

Exploring the potential mechanism and screening small molecule drugs for glaucoma by using bioinformatics approach

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Abstract. – **BACKGROUND:** Glaucoma is a neurodegenerative disease which is the second most common cause of blindness worldwide.

AIM: To investigate the mechanism of glaucoma and identify small molecule drugs.

MATERIALS AND METHODS: Gene expression profiles of GSE2378 were downloaded from Gene Expression Omnibus (GEO) database which included 15 astrocytes from 8 and 7 donors with and without glaucoma, respectively. Then the raw data were normalized by Robust Multichip Averaging and the differentially expressed genes (DEGs) were identified with limma package in R. Moreover, the Gene Ontology and pathway enrichment analyses were performed by GOEAST and Gene Set Analysis Toolkit V2, respectively. In addition, the potential target sites of transcription factors were detected using MSigDB. Finally, small molecule drugs were screened for glaucoma treatment by Connectivity Map.

RESULTS: A total of 961 DEGs between glaucoma and normal cells were identified. These DEGs were discovered mainly involved in cell surface, molecule binding, changes in protein activity and signal transduction. Additionally, the most significant pathway was pathway in cancer (FDR = 0.0051). Some DEGs shared target sites of the transcription factor, such as NFκB. (FDR = 0.0132) and PBX1 (FDR = 0.0158). Luteolin (enrichment = 0.87) can simulate the state of normal cells, while vancomycin (enrichment = -0.883) and Prestwick-1082 (enrichment = -0.882) might be potential pathogenic substances.

CONCLUSIONS: We hypothesize that glaucoma cells may be not only caused by the optic nerve cells themselves, but also caused by infections due to resistance decline. All these results may facilitate glaucoma treatment with a new breakthrough.

Key Words:

Glaucoma, Differentially expressed gene, Gene Ontology, Pathway enrichment analysis, Small molecules.

Introduction

Glaucoma is a dangerous eye disease that can cause blindness by optic nerve damage¹. Some glaucoma cases have rapid onset and great harm². Glaucoma has become the second leading cause of blindness globally according to statistics gathered by World Health Organization in 2002³. People over 40 years old are more likely to suffer from glaucoma, and it is popular in female patients^{4,5}. Therefore, the research and treatment of glaucoma are of great significance to human health.

With the development of bioinformatics, a number of researchers have utilized this approach to explore the potential molecular mechanism of glaucoma⁶. Johnson et al⁷ have discovered that during the pathogenesis of glaucoma the most significant up-regulated gene may affect cell cycle, cytoskeleton, and immune system process, and the down-regulated genes change glucose and lipid metabolism. Miao et al⁸ have predicted that the potential mechanism of glaucoma is the changes of signal transduction, response to stress, ECM genes, migration and cell adhesion functions. Paylakhi et al⁹ have found many of glaucoma related genes are enriched in biologic pathways, including focal adhesion and extracellular matrix. Gene set enrichment analysis of altered transcriptional pathways in glaucoma shows that the changes in apoptotic, pro-sur-

vival and pro-death may be associated with the pathogenesis of glaucoma¹⁰. Despite there are many studies about glaucoma, the mechanism of it are still not fully understood.

Many diseases are caused by hereditary mutations and an increasing number of the identified disease-related mutations occur in gene regulatory sequences. Laurila and Lähdesmäki¹¹ have investigated the effect of mutations on transcription factor binding affinity computationally. For example, the mutation in ALOX changes its binding status with transcription factor SPI1, which results in inflammatory effects¹². Mutation of HBD also affect its binding with transcription factor GATA1, which finally leads to δ -thalassemia¹³. Erb C. has discovered some risk factors can induce the transcription factor NF κ B to stimulate differential gene expression¹⁴. Therefore, the target sites of transcription factor play important roles in the pathogenesis of glaucoma.

