# Non-thyroidal illness syndrome in chronic diseases: role of irisin as modulator of antioxidants

A. MANCINI<sup>1</sup>, E. CAPOBIANCO<sup>1</sup>, C. BRUNO<sup>1</sup>, E. VERGANI<sup>1</sup>, M. NICOLAZZI<sup>2</sup>, A.M.R. FAVUZZI<sup>2</sup>, N. PANOCCHIA<sup>3</sup>, E. MEUCCI<sup>4</sup>, A. MORDENTE<sup>4</sup>, A. SILVESTRINI<sup>4</sup>

Abstract. - OBJECTIVE: Non-thyroidal-illness syndrome (NTIS) refers to condition found in chronic diseases that is an adaptive mechanism. However, oxidative stress is related to NTIS in a vicious circle, due to deiodinases alteration and negative effects of low T3 on antioxidant levels or activity. Muscle is one of the main targets of thyroid hormones and it can secrete a myokine named irisin, which is able to induce the browning of white adipose tissue, energy expenditure and protect against insulin resistance. Inconclusive data have been reported about irisin role in chronic diseases. Moreover, no correlation with antioxidants has been investigated. Therefore, we performed a case-control study with the primary endpoint to evaluate irisin levels in two models of NTIS, such as chronic heart failure (CHF) and chronic kidney disease (CKD) during haemodialytic treatment. The secondary endpoint was the correlation with total antioxidant capacity (TAC) to establish a possible role of irisin in the modulation of antioxidant

**PATIENTS AND METHODS:** Three groups of subjects were enrolled. Group A included CHF patients (n=18; aged  $70.22 \pm 2.78$  ys; BMI  $\pm 27.75 \pm 1.28$  kg/m²); Group B included CKD patients (n=29; aged  $67.03 \pm 2.64$ ; BMI  $24.53 \pm 1.01$ ); finally, 11 normal subjects (Group C) have been enrolled as controls. Irisin has been evaluated by ELISA method and Total Antioxidant Capacity (TAC) by spectrophotometric method.

**RESULTS:** Irisin was significantly higher in Group B vs. A and C groups (Mean  $\pm$  SEM: 20.18  $\pm$  0.61 ng/ml vs. 2.77  $\pm$  0.77 and 13.06  $\pm$  0.56, respectively; p<0.05); a significant correlation between irisin and TAC was observed in group B.

**CONCLUSIONS:** These preliminary data suggest a possible role of irisin in the modulation of

antioxidants in two chronic syndromes with low T3 (i.e., CHF and CKD) with differential pattern in these two models studied. Further insights are needed to confirm this pilot study, which could be the basis for a longitudinal investigation, to assess a prognostic role of irisin with possible therapeutic implications.

*Key Words:*NTIS, Thyroid, Heart failure, CKD.

### Introduction

Non-thyroidal-illness syndrome (NTIS) is a functional disorder present in acute and chronic sickness, in absence of thyroid disease, and considered an adaptive response rather than a real hypothyroidism<sup>1</sup>. The most common NTIS patient's condition is due to a low deiodination of fT4, leading to reduced circulating levels of fT3 ("low-fT3 syndrome"), that progressively involve also fT4 with low secretion<sup>2</sup>. In acute situations, this can be observed after starvation and critical illness, such as sepsis or major surgery, while low fT3 is commonly observed in chronic kidney and liver diseases, heart failure, and chronic inflammatory diseases<sup>3</sup>. A key role seems to be exerted by increased cytokines<sup>4</sup>, but other mechanisms can be also involved, such as alterations in thyroid hormones transport, clearance and modifications of membrane transporters<sup>2</sup>. Also, selenium element is involved, since it converts thyroids hormones through a mechanism of deiodination by selenoproteins<sup>5</sup>. Although debated, thyroid

<sup>&</sup>lt;sup>1</sup>Dipartimento di Medicina e Chirurgia Traslazionale, Università Cattolica del Sacro Cuore, Rome, Italy <sup>2</sup>Dipartimento di Scienze Cardiovascolari, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy

<sup>&</sup>lt;sup>3</sup>Dipartimento di Scienze Mediche e Chirurgiche, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy

<sup>&</sup>lt;sup>4</sup>Dipartimento di Scienze Biotecnologiche di Base, Cliniche Intensivologiche e Perioperatorie, Università Cattolica del Sacro Cuore, Rome, Italy

hormone replacement therapy is not usually required<sup>6,7</sup>. Moreover, at cellular levels, a reduced T3 bioavailability can be unsafe and express a "maladaptive" rather than an "adaptive" response. As consequences, intracellular oxidative stress has been hypothesized<sup>8,9</sup> to exert an active role. In fact, oxidative stress (OS) could exacerbate NTIS condition in a vicious circle, due to deiodinases alteration and negative effects of low T, on antioxidant levels or activity<sup>10</sup>. Thus, chronic inflammation and OS exert reciprocal influences leading to a worse clinical progression of such conditions<sup>11</sup>. Accordingly, based on experimental and clinical studies<sup>12</sup>, thyroid hormones (TH) levels in NTIS do not necessarily reflect its serum low concentrations and there is a tissue-specific thyroid hormone transport, receptor binding and hormone metabolism. Nevertheless, muscle is one of the main targets of TH and it is not surprising that many alterations are observed in NTIS condition. For instance, in mice, TH expression is decreased in sepsis and acute inflammation, but it seems not to be affected in chronic inflammation. The same scenario is observed in humans since the expression of monocarboxylate transporter 8 (MCT8) is lower after acute surgical stress than in prolonged illness13, however, a compensatory increase in MCT8 has been described in chronic diseases in rabbits<sup>13</sup>. Different regulations of deiodinases 2 and 3 (DIO2 and DIO3) modulate T3 intracellular levels with a subtle tuning, to compensate the systemic TH unbalance<sup>14</sup>.

