PCa recurrence (9,53). In addition, even the above mentioned CXCRA over-expression from PCSCs is highly indicative of PCa radioresistance, that's why the foreseeable inhibition of CXCRA/CXC ligand pathway might play a radiosensitizer role⁴⁷.

In the field of today's normal laboratory procedures, an elevated, immunoreactivity-tested, PCa cell Bcl2/Bax ratio, may be predictive of a worse response to radiation, it, for a long time now, suggesting the usefulness of such biomarker for prognostic evaluation of PCa radiotherapy⁵⁴.

Concluding remarks and new directions

In the last decades, more and more interesting and effective EBRT advances – from the 3D-conformal radiotherapy to the intensity-modulated one and even to the image-guided radiation delivery – together with adopting a suitable dose hypofractionation modality given that low alpha/beta PCa cell radiosensitivity ratio, have significantly improved, compared to the past, both biochemical and clinical outcomes for patients with organ-confined/locally advanced PCa undergone to radiation therapy^{4,6,55,56}. Nevertheless, despite progress in the EBRT procedures, the PCa radioresistance – with the disease progression or local recurrence, both emerging by PSA's levels continuously rising above the radiation therapy-reached nadir and pointed up by prostatic biopsy – still occurs up to third of PCa patients undergone radiotherapy^{3,11,37,57}. More thoroughly, according to recent data^{58,59}, the biopsy-defined primary Gleason (pG) grade may represent an independent predictor for biochemical recurrence-free survival (BRfs), distant metastasis(DMfs)and PCa-specific mortality (PCa-SM), as 8-year BRfs rate for pG3 amounting 77.6% versus pG4 (61.3%), pG3-DMfs 96.8% versus pG4 (84.3%), pG3 PCa-SM 3.7% versus pG4 81.00%.

The role of different kinase hyperactivity – such as of PI3K, MAPK/ERK, Janus tyrosine kinase – in sometimes inducing PCa radioresistance has been highlighted in detail as far as the aberrations of cell growth signaling pathway^{10-14,17}, as well as the antiapoptotic-radioresistance promoting effects of Bcl-2, clusterin, survivin and, in addition, HIF1 alpha – compared with proapoptotic effects of Bax, Bak, caspase sequence – have been clearly taken into consideration in regard to aberrant cell death signaling pathway^{25-38,49,50}.

Current genetic profile/transcriptional pattern studies can provide reliable specific data predictive of radiotherapy long-term outcomes, thus it allowing to identify PCa patients who, as not expected to be responsive to radiation, might be undergone an alternative therapy such as surgery or hormone/anticancer chemotherapeutic agents⁵. In this regard, EBRT combination with either neoadjuvant- or adjuvant ADT/androgen receptor antagonists, such as the novel enzalutamide, can sometimes lead to more effective PCa local control and disease-free survival, with particularly favourable prognosis for high-risk PCa patients^{4,22,60,61}.

It's of the utmost importance to take into consideration that in elderly cancer patients, different comorbidities can influence both the acute and late

radio- or combined radio-chemotherapy dependent toxicity, it leading to carry out customized radiotherapy/chemotherapy treatments⁶²⁻⁶⁴.

The implications of PCSCs in the tumor radioresistance development, with local recurrence and metastatic spread after radiotherapy, make more and more useful the resort to the identification of radioresistance-peculiar PCSC genes, such as the PCSC1- and PCSC2-RAN ones, highly predictive of the radiotherapy ineffectiveness⁸.

In the field of gene therapy, radiosensitizer siRNA-based novel bio-approaches are today under study, particularly to suppress CSC-associated specific signaling pathways^{8,9,45,46}.

Foreseeably, further developments of *customized* radiosensitizers – focusly targeting the highlighted PCa cell radioresistance-linked specific molecular aberrations – will improve the PCa radiotherapy outcome^{4,6,9}. In addition to various radiosensitizers acting at above-mentioned different signaling pathway levels, novel anti-cancer agents, such as cytoskeletal microtubule-stabilizing epothilone B-derived drugs, have been preclinically recognized to be provided with high potential of radiosensitizing various cancer cell types, among which the PCa cell ones, by inducing the G2-M cell cycle arrest and tumor growth delay⁶⁵⁻⁶⁷.