Although many drugs for the treatment of glaucoma have been found in the past decades, there is an urgent need to discover some molecular drugs with more efficient and selective. Based on the genes related with disease phenotype, researchers screen drugs to repress these genes. Drug versus Disease (DvD) provides a pipeline for the comparison of drug and disease gene expression profiles from public microarray repositories¹⁵. Yeh et al¹⁶ have screened trifluoperazine as a potential anti-cancer stem cell agent that could overcome EGFR-tyrosine kinase inhibitor and chemotherapy resistance. Chen et al¹⁷ have compared the expression pattern of CCA-related genes and genes perturbed by small molecules in connectivity map, and then NVP-AUY922 is regarded as an effective treatment option for patients with CCA. In addition, using microarray technology, Claerhout et al¹⁸ have discovered vorinostat as a candidate therapy for gastric cancer. Therefore, the bioinformatics approach has been widely used in the molecule drugs identification.

In this study, to explore the molecular mechanism of glaucoma, the differentially expressed genes (DEGs) between normal and glaucoma samples were identified and conducted functional analyses. In addition, the potential target sites of transcription factors were also detected, helping to regulate expression of DEGs. Moreover, small molecules for glaucoma treatment were screened out. Our findings set new insight into the pathogenesis of glaucoma and provide several molecular drugs which have the potential to combat the disease.

Materials and Methods

Data Source

The microarray data were downloaded from Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>, accession number GSE2378¹⁹) based on the GPL8300 [HG_U95Av2] platform. Total 15 samples were obtained which included 8 and 7 astrocytes from Caucasian American donors with and without glaucoma, respectively. Meanwhile, the original CEL files and the platform probe annotation information of HG_U95Av2 were downloaded for the present study.

Screen of Differentially Expressed Genes

The raw chip data were divided into two sets, including glaucoma cells and normal cells. We used R language to analyse chip data (version v.2.13.0)²⁰. First, RMA(Robust Multi-chip Averaging) method²¹ was used for chip normalization. Then the limma package in R²² was utilized to screen the DEGs and the multiple testing correction were conducted to adjust the raw p values to false discovery rate (FDR) by Benjamini and Hochberg (BH)²³. Finally, the genes with FDR < 0.05 were identified as the DEGs.

Detecting Significant Gene Ontology (GO) Terms of DEGs

To explore changes of DEGs occurring at the cellular level and detect the functions of DEGs, we elaborated gene function and cellular location based on Gene Ontology (GO)²⁴. GO enrichment analysis was performed using Gene Ontology Enrichment Analysis Software Toolkit (GOEAST)²⁵ based on hypergeometric algorithm. Finally, the significant cellular component, molecular function and biological process, in which the DEGs were involved, were identified with FDR less than 0.1, 0.1 and 0.05, respectively.

If DEGs contains valid IDs of k genes from a microarray with a total of t genes, for a given GO term, there are q genes within k and m genes within t , then the possibility that whether genes associated with this GO term is enriched by hypergeometric algorithm,

$$p(X = x > q) = \sum_{x=q}^m \frac{\binom{m}{x} \binom{t-m}{k-x}}{\binom{t}{k}}$$

Detecting Significant Pathways

After elaborating gene function and cellular location based on GO, we further explored how these DEGs interacted with other genes in a pathway. All the metabolic and non-metabolic pathways were obtained from KEGG (Kyoto encyclopedia of genes and genomes) Pathway database²⁶ and the enriched pathways were identified using Gene Set Analysis Toolkit V2^{27, 28} based on hypergeometric algorithm. If DEGs contains valid IDs of k genes from a microarray with a total of t genes, for a given pathway, there are q genes within k and m genes within t , the possibility is calculated as following,

$$p(X = x > q) = \sum_{x=q}^m \frac{\binom{m}{x} \binom{t-m}{k-x}}{\binom{t}{k}}$$

Then BH method²³ was used to adjust the raw p-values into false discovery rate (FDR). Finally, the significant pathways were identified with FDR < 0.05 and count > 2.