Irisin is a 112 amino acid protein of 12 kDa molecular weight, identified for its ability to induce browning of white adipose tissue, protect from insulin resistance, and increase energy expenditure<sup>15</sup>. In human being, irisin is mainly expressed in skeletal and heart muscle<sup>16</sup>. Irisin is produced by a proteolytic cleavage of fibronectin type III domain containing protein 5 (FNDC5) a transmembrane protein whose expression is induced by exercise and/or by increasing peroxisome proliferator-activated receptor (PPAR)-y co-activator  $1\alpha$  (PGC- $1\alpha$ ). Interestingly, irisin has been confirmed to carry out a thermogenic role through a pathway involving PGC-1a. Moreover, it is proposed that PGC-1α activation and/or oxidative stress<sup>17</sup> controls the expression FNDC5, which, when released in the blood, produces irisin peptide<sup>15</sup>.

Furthermore, irisin is highly expressed in myocardium, although its physiological effects in the heart remain not fully known and controversial<sup>18</sup>. While the precise physiological role of irisin in human being and its correlations with diseases remain unclear, some scholars<sup>19</sup> show that there is an association between irisin levels and comorbidities (e.g., type 2 diabetes mellitus and metabolic syndrome).

Moreover, obesity, T2DM and the metabolic syndrome have been strictly associated with cardiovascular disease (CVD), including coronary artery disease (CAD) and acute coronary syndromes (ACSs). Some studies<sup>19,20</sup> have confirmed that circulating irisin levels are positively associated of the 10-year CVD risk, compared with the general Framingham risk profile<sup>19</sup> and diminished expression of FNDC5 could be related with reduced aerobic performance in patients with CVD<sup>20</sup>.

Finally, a different model of NTIS is present in chronic kidney disease (CKD)<sup>21</sup>; consequently, low fT3 levels have been shown to be independent predictor of mortality in haemodialysis (HD) patients<sup>22</sup>.

Therefore, this pilot study was designed to evaluate, in a case-control study, as primary endpoint the circulating irisin levels in two models of NTIS, such as chronic heart failure (CHF) and chronic kidney disease (CKD) during haemodialytic treatment. The secondary endpoint was the evaluation of total antioxidant capacity (TAC) and its correlation with irisin in order to establish a possible role of this myokine in the modulation of antioxidant systems.

## **Patients and Methods**

Three groups of patients were selected to participate in this study and were enrolled after being explained the purposes and nature of the study, conducted in accordance with the declaration of Helsinki, as revised in 2013. The study protocol was approved by our Centre's Ethics Committee (School of Medicine, Catholic University, Rome, Italy) and written informed consent was obtained from all patients: patients affected by chronic heart failure (CHF, group A), chronic kidney disease treated by haemodialysis (HD, group B), and control group (group C).

All subjects involved in this study were admitted to the University Hospital "Policlinico Gemelli" Dept. of Internal Medicine. Two senior cardiologists separately confirmed the diagnosis of CHF (group A) based on clinical history, physical examination, laboratory and echocardiographic parameters, according to the European

Society of Cardiology Guidelines for the Management of Heart Failure<sup>23</sup>. Group A: 18 patients with Heart Failure with reduced ejection function (HFrEF) (15 males), aged 42-88 years (mean 69.2) were recruited. All of them were Caucasian; they were treated by conventional therapy according to ESC guidelines (beta-blockers 1, ACE-inhibitors 7, angiotensin receptor blockade 7, Diuretics 17). Comorbidities were present in HFrEF (33% T2DM, 39% hypertension, 44% atrial fibrillation, 6% peripheral atherosclerosis, 33% nonend stage chronic kidney disease, 16% COPD). All patients were classified according to NYHA classification (all belonged to class II or III) and levels of physical activity (which was confined to sedentary activity) were reported. Participants were excluded if they had uncontrolled hypertension (blood pressure > 140 mmHg/90 mmHg), alcoholism, drug abuse, abnormal hepatic function (transaminases > twice the upper limit of normal), end stage renal disease, malabsorption syndromes, gastro-esophageal reflux disease. Group B: 29 patients (16 males) affected by end-stage renal failure who received three times weekly hemodialysis for a period ranging from 8-336 months at the Hemodialysis Unit of the Catholic University, Rome, Italy, were screened for inclusion in the present study. Exclusion criteria were as follows: advanced heart failure (according to the criteria of the European Society of Cardiology), diagnosis of dementia based on DSM-IV criteria, history of alcohol or substance abuse, previous diagnosis of psychotic disorders, clinical instability requiring hospital admission. All patients received 4-h bicarbonate hemodialvsis three times a week, according to the schedule employed in our Hospital<sup>24</sup>. The blood flow ranged from 250 to 300 mL/min, with a dialysis rate flow of 500 mL/min. All patients of Group B were treated with high-permeability membranes. Membranes were not reused. Comorbidity was quantified using the Charlson comorbidity score index<sup>25</sup>. Diagnoses were collected according to the International Classification of Diseases, ninth edition, and Clinical Modification codes (International Classification of Diseases (ICD). www. who.int/ classifications/icd/en/). All drugs assumed by participants were coded according to the Anatomical, Therapeutic, and Chemical codes. (ATC/DDD Index 2013. www.whocc.no/ atc ddd index/). Interdialytic weight gain (ID-WG) and pre-dialysis systolic blood pressure of 10 consecutive hemodialysis sessions (the same used to record IDH) were recorded and mean and

median values were calculated  $^{26}$ . Finally, a group of normal subjects (Group C), aged  $50.4\pm3.42$  ys, with BMI  $25.07\pm2.11$  kg/m<sup>2</sup> was included as control group.

