

Prevalence of plasmid-mediated *qnr* determinants and gyrase alteration in *Klebsiella pneumoniae* isolated from a university teaching hospital in Malaysia

A.S. SAIFUL ANUAR, M.Y. MOHD YUSOF, S.T. TAY

Department of Medical Microbiology, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia

Abstract. – BACKGROUND AND OBJECTIVES:

The ciprofloxacin resistance of *Klebsiella* (*K.*) *pneumoniae* is mediated primarily through alterations in type II topoisomerase (*gyrA*) gene and plasmid-mediated quinolone resistance-conferring genes (*qnr*).

This study aimed to define the prevalence of plasmid-mediated quinolone resistance-conferring genes (*qnr*) and type II topoisomerase (*gyrA*) alterations of a population of ciprofloxacin-resistant ($n = 21$), intermediate ($n = 8$), and sensitive ($n = 18$) *K. pneumoniae* isolates obtained from a teaching hospital at Kuala Lumpur, Malaysia.

MATERIALS AND METHODS: A multiplex PCR assay was performed for simultaneous detection of *qnrA*, *qnrB* and *qnrS*. Sequence analysis of the amplified *gyrA* and *gyrB* regions of the isolates were performed.

RESULTS: The findings in this study revealed the emergence of a high prevalence (48.9%) of *qnr* determinants in our isolates. Four variants of plasmid-mediated *qnr* determinants (*qnrB1*, *qnrB6*, *qnrB10* and *qnrS1*) were detected from 11 (52.4%) ciprofloxacin-resistant, 5 (62.5%) intermediate and 7 (38.9%) sensitive isolates. *gyrA* alterations were detected from 18 (85.7%) ciprofloxacin-resistant isolates. Single *gyrA* alterations, Ser83→Tyr, Ser83→Ile, and Asp87→Gly, and double alterations, Ser83→Phe plus Asp87→Ala and Ser83→Tyr plus Asp87→Asn were detected. While ciprofloxacin resistance was significantly associated with *gyrA* alteration (Ser83, $p = 0.003$; Asp87, $p = 0.005$; double alteration, $p = 0.016$), no significant association of ciprofloxacin resistance was noted with the presence of *qnr* determinants ($p = 0.283$).

CONCLUSIONS: The findings in this study demonstrate the emergence of *qnr* determinants and *gyrA* alterations contributed to the development and spread of fluoroquinolone resistance in the Malaysian isolates.

Key Words:

Klebsiella pneumoniae, Gyrase A, Plasmid-mediated *qnr* determinants, Ciprofloxacin.

Introduction

Klebsiella (*K.*) *pneumoniae* is a frequent cause of urinary tract infections, nosocomial pneumonia, and intra-abdominal infections amongst hospitalized patients. In recent years, emergence of ciprofloxacin-resistant *K. pneumoniae* isolates has been widely reported in Europe, North America and Asia¹, along with the upsurge of resistance to other antibiotics. The ciprofloxacin resistance of *K. pneumoniae* is mediated primarily through alterations in type II topoisomerase (*gyrA*) gene or changes in the drug entry and efflux^{2,3}. More recently, low-level resistance to quinolones conferred by plasmid-mediated quinolone resistance-conferring genes (*qnr*) has been reported¹. *qnr* genes protect DNA gyrase and topoisomerase from inhibition by quinolones and have been often associated with genes that confer resistance to other classes of antibiotics (e.g. β -lactams and aminoglycosides). In addition, isolates with *qnr* resistance may represent a serious concern in the clinical environment as it can contribute to the rapid development and spread of fluoroquinolone resistance.

Limited information is available on the ciprofloxacin-resistance mechanisms of *K. pneumoniae* isolates in the Southeast Asia region, including Malaysia. The aim of this study was to define the prevalence of plasmid-mediated *qnr* resistance and *gyrA* alteration of a population of ciprofloxacin-resistant ($n = 21$), intermediate ($n = 8$), and sensitive ($n = 18$) *K. pneumoniae* clinical isolates from University Malaya Medical Center (UMMC), a Teaching Hospital at Kuala Lumpur, Malaysia.

Materials and Methods

Klebsiella Clinical Isolates

These were multi-drug resistant isolates obtained from various types of clinical specimens in

Table I. Detection of plasmid-mediated *qnr* genes in *K. pneumoniae* isolates.