Potential Transcription Factor Target Sites

The gene enrichment analysis was performed based on gene annotation data in MSigDB (molecular signature database, <http://www.broadinstitute.org/gsea/msigdb/index.jsp>) using hypergeometric algorithm, and then BH adjustment was utilized for multiple test correction. Finally, the potential target sites of the transcription factors were identified with FDR < 0.05.

Expression Profiles of cell Lines Perturbed by Small Molecules

Connectivity Map²⁹ (CMap, <http://www.broad.mit.edu/ConnectivityMap/>) consists of more than 7,000 gene expression profiles treated with 1,309 small molecules. These expression profiles represent about 6,000 instances, each of which comprises a treatment and vehicle pair. By comparing the expression pattern of DEGs and the genes perturbed by molecules in CMap, a list of significant molecules related to the disease were identified with FDR < 0.05. Finally, a correlation score for each perturbagen was calculated, ranging from -1 to 1.

KS scores for both up (KS_{up}) and down (KS_{down}) were calculated as follows:

$$a = \max_{j=1}^t \left[\frac{j}{t} - \frac{V(j)}{N} \right] \quad b = \max \left[\frac{V(j)}{N} - \frac{j-l}{t} \right]$$

$$KS_{up/down} = \begin{cases} a, (a > b) \\ -b, (b > a) \end{cases}$$

Where, t is the number of genes in either the up- or down-regulated gene group, N is the total number genes in array, j denotes the j th gene in the rank ordered up- or down-regulated groups according to the extent of differential expression, $V(j)$ denotes the position of the j th gene in the rank ordered whole gene list (also ranked according to the extent of differential expression). The connectivity S scores set to zero where KS_{up} and KS_{down} have the same sign, otherwise, to be $KS_{up} - KS_{down}$.

Results

DEGs Between Glaucoma Cells and Normal Cells

We identified DEGs between glaucoma cells and normal cells using linear models with FDR < 0.05. Finally, 1110 probes were discovered differential expression, which involved 961 DEGs.

Enriched GO terms of DEGs

DEGs were enriched into GO terms using GOEAST from three aspects, including cellular component, molecular function and biological process.

From the aspect of cellular component (Figure 1), differential genes of glaucoma mainly involved in cell surface and extracellular matrix, including cell membrane and extracellular matrix composition. In addition, DEGs were found mainly affected the molecular functions included molecule binding and changes in protein activity (Figure 2). Finally, multiple biological processes in which the DEGs anticipated, including signal transduction, MAPK Cascade pathway, JAK-STAK pathways, cell response were identified (Figure 3).

Pathways Disturbed in Glaucoma Cells

After the pathway enrichment analysis, total 12 pathways with FDR < 0.05 and count > 2 were identified (Table I). Among these pathways, the most significant one was pathway in cancer (FDR = 0.0051), and the other pathways