Between 08.00 and 09.00 a.m., at fasting, a venous withdraw was performed; the blood was collected in a test tube containing heparin as anticoagulant and immediately centrifuged (4°C at 2500× g for 10 min). Samples were subsequently aliquoted in 2 ml and stored at -80°C until assayed. In HD patients, the blood sample was collected during the long interval (72 h after the last dialysis, in order to minimize the effect of acute heparin administration) through the arteriovenous fistula or the central venous catheter immediately before their scheduled hemodialysis session at the beginning of the week.

Fasting glucose, total and HDL cholesterol, triglycerides, uric acid, transaminases and albumin levels were quantified using ADVIA 2400 automatic analyzer (Siemens, Italy). Plasma concentrations of NT-proBNP were measured by immunochemiluminometric assays on a Roche modular E170 analyzer (Roche diagnostic; Indianapolis, IN, USA). Plasma fT3, fT4, TSH were assayed by ECLIA. Lower T3 syndrome was defined as the presence of fT3 values under the median for each group (2.7 ng/ml in CHF and 2.3 ng/ml in CKD).

Total antioxidant capacity (TAC) was estimated with the method of Rice-Evans<sup>27</sup>, as modified in our laboratory<sup>28</sup>. TAC determination is based on the interaction between the system H<sub>2</sub>O<sub>2</sub>-methmyoglobin with the chromogen ABTS, which radical form is spectrophotometrically detectable (at 740 nm). The latency time (LAG phase) expressed in second before the appearance of radical species is proportional to the antioxidant concentration in the sample.

An enzyme immunoassay kit (Cat. No. EK-067-029 from Phoenix Pharmaceuticals, Karlsruhe, Germany) was used for evaluation of circulating irisin concentrations according with the manufacturer's protocol. The intra- and inter-assay variations were less than 10% and 15%, respectively; detection limit ranges from 0.1 to 1000 ng/ml. The ELISA kit used for detection of irisin was previously validated by MS<sup>29</sup>. Optical density at 450 nm was measured, with a reading time of 1 sec., using a microtiter plate reader (Victor3; Perkin Elmer, Waltham, MA, USA) with precision at 450 nm < 0.5% and temperature control set at 25°C. Analyses were performed in duplicate.

**Table I.** Basal and anthropometric features in patients studied.

	Age (ys)	Sex (M/F)	BMI (Kg/m²)	DM (%)	Hypertension (%)
CHF	$70.22 \pm 2.78$	15/3	$27.75 \pm 1.28$	46.2	46.7
HD	$67.03 \pm 2.64$	16/13	$24.53 \pm 1.01$	35	84.2

A complete echocardiographic evaluation was performed in CHF patients (Echocardiography Philips, Affiniti 70c), measuring the following parameters: left ventricular ejection fraction (LVEF), left ventricular end-diastolic diameter (LVEDD), left ventricular end-systolic diameter (LVESD), left ventricular end-diastolic volume (LVEDV), left ventricular end-systolic volume (LVESV), Septal thickness (IVS), Posterior wall thickness (PW), Peak E-wave velocity (E), Peak A-wave velocity (A), E/A ratio, Pulsed-wave TDI E' velocity (E'), E/E' ratio, Deceleration time (DT), left atrial volume (LAV), indexed atrial volume (LAVI), systolic pulmonary artery pressure (SPAP), tricuspid annular plane systolic excursion (TAPSE) and tricuspid peak velocity (TPV).

#### Statistical Analysis

The Mann-Whitney U test was employed to evaluate differences between the two groups of subjects. A *p*-value of 0.05 was considered statistically significant. Chi-square test was employed to evaluate categorical variables (presence or not of Low fT3) and parts of whole. Linear regression analysis was employed to correlate irisin with the other studied parameters.

## Results

Clinical characteristics and comorbidities of subjects affected by chronic heart failure (CHF,

**Table II.** Metabolic parameters in patients studied.

	CHF	HD
Glycemia (mg/dl)	$80.2 \pm 5.69$	$86.43 \pm 3.24$
Cholesterol (mg/dl)	$153.62 \pm 10.91$ $40.01 \pm 3.68$	$148.22 \pm 6.06$ $41.45 \pm 2.12$
HDL (mg/dl) Triglycerides (mg/dl)	$95.56 \pm 9.14$	$160.23 \pm 17.15$
Uric acid (mg/dl)	$5.81 \pm 0.3$	$6.14 \pm 0.2$
Creatinine (mg/dl)	$1.33 \pm 0.21$	$9.48 \pm 0.35$
AST (U/l)	$20.25 \pm 1.61$	$11.78 \pm 0.75$
ALT (U/l) Albumin (g/l)	$17.54 \pm 2.28$ $39 \pm 1.84$	$10.85 \pm 1.03$ $19.05 \pm 4.77$
1110 uniiii (g, 1)	57 - 1.0 .	19.00 - 1.77

group A) and chronic kidney diseases treated by hemodialysis (HD, group B) are reported in Table I.