Ciprofloxacin susceptibility	No. (%) strain with <i>qnr</i> determinants				
	<i>qnrB1</i>	<i>qnrB6</i>	<i>qnrB10</i>	<i>qnrS1</i>	Total
Sensitive (n = 18)	2 (11.1)	0 (0)	1 (5.6)	4 (22.2)	7 (38.9)
Intermediate (n = 8)	2 (25.0)	1 (12.5)	2 (25.0)	0 (0)	5 (62.5)
Resistant (n = 21)	4 (19.0)	1 (4.8)	2 (9.5)	4 (19.0)	11 (52.4)
Total (n = 47)	8 (17.0)	2 (4.3)	5 (10.6)	8 (17.0)	23 (48.9)

the Microbiology Diagnostic Laboratory from 2006-2008. The identity of the isolates was confirmed based on the sequence analysis of the internal transcribed spacer gene region, as described by Wang et al⁴. Minimum inhibitory concentrations (MICs) of the isolates to ciprofloxacin were confirmed using E tests strips (bioMérieux SA, Marcy L'Etoile, France) according to the manufacturer's instructions. Ciprofloxacin resistance was defined as minimum inhibitory concentration (MIC) ≥ 4 $\mu\text{g/ml}$, intermediate as MIC = 2 $\mu\text{g/ml}$ and sensitive as MIC ≤ 1 $\mu\text{g/ml}$ ⁵.

Amplification and Sequence Analysis of Resistance Genes

A multiplex polymerase chain reaction (PCR) assay was performed for the simultaneous detection of *qnrA*, *qnrB* and *qnrS* as described by Cattoir et al⁶, using boiled bacterial extracts as DNA templates. The *gyrA* and *gyrB* regions of the isolates were amplified as described by Ling et al⁷. The amplicons were purified using GeneAll (PCR SV PCR kit, Seoul, Korea) and the subsequent sequencing reaction was performed with the Big Dye[®] Terminator V3.1 Cycle Sequencing Kit (Applied Biosystems, CA, USA) on an ABI-

377 genetic Analyzer (Applied Biosystems, Lincoln Centre Drive, Foster City, CA, USA). The *qnr* sequences were subjected to BLAST search for matching sequences in the GenBank database. The gyrase nucleotide sequences and the deduced amino acids were compared with that of *K. pneumoniae* ATCC (American Type Culture Collection) 13883 (GenBank accession number DQ673325) using ClustalW alignment program.

Statistical Analysis

The association of each resistance mechanism for ciprofloxacin-resistance was analysed using the Mann-Whitney test of the SPSS statistical package (version 13.0) (SPSS Inc., Chicago, IL, USA). A *p* value of < 0.05 was considered statistically significant.

Results

The ciprofloxacin MICs of our isolates ranged from 0.25 $\mu\text{g/ml}$ to ≥ 32 $\mu\text{g/ml}$, with MIC₅₀ and MIC₉₀ being 2 $\mu\text{g/ml}$ and ≥ 32 $\mu\text{g/ml}$, respectively. The ciprofloxacin MICs for resistant isolates were as follows: 4 $\mu\text{g/ml}$, 2 isolates (9.5%); 8

Table II. Alterations in *gyrA* gene regions of *K. pneumoniae* isolates.

Ciprofloxacin susceptibility	No. (%) isolate with <i>gyrA</i> alterations							
	Single alteration	Double alteration	codon Ser83 (TCC)			codon Asp87 (GAC)		
			Tyr (TAC)	Ile (ATC)	Phe (TTC)	Gly (GGC)	Ala (GCC)	Asn (AAC)
Sensitive (n = 18)	5 (27.8)	0 (0)	5 (27.8)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
Intermediate (n = 8)	5 (62.5)	0 (0)	4 (50)	0 (0)	0 (0)	1 (12.5)	0 (0)	0 (0)
Resistant (n = 21)	10 (47.6)	8 (38.1)	6 (28.6)	6 (28.6)	5 (23.8)	1 (4.8)	5 (23.8)	3 (14.3)
Total (n = 47)	20 (42.6)	8 (17.0)	15 (31.9)	6 (12.8)	5 (10.6)	2 (4.3)	5 (10.6)	3 (6.4)

Ser, serine; Asp, Aspartic acid; Tyr, tyrosine; Ile, isoleucine; Phe, phenylalanine; Gly, glycine; Ala, alanine; Asn, asparagine.