On the basis of deepened radiobiological features – such as high linear energy transfer, radiobiological effectiveness, tumor dose delivery targeting, tumor/healthy tissue damage ratio – both hadron beam proton/neutron- and carbon ion radiation-therapy seem to successfully overcome both tumor intrinsic and extrinsic radioresistance classical conditions (CSC-related risk, crucial alpha/beta cancer cell sensitivity ratio, microenvironmental hypoxia, etc.) meanwhile better sparing pelvic organs in comparison with EBRT⁶⁸⁻⁷².

Conflict of Interest

The Author declares that he has no conflict of interests.

References

- 1) SIEGEL R, MA J, ZOU Z, JEMAL A. Cancer statistics. *Ca Cancer J Clin* 2014; 64: 9-29.
- 2) FERLAY J, STELIAROVA-FOUCHER E, LORTET-TIEULENT J, ROSSO S, COEBERGH JWW, COMBER H, BRAY F, FORMAN D. Cancer incidence and mortality patterns in Europe: estimates for 40 countries in 2012. *Eur J Cancer* 2013; 49: 1374-1403.
- 3) CHANG L, GRAHAM PH, HAO J, NI J, BUCCI J, COZZI P, KEARSLEY JH, LI Y. Emerging roles of radioresistance in prostate cancer metastasis and radiation therapy. *Cancer Metastasis Rev* 2014, Jan 21 (Epub ahead of print).
- 4) POLKINGHORN WR, ZELEFSKY MI. Improving outcomes in high-risk prostate cancer with radiotherapy. *Rep Pract Oncol Radiother* 2013; 18: 333-337.
- 5) AGEMY L, KELA I, WAKS T, PFEFFER RM, BAR-SHIRA A, ORR-URTREGER A, DOMANY E, ESHHAR Z. Gene expression profiles predict sensitivity of prostate cancer to radiotherapy. *J Cancer Ther* 2013; 4: 11-26.
- 6) HAUSTERMANS K, ISEBAERT S. Prostate cancer: what is the next step for the radiation oncologist? *P Belg Roy Acad Med* 2013; 2: 111-126.
- 7) GHOTRA VP, GELDOLF AA, DANEN EH. Targeted radiosensitization in prostate cancer. *Curr Pharm Des* 2013; 19: 2819-2828.
- 8) CHUMSRI A, SHAH P. Radiation resistance of cancer stem cells as an obstacle in cancer therapy. *Mol Cell Pharmacol* 2013; 5: 39-49.
- 9) RYCAJ K, TANG DG. Cancer stem cells and radioresistance. *Int J Radiat Biol* 2014 Mar 7 (Epub ahead of print).
- 10) SKVORTSOVA I, SKVORTSOV S, STASYK T, RAIU U, POPPER B, SCHIESTI B, VON GUGGENBERG E, NEHER A, BONN GK, HUBER LA, LUKAS P. Intracellular signaling pathways regulating radioresistance of human prostate carcinoma cells. *Proteomics* 2008; 8: 4521-4533.
- 11) CHANG L, GRAHAM PH, HAO J, BUCCI J, COZZI PJ, KEARSLEY JH, LI Y. Acquisition of EMT and cancer stem cell phenotypes is associated with activation of PI3K/Akt/mTOR pathway in prostate cancer radioresistance. *Cell Death Dis* 2013; 4: e875.
- 12) LEHMANN BD, MCCUBREY JA, TERRIAN DM. Radiosensitization of prostate cancer by priming the wild-type p53-dependent cellular senescence pathway. *Cancer Biol Ther* 2007; 6: 1165-1170.
- 13) DENT P, YACCOUB A, FISHER PB, HAGAN MP, GRANT S. MAPK pathways in radiation responses. *Oncogene* 2003; 22: 5885-5896.
- 14) AFFOLTER A, DRIGOTAS M, FRUTH K, SCHMIDTMANN I, BROCHHAUSEN C, MANN W, BRIEGER J. Increased radioresistance via G12SK-Ras by compensatory upregulation of MAPK and PI3K pathways in epithelial cancer. *Head Neck* 2013; 35: 220-228.
- 15) WU K, XIE D, ZOU Y, ZHANG T, PONG RC, XIAO G, FAZLI L, GLEAVE M, HE D, BOOTHMAN D, HSIEH JT. The mechanism of DAB2IP in chemoresistance of prostate cancer cells. *Clin Cancer Res* 2013; 19: 4740-4749.
- 16) YU L, TUMATI V, TSENG SF, HSU FM, KIM D, HONG D, HSIEH JT, JACOBS C, KAPUR P, SAHA D. DAB 21P regulates autophagy in prostate cancer response to combined treatment of radiation and DNA-PKcs inhibitor. *Neoplasia* 2012; 14: 1203-1212.
- 17) KUGER S, COREK E, POLAT B, KAMMERER U, FLENTJE M, DJUZENOVA CS. Novel PI3K and mTOR inhibitor