Mean (±SEM) values of hematochemical and thyroid hormones levels are reported in Table II and III, respectively. The prevalence of low T3 was similar in the two groups (Group A 43.75%; Group B 37.93%; n.s. using Chi-square test) as shown in Figure 1. Figure 2 shows the mean (±SEM) of plasmatic TAC values, which were not different between groups. Figure 3 shows the mean (±SEM) levels of irisin. Significantly higher levels of irisin were observed in HD patients (group B) vs. CHF (group A) and controls (group C); CHF also showed lower levels vs. controls.

When considering correlation studies, using irisin as independent variable, no significant correlation was showed between irisin and fT3/fT4/BMI/TAC; on the contrary, a significant positive correlation between irisin and LAG was present in patients with HD (shown in Figure 4).

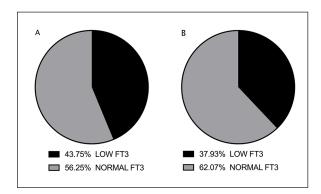
#### Discussion

At the best of our knowledge, this is the first paper that reports a correlation between irisin and LAG in HD patients. The same correlation was not observed in CHF patients, in which a very low level of irisin were revealed.

The interest concerning irisin, in CHF condition, started from the experiments in zebrafish, in which it increases diastolic volume, heart frequency and cardiac output<sup>30</sup>. Interesting results were reported in murine models with diabetic cardiomyopathy: low dose of recombinant irisin treatment diminished cardiac fibrosis and ame-

**Table III.** Thyroid function parameters in patients studied.

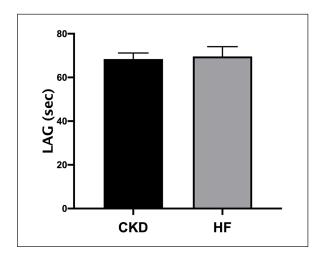
	TSH	fT3	fT4
	mcUI/ml	pg/ml	pg/ml
CHF HD	$2.5 \pm 0.8$ $2.01 \pm 0.28$	$2.7 \pm 0.15$ $2.29 \pm 0.08$	$11.82 \pm 0.57$ $9.26 \pm 0.27$



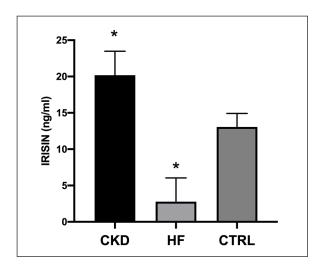
**Figure 1.** Prevalence of low T3 syndrome in the two groups of chronic diseases enrolled in the study (A= chronic heart failure with reduced ejection fraction; B = chronic kidney disease under haemodialytic treatment).

liorates left ventricular function; however, in higher doses the same effects were not observed; on the contrary, it increased collagen deposition, showing a dose-related effect. A possible explanation is an increased expression of matrix metalloproteinases *via* MAPK and proliferation of fibroblasts at myocardial level<sup>31</sup>. Accordingly, in murine model of chronic heart failure with reduced ejection fraction, the expression in skeletal muscle of FNDC5 and PGC-1a is depressed, with a consequent decrease of serum irisin levels<sup>32</sup>.

Interestingly, the incubation of murine skeletal muscle cells with TNF-a and IL1B, or both, further decreased the expression of the same pathway suggesting an interference of chronic inflammation and endocrine function of muscle<sup>32</sup>.



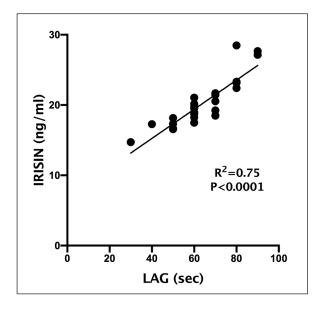
**Figure 2.** Mean  $\pm$  SEM levels of LAG phase (sec) as measure of total antioxidant capacity in the two groups of chronic illness.



**Figure 3.** Mean  $\pm$  SEM of circulating irisin levels in the two groups of chronic illness (CKD and HF) and control subjects (CTRL). \*p<0.05.

Despite the correlation between inflammation and OS, no data were reported about the correlation between irisin and antioxidants.

Recently, we demonstrated however different pattern of irisin and antioxidants in two kinds of CHF, with preserved or reduced ejection faction. In the first, in fact, we observed higher levels than in the latter<sup>33</sup>; among the possible explanations, an increased oxidative stress, and a compensatory increase of irisin, was hypothesized<sup>34</sup>.



**Figure 4.** Correlation between circulating irisin levels and LAG in the CKD group of patients.

Accordingly, in the present work, we confirmed lower irisin levels in CHF with reduced ejection fraction (group B) compared with the other two groups.

In HD patients (group A), the situation is more complex. CKD is a condition with increased OS through different mechanisms<sup>35</sup>, including lipotoxicity (both induced by reduced b-oxidation of free fatty acids, and lipid absorption by the CD36 receptor), increased mitochondrial ROS production and deregulation of mitophagy<sup>36,37</sup>. Furthermore, HD induces a chronic inflammation through the activation of polymorphonuclear cells and monocytes, and the consequent stimulation of myeloperoxidases and NADPH<sup>38</sup>.