µg/ml, 1 isolate (4.8%); 12 µg/ml, 3 isolates (14.3%); and ≥ 32 µg/ml, 15 isolates (71.4%).

The prevalence and diversity of *qnr* determinants of our isolates are shown in Table I. Four *qnr* determinants (*qnrB1*, *qnrB6*, *qnrB10* and *qnrS1*) were identified; with *qnrB* detected more frequently than *qnrS*. No *qnrA* was detected from our isolates. *qnr* determinants were detected in 11 (52.4%) of the ciprofloxacin-resistant isolates, as well as 7 (38.9%) and 5 (62.5%) sensitive and intermediate isolates, respectively. No significant association of ciprofloxacin resistance with the presence of *qnr* was noted ($p = 0.283$). Of 23 *qnr* positive isolates, 12 (52.2%) had *gyrA* alterations. Three ciprofloxacin-resistant isolates (MICs of 8, 12, ≥ 32 µg/ml) with *qnr* determinants (*qnrB6*, *qnrS1*, *qnrB1*) had no *gyrA* alteration.

Alterations in the *gyrA* region, as indicated in Table II, were detected in 28 (59.6%) isolates, of which 20 (42.6%) and 8 (17.0%) had single and double alterations, respectively. The *gyrA* alterations involve codons at the position 83 (Ser) and 87 (Asp). Of the 21 ciprofloxacin-resistant isolates, 10 (47.6%) and 8 (38.1%) had single and double alterations in the *gyrA* region, respectively. Three types of single alterations were noted in the resistant isolates: Ser83→Tyr (3 isolates), Ser83→Ile (5 isolates), and Asp87→Gly (2 isolates). Two types of double alterations were detected in the resistant isolates (all MICs ≥32 µg/ml): Ser83→Phe plus Asp87→Ala (5 isolates), and Ser83→Tyr plus Asp87→Asn (3 isolates). Single alteration (Ser83→Tyr substitution) was the only alteration detected in 5 (27.8%) and 4 (50%) sensitive and intermediate isolates, respectively. When the intermediate isolates were grouped as resistant isolates, significant association of ciprofloxacin resistance was observed with *gyrA* alteration at Ser83 ($p = 0.003$), Asp87 ($p = 0.005$) and double alterations ($p = 0.016$).

Discussion

High prevalences of *qnr* determinants have been reported in many geographical regions¹. This study reported the emergence of a high prevalence (48.9%) of *qnr* determinants in the Malaysian *K. pneumoniae* isolates. Isolates with *qnr* determinants are known to harbor multiple ciprofloxacin resistance mechanisms including alterations in *gyrA*⁸ or decreased drug permeability⁹, thus, facilitating high ciprofloxacin resistance. However, acquisition of *qnr* genes

does not render a wild-type organism fluoroquinolone insusceptible¹. This is also observed in this study whereby no significant association of ciprofloxacin resistance was observed with the presence of *qnr* determinants.

All *K. pneumoniae* isolates with double alterations at the *gyrA* region show high ciprofloxacin MICs (≥ 32 µg/ml) in this study. The *gyrA* alterations detected from 18 (85.7%) of 21 ciprofloxacin-resistant isolates in this study are similar to those reported in other geographical regions such as Japan, North America and Europe^{2,10,11}. Although *gyrA* alteration (Ser83→Leu) has been described in *K. pneumoniae* isolates from two Asian countries, China¹² and Singapore¹³, it was not observed in this study. The alteration in *gyrB* gene has been reported in members of *Enterobacteriaceae* such as *Salmonella*¹⁴ and *Proteus mirabilis*¹⁵. However; it has not been reported for *K. pneumoniae*. Similarly, no *gyrB* alteration was detected in this study.

Conclusions

These findings demonstrate that the emergence of *qnr* determinants and *gyrA* alterations contributed to the development and spread of fluoroquinolone resistance in the Malaysian isolates. A correct use of antibiotics and continuously monitoring of resistance patterns in *K. pneumoniae* in our hospital settings are necessary.

Acknowledgements

This project was funded by the University of Malaya Post-graduate research grants: PS258/2010A, PV086/2011B and RG378/11HTM

Conflict of Interest

None declared.