- NVP-BEZ235 radiosensitizes breast cancer cell lines under normoxic and hypoxic conditions. *Breast Cancer* 2014; 8: 39-49.
- 18) XIAO W, GRAHAM PH, POWER CA, HAO J, KEARSLEY JH, LI Y. CD44 is a biomarker associated with human prostate cancer radiation sensitivity. *Clin Exp Metastasis* 2012; 29: 1-9.
 - 19) CHÈNE P. Inhibition of the p53-MDM2 interaction:targeting a protein-protein interface. *Mol Cancer Res* 2004; 2: 20-28.
 - 20) VASSILEV LT, VU B, GRAVES B, CARVAJAL D, PODIASKI F, FILIPOVIC Z, KONG N, KAMMIOTT U, LUKACS C, KLEIN C, FOTOUHI N, LIU EA. In vivo activation of the p53 pathway by small-molecule antagonists of MDM2. *Science* 2004;303:844-848.
 - 21) IMPICCIATORE G, SANCILLO S, MISCIA S, DI PIETRO R. Nutlins and ionizing radiation in cancer therapy. *Curr Pharm Des* 2010; 16: 1427-1442.
 - 22) WU CT, CHEN WC, LIAO SK, HSU CL, LEE KD, CHEN MF. The radiation response of hormone resistant prostate cancer induced by long-term hormone therapy. *Endocrin Relat Cancer* 2007; 14: 633-643.
 - 23) MURPHY TM, PERRY AS, LAWIER M. The emergence of DNA methylation as a key modulator of aberrant cell death in prostate cancer. *Endocr Relat Cancer* 2008; 15: 11-25.
 - 24) JIANG W, LI YQ, LIU A, SUN Y, HE QM, JIANG N, XU YF, CHEN L, MA J. 5-Azacytidine enhances the radiosensitivity of CNE2 and SUNE1 cells in vitro and in vivo possibly by altering DNA methylation. *PLoS ONE* 2014; 9: e93273.
 - 25) CHEN T, CHEN M, CHEN J. Ionizing radiation potentiates dihydroartemisin-induced apoptosis of A549 cells via a caspase-8 dependent pathway. *PLoS ONE* 2013; 8: e59827.
 - 26) WATTERS D. Molecular mechanisms of ionizing radiation-induced apoptosis. *Immunol Cell Biol* 1999;77:263-271.
 - 27) YU J, ZHANG L. Apoptosis in human cancer cells. *Curr Opin Oncol* 2003; 16: 19-24.
 - 28) ZIMMERMANN KG, BONZON C, GREEN DR. The machinery of programmed cell death. *Pharmacol Ther* 2001; 92: 57-70.
 - 29) FRISH SM, SCREATON RA. Anoikis mechanisms. *Curr Opin Biol* 2001; 13: 555-562.
 - 30) AN J, CHERVIN AS, NIE A, DUCCOFF HS, HUANG Z. Overcoming the radioresistance of prostate cancer cells with a novel Bcl2 inhibitor. *Oncogene* 2007; 26: 652-661.
 - 31) ZHU B, KYPRIANOU N. Transforming growth factor beta and prostate cancer. *Cancer Treat Res* 2005; 126: 157-173.
 - 32) MCKENZIE S, KYPRIANOU N. Apoptosis evasion: the role of survival pathways in prostate cancer progression and therapeutic radioresistance. *J Cell Biochem* 2006; 97: 18-32.
 - 33) FRISH SM, RUOSLAHTI E. Integrins and anoikis. *Curr Opin Cell Biol* 1997; 9: 701-706.
 - 34) WANG T, HUANG J, ALAVIAN MR, GOEL HL, PLESCIA J, ALTIERI DC, LANGUINO LR, FITZGERALD TJ. Alpha v beta3 integrin controls survivin expression and promotes resistance of prostate cancer cells to ionizing radiation. *ASCO-Genitourinary Cancer Symposium, Orlando, US, 2009 February, 26-28.*