In the present study, we describe, in disagreement with data reported by other authors, higher levels of irisin in CKD than in controls and CHF patients. Surprisingly, results reported elsewhere in literature are still conflicting<sup>39-43</sup>. In patients with CKD irisin was associated with fat mass, BMI and glomerular filtration rate (GFR), with the lowest levels observed in the 5th stage of CKD<sup>39</sup>. Adult obese Chinese patients with higher irisin levels showed reduced prevalence of CKD<sup>40</sup>. A correlation with GFR was reported in some studies<sup>41,42</sup> but not with microalbuminuria<sup>42</sup>. Another article<sup>43</sup>, confirming lower irisin levels in CKD, however, found that in peritoneal dialysis, irisin concentrations were higher than in HD. A recent meta-analysis<sup>44</sup>, focused on irisin levels in CKD patients, confirmed that this peptide was decreased in CKD patients, mainly in dialysis patients.

To be clear, some limits could explain these contrasting results:

1) The normal range of circulating irisin in CKD is not known; conflicting data are reported about irisin levels in CKD<sup>41</sup>. Meta-analysis<sup>44</sup> showed that eight studies reported significantly different irisin levels in CKD vs. control, while other studies did not find this correlation. The CKD population was heterogeneous, including non-dialysis and patients treated with hemodialysis or peritoneal dialysis with differences that have been reported in these subgroups<sup>44</sup>. It has been also shown, in in vivo animal models, using irisin radiolabeled with 125I and SPECT/ TC imaging, that the metabolic clearance of irisin was related to liver and kidney, with an integrated action<sup>45</sup>. However, the specific contribution of these two systems could be modified in pathological states.

- 2) Several studies are observational and performed in a small sample, therefore a causal relationship between CKD and irisin is not still fully demonstrated. Moreover, the etiology of CKD could also be decisive. For instance, the study of Liu et al<sup>46</sup>, which described the relationship between GFR and irisin, included patients with type 2 diabetes. It is known that diabetes itself can show irisin modifications which is important for the pathogenesis and clinical course of the disease<sup>45</sup>.
- 3) Ethnic interferences can be important; a meta-analysis<sup>47</sup> showed higher levels of irisin in non-Asian than in Asian population. Accordingly, stratifying subgroups by geographic region, irisin concentrations was different, even if underlying mechanisms were not clarified.
- 4) The impact of the level of physical activity is not considered with irisin in most studies, although exercise is well known to affect irisin levels; different kind of effects are exerted by acute or chronic activity<sup>15,48</sup> and by resistance *vs.* endurance effort<sup>49</sup>. We have not measured the level of activity in our patients; however, it has been shown that neither a resistance training program<sup>50</sup> nor acute intradialytic strength exercise<sup>51</sup> were able to induce irisin variations in patients under hemodialytic treatment. In our cohort, the lack of a positive factor, such as exercise, was surely more determinant in chronic heart failure.
- 5) There is still considerable heterogeneity in reports on circulating levels of irisin in humans. Moreover, ELISA methods show significant variations depending on the manufacturer kit employed.
  - In fact, irisin levels measured with different commercial ELISAs were reported in a range from picograms to micrograms per milliliter of plasma. Thus, measuring circulating irisin remains challenging.
- 6) Last but not least, the impact of malnutrition and sarcopenia on irisin levels is not always considered. Irisin, which originates from muscle, is evidently related to sarcopenia, even if this topic has been investigated especially in relation to cardiovascular risk factor<sup>52</sup>. Again, our cohort was consisting of normal weight subjects, therefore, the observed irisin values should not have been influenced by this factor.

Moreover, in our cohort, with normal weight patients, the higher irisin levels observed could be furthermore explained by its protective effect on oxidative stress; this hypothesis is reinforced by the correlation with LAG and by the similarity with another condition characterized by increased OS as HFpEF<sup>33</sup>.

The role of irisin and its antioxidant role is described in experimental animal models. Liu et al<sup>53</sup> demonstrated a protective effect of irisin on the damage produced in renal ischemia-reperfusion by suppressing p53. Other experiments supported this hypothesis: irisin treatment ameliorates OS induced by H<sub>2</sub>O<sub>2</sub> in murine hepatocytes<sup>54</sup> and OS induced by palmitic acid in hepatocytes of mice and humans<sup>55</sup>. Other studies<sup>56</sup> confirmed similar result, demonstrating the antioxidant properties of irisin in a mice model of T2DM and in human cells of diabetic patients. Accordingly, irisin exerts a protective effect on endothelial function in the same models<sup>57,58</sup>.

The lack of difference of LAG between HD and CHF patients is not contrasting with our hypothesis, since many factors can influence this parameter, such as the levels of uric acid, which is increased in HD, but very likely is the main component of LAG assay.

In spite of all precautions, our study may be still subject to certain biases, and some main potential restrictions should be considered, other than those above reported. The number of subjects in the groups is slightly small, so the statistical power of the study is limited; consequently, our findings will need to be confirmed in a larger population and still require further studies. Moreover, the moment of the disease history in which the patient is evaluated may affect irisin and antioxidants levels. Therefore, the study design and the power analysis cannot draw a cause-effect relationship.

#### Conclusions

These preliminary data suggest a possible role of irisin in modulation of antioxidants in two chronic syndromes with low T3 with differential pattern in these two models studied (i.e., CHF and HD). Further insights are needed to confirm this pilot study, which could be the basis for a longitudinal investigation, to assess a prognostic role of irisin evaluation with possible therapeutic implications.

#### **Conflict of Interest**

The Authors declare that they have no conflict of interests.

#### Acknowledgements

None.

#### **Informed Consent**

Written informed consent was obtained from all patients.