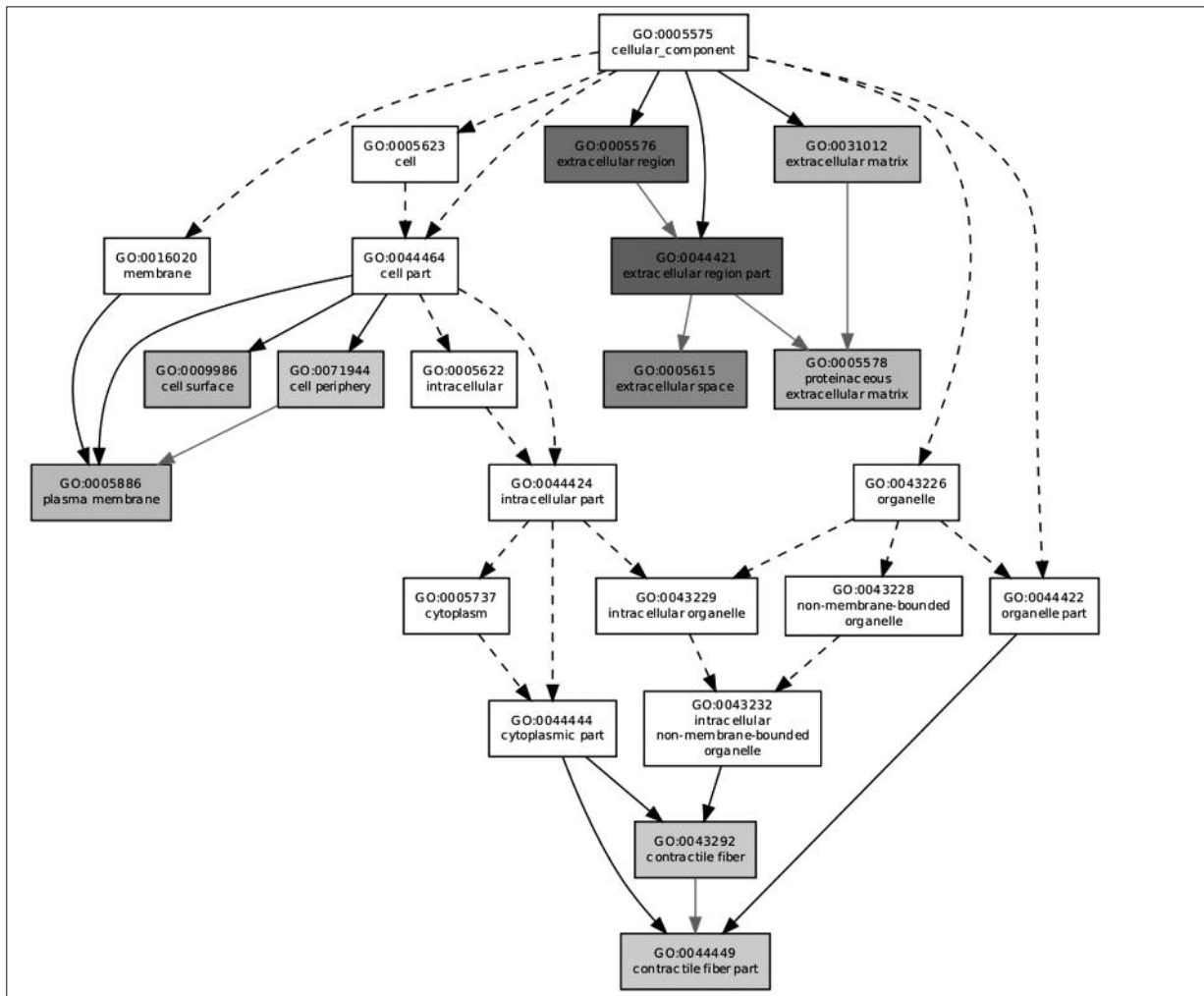


Figure 1. Enriched cellular components of DEGs. Significantly enriched terms are colored gray (FDR < 0.1) and the deeper color represents the more significance.

mainly included MAPK signaling pathway (FDR = 0.0219) and Jak-STAT signaling pathway (0.0306).

Potential Transcription Factor Target Sites

The potential target sites of the transcription factors were identified based on MSigDB with FDR < 0.05 (Table II). The potential target sites of 30 transcription factors were detected, including NF κ B (FDR = 0.0132) and PBX1 (FDR = 0.0158).

Glaucoma Related Small Molecules

In order to screen out small molecule drugs, we performed computational bioinformatics analysis of DEGs using the CMap. Total 20 related small molecules with a highly significant correlation were identified, including 13 nega-

tive molecules and 7 positive ones. The most significant molecules included Luteolin (enrichment = 0.87), vancomycin (enrichment = -0.883) and Prestwick-1082 (enrichment = -0.882) (Table III).