 - 35) ZHANG J. Poly(ADP-ribose)polymerase inhibitor: an involving paradigm in the treatment of prostate cancer. *Asian J Androl* 2014; 16: 401-406.
 - 36) JELINIC P, LEVINE DA. New insights of PARP inhibitors' effects on cell cycle and homology-directed DNA damage repair. *Mol Cancer Ther* 2014; 13: 1645-1654.
 - 37) BECKHAM TH, CHENG JC, LU P, SHAO Y, TROYER D, LANCE R, MARRISON ST, NORRIS JS, LIU X. Acid ceramidase induces sphingosine kinase 1/S1P receptor-2-mediated activation of oncogenic Akt signaling. *Oncogenesis* 2013; 2: e49.
 - 38) KOLESNICK R, FUKS Z. Radiation and ceramide-induced apoptosis. *Curr Opin Cell Biol* 2003; 22: 5897-907.
 - 39) LIU X, CHENG JC, TURNER LS, ELOJEIMI S, BECKHAM TH, BIELAWSKA A, KEANE TE, HANNUN YA, NORRIS JS. Acid ceramidase upregulation in prostate cancer: role in tumor development and implications for therapy. *Expert Opin Ther Targets* 2009; 13: 1449-1458.
 - 40) BECKHAM TH, CHENG JC, LU P, MARRISON S, NORRIS JS, LIU X. Acid ceramidase promotes nuclear export of PTEN through sphingosine-phosphate mediated Akt signaling. *PLoS ONE* 2013; oct 1.
 - 41) MORAD SA, LEVIN JC, TAN SF, FOX TE, FEITH DJ, CABOT MC. Novel off-target effect of tamoxifen inhibition of acid ceramidase activity in cancer cells. *Biochim Biophys Acta* 2013; 1831: 1656-664.
 - 42) KIM YS, KANG MJ, CHO YM. Low production of ROS and high DNA repair mechanism of radioresistance of prostate cancer stem cells. *Anticancer Res* 2013; 33: 4469-4474.
 - 43) CASTELLON EA, VALENZUELA R, LILIO J, CASTILLO V, CONTRERAS R, GALLEGOS I, MERCADO A, HUIDOBRO C. Molecular signature of cancer stem cells isolated from prostate carcinoma and expression of stem markers in different Gleason grades and metastasis. *Biol Res* 2012; 45: 297-305.
 - 44) LOVEJOY CA, CORTEZ D. Common mechanisms of PIKK regulation. *DNA Repair* 2009; 8: 1004-1008.
 - 45) HU YY, ZHENG MH, ZHANG R, LIANG Y, HAN H. Notch signaling pathway and cancer metastasis. *Adv Exp Med Biol* 2012; 727: 186-198.
 - 46) YAO Y, WANG L, ZHANG H, WANG H, ZHAO X, WHANG Y, ZHANG Y, ZHANG L, FAN X, QIAN G, HU JF, GE S. A novel anticancer therapy that simultaneously targets aberrant p53 and Notch activity in tumors. *PLoS ONE* 2012; 7: e46627.
 - 47) TRAUTMANN F, COJOC M, KURTH I, MELIN N, BOUCHEZ LC, DUBROVSKA A, PEITZSCH C. CXCR4 as biomarker for radioresistant cancer stem cells. *Int J Radiat Biol* 2014 (Epub ahead of print).
 - 48) DUHAGON MA, HURT EM, SOTELO-SILVEIRA JR, ZHANG X, FARRAR WT. Genomic profiling of tumor initiating prostatespheres. *BMC Genomics* 2010; 11: 324-330.