#### **Ethics Statement**

The study protocol was approved by our Centre's Ethics Committee (School of Medicine, Catholic University, Rome, Italy). The study was conducted according to the Declaration of Helsinki.

#### **Authors' Contribution**

Conceptualization: AM, AS; Data collection: CB, EV, EC, AMN; Statistical Analysis: CB, EV; Writing: AM, EC, CB, EV; Supervision: AM, AS, EM, AM, AMRF, NP; Experimental procedures: AS, EM, AM.

## **Funding**

The study was supported by "Fondi di Ateneo Linea D1" to AS and AM (n. R4124500893/2019).

#### **ORCID ID**

Mancini A 0000-0002-9417-6810; Bruno C 0000-0002-3489-0238; Vergani E 0000-0002-8444-0091; Silvestrini A 0000-0002-2005-3746.

# References

- Chopra IJ. Clinical review 86: Euthyroid sick syndrome: is it a misnomer? J Clin Endocrinol Metab 1997; 82: 329-334.
- De Groot LJ. Non-thyroidal illness syndrome is a manifestation of hypothalamic-pituitary dysfunction, and in view of current evidence, should be treated with appropriate replacement therapies. Crit Care Clin 2006; 22: 57-86.
- 3) McIver B, Gorman CA. Euthyroid sick syndrome: an overview. Thyroid 1997; 7: 125-132.
- Bartalena L, Brogioni S, Grasso L, Velluzzi F, Martino E. Relationship of the increased serum interleukin-6 concentration to changes of thyroid function in nonthyroidal illness. J Endocrinol Invest 1994; 17: 269-274.
- Silvestrini A, Mordente A, Martino G, Bruno C, Vergani E, Meucci E, Mancini A. The Role of Selenium in Oxidative Stress and in Nonthyroidal Illness Syndrome (NTIS): An Overview. Curr Med Chem 2020; 27: 423-449.
- Chopra IJ, Huang TS, Boado R, Solomon DH, Teco GNC. Evidence against benefit from replacement doses of thyroid hormones in nonthyroidal

- illness (NTI): studies using turpentine oil-injected rat. J Endocrinol Invest 1987; 10: 559-564.
- Wartofsky L, Burman KD. Alterations in thyroid function in patients with systemic illness: the "euthyroid sick syndrome." Endocr Rev 1982; 3: 164-217.
- Mancini A, Corbo GM, Gaballo A, Raimondo S, Di Segni C, Gigliotti P, Silvestrini A, Valente S, Littarru GP, Pontecorvi A, Meucci E. Relationship between plasma antioxidants and thyroid hormones in chronic obstructive pulmonary disease. Exp Clin Endocrinol Diabetes 2012; 120: 623-628.
- Mancini A, Raimondo S, Di Segni C, Persano M, Gadotti G, Silvestrini A, Festa R, Tiano L, Pontecorvi A, Meucci E. Thyroid hormones and antioxidant systems: focus on oxidative stress in cardiovascular and pulmonary diseases. Int J Mol Sci. 2013 14: 23893-23909.
- St Germain DL, Galton VA, Hernandez A. Minireview: Defining the roles of the iodothyronine deiodinases: current concepts and challenges. Endocrinology 2009; 150: 1097-1107.
- Mancini A, Di Segni C, Raimondo S, Olivieri G, Silvestrini A, Meucci E, Currò D. Thyroid Hormones, Oxidative Stress, and Inflammation. Mediators Inflamm. 2016; 2016: 6757154.
- Fliers E, Boelen A. An update on non-thyroidal illness syndrome. J Endocrinol Invest 2021; 44: 1597-1607.
- Mebis L, Paletta D, Debaveye Y, Ellger B, Langouche L, D'Hoore A, Darras VM, Visser TJ, Van den Berghe G. Expression of thyroid hormone transporters during critical illness. Eur J Endocrinol 2009; 161: 243-250.
- 14) Arrojo E, Drigo R, Bianco AC. Type 2 deiodinase at the crossroads of thyroid hormone action. Int J Biochem Cell Biol 2011; 43: 1432-1441.
- 15) Bostrom P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, Rasbach KA, Boström EA, Choi JH, Long JZ, Kajimura S, Zingaretti MC, Vind BF, Tu H, Cinti S, Højlund K, Gygi SP, Spiegelman BM. A PGC1 alpha dependent myokine that drives brown fat like development of white fat and thermogenesis. Nature 2012; 481: 463-468
- 16) Huh JY, Panagiotou G, Mougios V, Brinkoetter M, Vamvini MT, Schneider BE, Mantzoros CS. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise. Metabolism. 2012 61: 1725-1738.
- 17) Berthold HK, Rizzo M, Spenrath N, Montalto G, Krone W, Gouni-Berthold I. Effects of lipid-lowering drugs on high-density lipoprotein subclasses in healthy men-a randomized trial. PLoS One 2014; 9: e91565.
- 18) Aronis KN, Moreno M, Polyzos SA, Moreno-Navarrete JM, Ricart W, Delgado E, de la Hera J, Sahin-Efe A, Chamberland JP, Berman R, Spiro A 3rd, Vokonas P, Fernández-Real JM, Mantzoros CS. Circulating irisin levels and coronary