Discussion

Glaucoma is a dangerous eye disease that can cause blindness by optic nerve damage. Some glaucoma cases have rapid onset and great harm. In this study, we identified 961 DEGs based on gene expression profile of glaucoma cells and normal cells. Then the DEGs were discovered involved in several intracellular signal and metabolic processes during the GO and KEGG pathway analyses. Additionally, the potential target

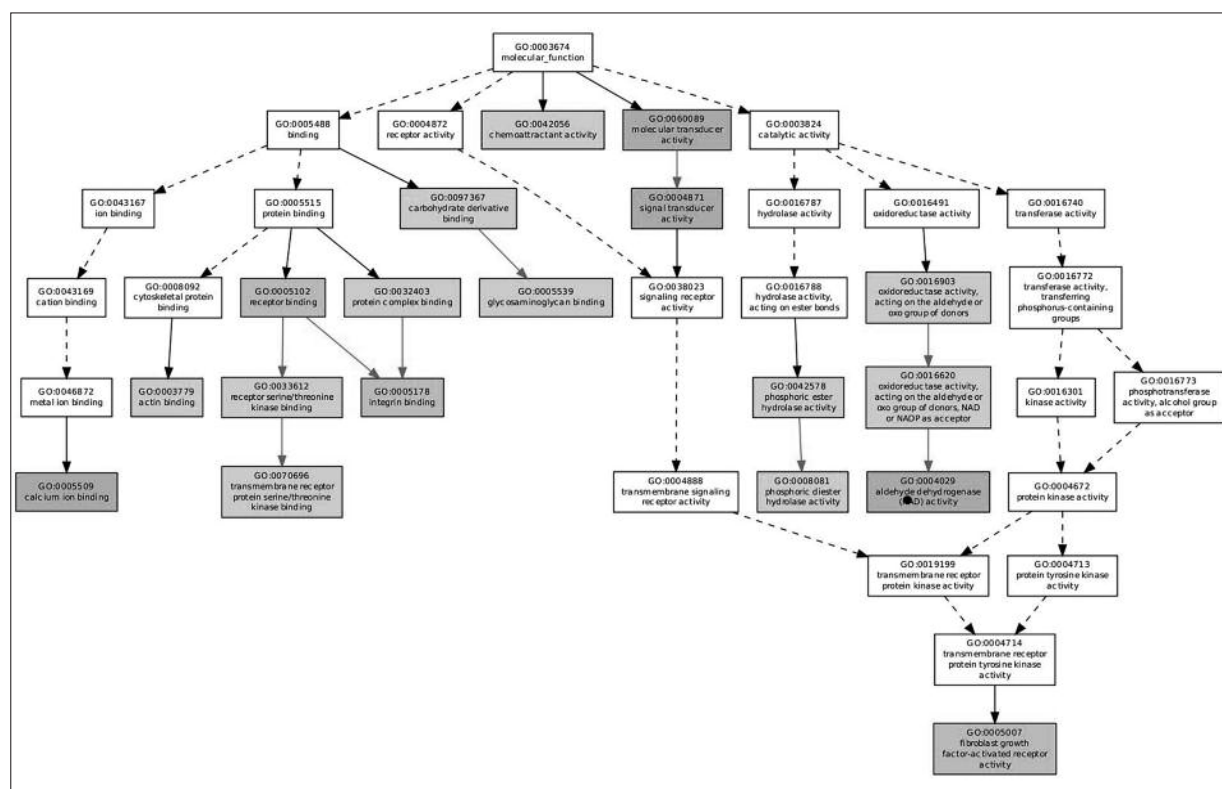


Figure 2. Enriched molecular function of DEGs. Significantly enriched terms are colored gray (FDR < 0.1) and the deeper color represents the more significance.

sites of the transcription factor were detected. Finally, small molecule drugs were identified for glaucoma treatment.