- 49) HENNESSEY D, MARTIN LM, ATZBERGER A, LYNCH TH, HOLLYWOOD D, MARIGNOL L. Exposure to hypoxia following irradiation increases radioresistance in prostate cancer cells. *Urol Oncol* 2013; 31: 1106-1116.
- 50) ALBERTI C. Prostate cancer progression and surrounding microenvironment. *Int J Biol Markers* 2006; 21: 88-95.
- 51) TANG Y, CHEN R, HUANG Y, LI G, HUANG Y, CHEN J, DUAN L, ZHU BT, THRASHER JB, ZHANG X, LI B. Natural compound Alternol induce oxidative stress-dependent apoptotic cell death in prostate cancer cells. *Mol Cancer Ther* 2014; 13: 1526-1536.
- 52) NI J, COZZI PJ, HAO JL, BERETOV J, CHANG L, DUAN W, SHIGDAR S, DELPRADO W, GRAHAM PH, BUCCI J. CD44 variant 6 is associated with prostate cancer metastasis and chemo-/radioresistance. *Prostate* 2014; 74: 602-617.
- 53) SKVORTSOV S, DEBBAGE P, CHO WC, LUKAS P, SKVORTSOVA I. Putative biomarkers and therapeutic targets associated with radiation resistance. *Expert Rev Proteomics* 2014; 11: 207-214.
- 54) MACKAY TJ, BORKOWSKI A, AMIN P, JACOBS S, KYPRIANOU N. Bcl 2 /Bax ratio as a predictive marker for therapeutic response to radiotherapy in patients with prostate cancer. *Urology* 1998; 52: 1085-1090.
- 55) WONG WM, WALLNER KE. The case of hypofractionation of localized prostate cancer. *Rev Urol* 2013; 15: 113-117.
- 56) MORGIA G, DE RENZIS C. Cyberknife in the treatment of prostate cancer: a revolutionary system. *Eur Urol* 2009; 56: 40-42.
- 57) HEIDENREICH A, RICHTER S, THUER D, PLISTER D. Prognostic parameters, complications and oncologic and functional outcome of salvage radical prostatectomy for locally recurrent prostate cancer after 21st-century radiotherapy. *Eur Urol* 2010; 57: 437-443.
- 58) SPRATT DE, ZUMSTEG Z, GHADJAR P, PANGASA M, PEI X, FINE SW, YAMADA Y, KOLLMEIER M, ZELEFSKY MJ. *Int J Radiat Oncol Biol Phys* 2013; 85: 1254-1261.
- 59) ZUMSTEG ZS, SPRATT DE, PEI I, ZHANG Z, YAMADA Y, KOLLMEIER M, ZELEFSKY MJ. A new risk classification system for therapeutic decision making with intermediate-risk prostate cancer patients undergoing dose-escalated external-beam radiation therapy. *Eur Urol* 2013; 64: 895-902.
- 60) MUNOZ F, GUARNERI A, BOTTICELLA A, GABRIELE P, MORETTO F, PANAIÀ R, RUGGIERI A, D'URSO L, MUTO G, FILIPPI AR, RAGONA R, RICARDI U. Salvage external beam radiotherapy for recurrent prostate adenocarcinoma after high-intensity focused ultrasound as primary treatment. *Urol Int* 2013; 90: 288-293.
- 61) CAMBRIA R, CATTANI F, JERECZEK-FOSSA BA, PANSINI F, CIARDO D, VIGORITO S, RUSSO S, ZERINI D, COZZI L, ORECCHIA R. Planning study to compare dynamic and rapid arc techniques for postprostatectomy radiotherapy of prostate cancer. *Strahlenther Onkol* 2014; 190: 799-805.