- heart disease: Association with future acute coronary syndrome and major adverse cardiovascular events. Int J Obes. 2015 39: 156-161.
- 19) Park KH, Zaichenko L, Brinkoetter M, Thakkar B, Sahin-Efe A, Joung KE, Tsoukas MA, Geladari EV, Huh JY, Dincer F, Davis CR, Crowell JA, Mantzoros CS. Circulating irisin in relation to insulin resistance and the metabolic syndrome. J Clin Endocrinol Metab. 2013 98: 4899-4907
- 20) Lecker SH, Zavin A, Cao P, Thakkar B, Sahin-Efe A, Joung KE, Tsoukas MA, Geladari EV, Huh JY, Dincer F, Davis CR, Crowell JA, Mantzoros CS. Expression of the irisin precursor fndc5 in skeletal muscle correlates with aerobic exercise performance in patients with heart failure. Circ Hear Fail. 2012; 5: 812-818.
- Mohamedali M, Reddy Maddika S, Vyas A, Iyer V, Cheriyath P. Thyroid disorders and chronic kidney disease. Int J Nephrol 2014; 2014: 520281.
- 22) Zoccali C. Asymmetric dimethylarginine in end-stage renal disease patients: a biomarker modifiable by calcium blockade and angiotensin II antagonism? Kidney Int 2006; 70: 2053-2055.
- 23) Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JG, Coats AJ, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GM, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. Eur Heart J 2016; 37: 2129-2200.
- 24) Panocchia N, Tonnara G, Minacori R, Sacchini D, Bossola M, Tazza L, Gambaro G, Spagnolo AG. Survey on advance care planning of Italian outpatients on chronic haemodialysis. BMJ Support Palliat Care 2017; 7: 419-422.
- 25) Di Iorio B, Cillo N, Cirillo M, Gaspare De Santo N. Charlson. Comorbidity Index is a predictor of outcomes in incident hemodialysis patients and correlates with phase angle and hospitalization. Int J Artif Organs 2004; 27: 330-336.
- 26) Bossola M, Laudisio A, Antocicco M, Panocchia N, Tazza L, Colloca G, Tosato M, Zuccalà G. Intradialytic hypotension is associated with dialytic age in patients on chronic hemodialysis. Ren Fail 2013 35: 1260-1263.
- Rice-Evans C, Miller NJ. Total antioxidant status in plasma and body fluids. Methods Enzymol 1994; 234: 279-293.
- 28) Mancini A, Leone E, Festa R, Grande G, Di Donna V, De Marinis L, Pontecorvi A, Tacchino RM, Littarru GP, Silvestrini A, Meucci E. Evaluation of antioxidant systems (coenzyme Q10 and total antioxidant capacity) in morbid obesity before and after biliopancreatic diversion. Metabolism 2008; 57: 1384-1389.
- 29) Polyzos SA, Mantzoros CS. An update on the validity of irisin assays and the link between irisin

- and hepatic metabolism. Metabolism 2015; 64: 937-942.
- Sundarrajan L, Yeung C, Hahn L, Weber LP, Unniappan S. Irisin regulates cardiac physiology in zebrafish. PLoS One 2017; 12: e0181461.
- 31) Liu X, Mujahid H, Rong B, Lu QH, Zhang W, Li P, Li N, Liang ES, Wang Q, Tang DQ, Li NL, Ji XP, Chen YG, Zhao YX, Zhang MX. Irisin inhibits high glucose-induced endothelial-to-mesenchymal transition and exerts a dose-dependent bidirectional effect on diabetic cardiomyopathy. J Cell Mol Med 2018; 22: 808-822.
- 32) Matsuo Y, Gleitsmann K, Mangner N, Werner S, Fischer T, Bowen TS, Kricke A, Matsumoto Y, Kurabayashi M, Schuler G, Linke A, Adams V. Fibronectin type III domain containing 5 expression in skeletal muscle in chronic heart failure-relevance of inflammatory cytokines. J Cachexia Sarcopenia Muscle 2015; 6: 62-72.
- 33) Silvestrini A, Bruno C, Vergani E, Venuti A, Favuzzi AMR, Guidi F, Nicolotti N, Meucci E, Mordente A, Mancini A. Circulating irisin levels in heart failure with preserved or reduced ejection fraction: A pilot study. PLoS One 2019; 14(1): e0210320.
- 34) Mancini A, Vergani E, Bruno C, Olivieri G, Di Segni C, Silvestrini A, Venuti A, Favuzzi A, Meucci E. Oxidative stress as a possible mechanism underlying multi-hormonal deficiency in chronic heart failure. Eur Rev Med Pharmacol Sci 2018; 22: 3936-3961.
- 35) Aranda-Rivera AK, Cruz-Gregorio A, Aparicio-Trejo OE, Pedraza-Chaverri J. Mitochondrial Redox Signaling and Oxidative Stress in Kidney Diseases. Biomolecules 2021; 11: 1144.
- Stadler K, Goldberg IJ, Susztak K. The evolving understanding of the contribution of lipid metabolism to diabetic kidney disease. Curr Diab Rep 2015; 15: 40.
- 37) Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007; 39: 44-84.
- 38) Piepoli MF, Kaczmarek A, Francis DP, Davies LC, Rauchhaus M, Jankowska EA, Anker SD, Capucci A, Banasiak W, Ponikowski P. Reduced peripheral skeletal muscle mass and abnormal reflex physiology in chronic heart failure. Circulation 2006; 114: 126-134.
- 39) Liu JJ, Wong MDS, Toy WC, Liu JJ, Toy WC, Wong MD, Tan CS, Tavintharan S, Wong MS, Sum CF, Lim SC. Elevated undercarboxylated and reduced carboxylated osteocalcin are associated with metabolic syndrome in middle age Asian females. Lower circulating irisin is associated with type 2 diabetes mellitus. J Diabetes Complications 2013; 27: 365-369.
- 40) Yang S, Xiao F, Pan L, Zhang H, Ma Z, Liu S, Liu Y, Zhang W, Zeng X, Liu C, Li X, Li X, Li Z. Association of serum irisin and body composition with