References

- 1) STRAHILEVITZ J, JACOBY GA, HOOPER DC, ROBICSEK A. Plasmid-mediated quinolone resistance: a multifaceted threat. *Clin Microb Rev* 2009; 22: 664-689.
- 2) DEGUCHI T, FUKUOKA A, YASUDA M, NAKANO M, OZeki S, KANEMATSU E, NISHINO Y, ISHIHARA S, BAN Y, KAWADA Y. Alterations in the *GyrA* subunit of DNA gyrase and the ParC subunit of topoisomerase IV in

- quinolone-resistant clinical isolates of *Klebsiella pneumoniae*. Antimicrob Agents Chemother 1997; 41: 699-701.
- 3) JACOBY GA. Mechanisms of resistance to quinolones. Clin Infect Dis 2005; 41(Suppl 2): S120-126.
 - 4) WANG M, CAO B, YU Q, LIU L, GAO Q, WANG L, FENG L. Analysis of the 16S-23S rRNA gene internal transcribed spacer region in *Klebsiella* Species. J Clin Microb 2008; 46: 3555-3563.
 - 5) CLINICAL AND LABORATORY STANDARDS INSTITUTE (CLSI). Performance standards for antimicrobial susceptibility testing; 19th informational supplement. Wayne, PA. 2009; pp. M100-S19.
 - 6) CATTOIR V, POIREL L, ROTIMI V, SOUSSY C, NORDMANN P. Multiplex PCR for detection of plasmid-mediated quinolone resistance *qnr* genes in ESBL-producing enterobacterial isolates. J Antimicrob Chemother 2007; 60: 394-397.
 - 7) LING JM, CHAN EW, LAM AW, CHENG AF. Alterations in topoisomerase genes of fluoroquinolone-resistant *Salmonella* in Hong Kong. Antimicrob Agents Chemother 2003; 47: 3567-3575.
 - 8) LASCOLS C, ROBERT J, CATTOIR V, BEBEAR C, CAVALLO JD, PODGLAJEN I, PLOY MC, BONNET R, SOUSSY CJ, CAMBAU E. Type II topoisomerase alterations in clinical isolates of *Enterobacter cloacae* and other enterobacterial species harbouring the *qnrA* gene. Int J Antimicrob Agents 2007; 29: 402-409.
 - 9) CHENIA HY, PILLAY B, PILLAY D. Analysis of the mechanisms of fluoroquinolone resistance in urinary tract pathogens. J Antimicrob Chemother 2006; 58: 1274-1278.
 - 10) MARTÍNEZ-MARTÍNEZ L, PASCUAL A, CONEJO MDEL C, GARCÍA I, JOYANES P, DOMÉNECH-SÁNCHEZ A, BENEDI VJ. Energy-dependent accumulation of norfloxacin and porin expression in clinical isolates of *Klebsiella pneumoniae* and relationship to extended-spectrum β -lactamase production. Antimicrob Agents Chemother 2002; 46: 3926-3932.
 - 11) YOSHIDA H, BOGAKI M, NAKAMURA M, NAKAMURA S. Quinolone resistance-determining region in the DNA gyrase *gyrA* gene of *Escherichia coli*. Antimicrob Agents Chemother 1990; 34: 1271-1272.
 - 12) FU Y, GUO L, XU Y, ZHANG W, GU J, XU J, CHEN X, ZHAO Y, MA J, LIU X, ZHANG F. Alteration of *GyrA* amino acid required for ciprofloxacin resistance in *Klebsiella pneumoniae* isolates in China. Antimicrob Agents Chemother 2008; 52: 2980-2983.
 - 13) SCHNEIDERS T, AMYES SG, LEVY SB. Role of *AcrR* and *ramA* in fluoroquinolone resistance in clinical *Klebsiella pneumoniae* isolates from Singapore. Antimicrob Agents Chemother 2003; 47: 2831-2837.
 - 14) GENSBERG K, JIN YF, PIDDOCK LJV. A novel *gyrB* alteration in a fluoroquinolone-resistant clinical isolate of *Salmonella typhimurium*. FEMS Microb Lett 1995; 132: 57-60.
 - 15) WEIGEL LM, ANDERSON GJ, TENOVER FC. DNA gyrase and topoisomerase IV alterations associated with fluoroquinolone resistance in *Proteus mirabilis*. Antimicrob Agents Chemother 2002; 46: 2582-2587.