- 62) FIORICA F, BERRETTA M, COLOSIMO C, BERRETTA S, RISTAGNO M, PALMUCCI S, LLESHI A, URSINO S, FISICHELLA R, SPARTÀ D, STEFANELLI A, CAPPELLANI A, TIRELLI U, CARTEI F. Safety and efficacy of radiotherapy treatment in elderly patients with localized prostate cancer: a retrospective analysis. *Arch Geront Geriatr* 2010; 51: 277-282.
- 63) FIORICA F, STEFANELLI A, FISICHELLA R, TIRELLI U, BERRETTA M. Comorbidity assessment and radiotherapy in elderly cancer patients. *Eur Rev Med Pharmacol Sci* 2012; 16: 1605-1606.
- 64) MORGIA G, RUSSO GL, BERRETTA M, PRIVITERA S, KIRKALI Z. Genito-urological cancers in elderly patients. *Anticancer Agents Med Chem* 2013; 13: 1391-1405.
- 65) DAVSON NA. Epothilones in prostate cancer: review of clinical experience. *Ann Oncol* 2007; 18(Suppl. 5): 22-27.
- 66) KONG Z, RAGHAVAN P, XIE D, BOIKE T, BURMA S, CHEN D, CHAKRABORTY A, HSIEH JT, SAHA D. EpothiloneB confers radiation dose enhancement in DAB2IP gene knock-down radiosensitive prostate cancer cells. *Int J Radiat Oncol Biol Phys* 2010; 78: 1210-1218.
- 67) ALBERTI C. Taxane- and epothilone-based chemotherapy: from molecule cargo cytoskeletal logistics to management of castration-resistant prostate carcinoma. *Eur Rev Med Pharmacol Sci* 2013; 17: 1658-1664.
- 68) SNYDER M, JOINER MC, KONSKI A, BOSSENBERGER T, BURMEISTER J. Dose escalation in prostate cancer using intensity modulated neutron radiotherapy. *Radiation Oncol* 2011; 99: 201-206.
- 69) FELLIN F, AZZERONI R, MAGGIO A, LORENTINI S, COZZARINI C, DI MUZIO N, FIORINO C, CALANDRINO R, SCHWARZ M. Helical tomotherapy and intensity modulated proton therapy in the treatment of dominant intraprostatic lesion: a treatment planning comparison. *Radiation Oncol* 2013; 107: 207-212.
- 70) MOLINELLI S, MAIRANI A, MIRANDOLA A, VILCHES-FREIXAS G, TESSONIER T, GIORDANENGO S, PARODI K, CIOCCA M, ORECCHIA R. Dosimetric accuracy assessment of treatment plan verification system for scanned proton beam radiotherapy: one-year experimental results and Monte Carlo analysis of the involved uncertainties. *Phys Med Biol* 2013; 58: 3837-3847.
- 71) SCHIAFF CD, KRAUZE A, BELARD A, O'CONNELL JJ, CAMPHAUSEN KA. Bringing the heavy: carbon ion therapy in the radiobiological and clinical context. *Radiation Oncol* 2014; 9: 88.
- 72) ALBERTI C. Organ-confined prostate carcinoma radiation brachytherapy compared with external either photon- or hadron-beam radiation therapy. Just a short up-to-date. *Eur Rev Med Pharmacol Sci* 2011; 15: 769-774.