- chronic kidney disease in obese Chinese adults: a cross-sectional study. BMC Nephrol 2015; 16:
- Maciorkowska M, Musiałowska D, Małyszko J. Adropin and irisin in arterial hypertension, diabetes mellitus and chronic kidney disease. Adv Clin Exp Med 2019; 28: 1571-1575.
- 42) Liu JJ, Liu S, Wong MD, Tan CS, Tavintharan S, Sum CF, Lim SC. Relationship between circulating irisin, renal function and body composition in type 2 diabetes. J Diabetes Complications 2014; 28: 208-213.
- 43) Rodríguez-Carmona A, Pérez Fontán M, Sangiao Alvarellos S, García Falcón T, Pena Bello ML, López Muñiz A, Cordido F. Serum levels of the adipomyokine irisin in patients with chronic kidney disease. Nefrologia 2016; 36: 496-502.
- 44) Gan W, Chen W, Li T, Shao D, Xu F, Huo S, Li C, Yang Z, Zeng X. Circulating irisin level in chronic kidney disease patients: a systematic review and meta-analysis. Int Urol Nephrol 2021; 54: 1295-1302.
- 45) Lv J, Pan Y, Li X, Cheng D, Ju H, Tian J, Shi H, Zhang Y. Study on the distribution and elimination of the new hormone irisin in vivo: new discoveries regarding irisin. Horm Metab Res 2015; 47: 591-595.
- 46) Liu JJ, Liu S, Wong MDS, Liu JJ, Liu S, Wong MD, Tan CS, Tavintharan S, Sum CF, Lim SC. Relationship between circulating irisin, renal function and body composition in type 2 diabetes. Relationship between circulating irisin, renal function and body composition in type 2 diabetes. J Diabetes Complications 2014; 28: 208-213.
- 47) Qiu S, Cai X, Yin H, Zügel M, Sun Z, Steinacker JM, Schumann U. Association between circulating irisin and insulin resistance in non-diabetic adults: A meta-analysis. Metabolism 2016; 65: 825-834.
- 48) Löffler D, Müller U, Scheuermann K, Friebe D, Gesing J, Bielitz J, Erbs S, Landgraf K, Wagner IV, Kiess W, Körner A. Serum irisin levels are regulated by acute strenuous exercise. J Clin Endocrinol Metab 2015; 100: 1289-1299.
- Tsuchiya Y, Ando D, Takamatsu K, Goto K. Resistance exercise induces a greater irisin response than endurance exercise. Metabolism 2015; 64: 1042-1050.
- 50) Moraes C, Leal VO, Marinho SM, Barroso SG, Rocha GS, Boaventura GT, Mafra D. Resistance exercise training does not affect plasma irisin levels of hemodialysis patients. Horm Metab Res 2013; 45: 900-904.
- 51) Esgalhado MGBM, Stockler-Pinto MB, Cardozo LFM de F, Barboza JE, Mafra D. Does high intensity exercise affects irisin plasma levels in hemodialysis patients? A pilot study. J Bras Nefrol. 2018; 40: 53-58.
- 52) Barbalho SM, Flato UAP, Tofano RJ, Goulart RA, Guiguer EL, Detregiachi CRP, Buchaim DV, Araú-

- jo AC, Buchaim RL, Reina FTR, Biteli P, Reina DO-BR, Bechara MD. Physical Exercise and Myokines: Relationships with Sarcopenia and Cardiovascular Complications. Int J Mol Sci 2020; 21: 3607.
- 53) Liu Y, Fu Y, Liu Z, Shu S, Wang Y, Cai J, Tang C, Dong Z. Irisin is induced in renal ischemia-reperfusion to protect against tubular cell injury via suppressing p53. Biochim Biophys Acta Mol Basis Dis 2020; 1866: 165792.
- 54) Batirel S, Bozaykut P, Mutlu Altundag E, Kartal Ozer N, Mantzoros CS. The effect of Irisin on antioxidant system in liver. Free Radic Biol Med 2014; 75: S16.
- 55) Park MJ, Kim D II, Choi JH, Heo YR, Park SH. New role of irisin in hepatocytes: The protective effect of hepatic steatosis in vitro. Cell Signal 2015; 27: 1831-1839.
- 56) Zhu D, Wang H, Zhang J, Zhang X, Xin C, Zhang F, Lee Y, Zhang L, Lian K, Yan W, Ma X, Liu Y, Tao L. Irisin improves endothelial function in type 2 diabetes through reducing oxidative/nitrative stresses. J Mol Cell Cardiol 2015; 87: 138-147.
- Moncada S, Higgs EA. The discovery of nitric oxide and its role in vascular biology. Br J Pharmacol 2006; S193-S201.
- 58) Symons JD, McMillin SL, Riehle C, Tanner J, Palionyte M, Hillas E, Jones D, Cooksey RC, Birnbaum MJ, McClain DA, Zhang QJ, Gale D, Wilson LJ, Abel ED. Contribution of insulin and Akt1 signaling to endothelial nitric oxide synthase in the regulation of endothelial function and blood pressure. Circ Res 2009; 104: 1085-1094.