From GO and KEGG pathway enrichment analyses, a lot of changes occur in glaucoma cells compared to normal cells. Besides intracellular changes in signal, metabolic processes are also dramatically changed. DEGs involve in many biological aspects, including cell surface, extracellular matrix and intracellular shrinkage fiber, which is consistent with the previous studies^{30,31}. Changes in terms, such as the calcium ion binding, actin binding, kinase binding, integrin protein binding, and the activity of the signal transduction indicates that glaucoma cells may change in signal transduction^{32,33}. Metabolic related pathways are also related with glaucoma, such as glycolysis/gluconeogenesis, valine, leucine and isoleucine degradation³⁴⁻³⁷. These abnormal metabolites also help early diagnosis and treatment of glaucoma. Cell morphology related pathways are also included such as regulation of actin cytoskeleton³⁸. Signaling transduction related pathways involves MAPK signaling pathway, Jak-STAT signaling pathway and focal adhe-

sion^{7,39,40}. DEGs of glaucoma enrich in multiple pathways, some of which are also shared by other diseases, such as pathway in cancer. Therefore, further study on these same pathways not only provides a theoretical basis for glaucoma treatment, but also facilitates the treatment of other diseases. Besides, immune related pathway is also included such as complement and coagulation cascades^{2,41}.

In vitro and *in vivo* in a monkey model of experimental glaucoma (ExpG), NF- κ B mRNA expression and nuclear NF- κ B protein was higher in glaucomatous optic nerve heads (ONH) astrocytes than in normal ONH astrocytes⁴². The transcriptional regulatory activity of FOXC1, whose mutations results in an increased susceptibility to glaucoma, is impaired by PBX1 in a filamin A-mediated manner⁴³. Combining the previous and the present results, we predict that the target sites of transcription factor play important roles in the pathogenesis of glaucoma.

A number of small molecules are obtained by comparing the expression pattern of DEGs and that of genes perturbed by small molecules. Small molecules vancomycin can simulate the

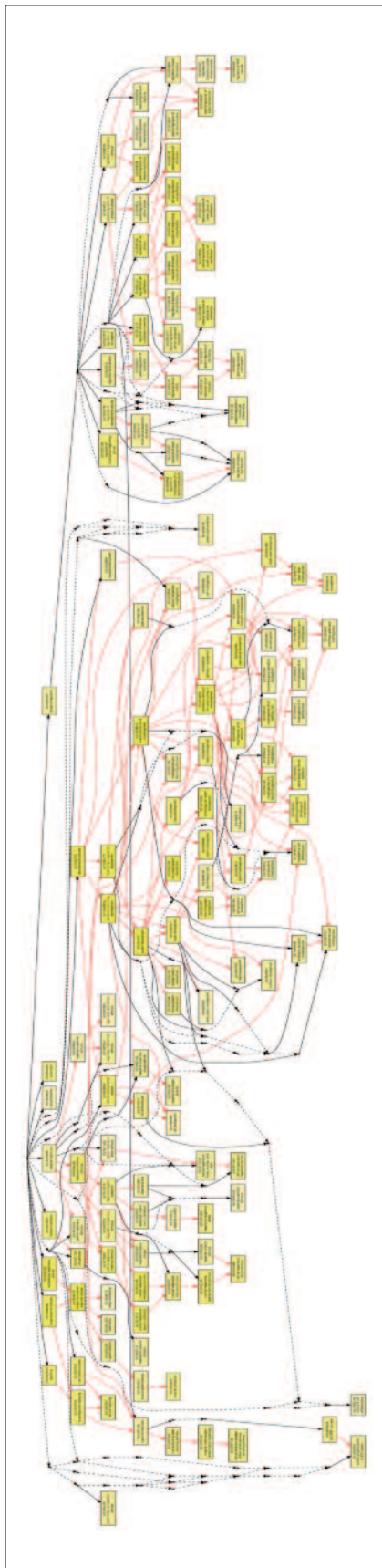


Figure 3. Enriched biological processes of DEGs. Significantly enriched terms are colored gray (FDR < 0.05) and the deeper color represents the more significance.

Table I. The significant pathways in glaucoma cells with FDR < 0.05.

KEGG pathway	FDR
Pathways in cancer	0.0051
Focal adhesion	0.0051
Ascorbate and aldarate metabolism	0.0204
Regulation of actin cytoskeleton	0.0219
Complement and coagulation cascades	0.0219
MAPK signaling pathway	0.0219
Glycolysis/Gluconeogenesis	0.0219
Limonene and pinene degradation	0.0230
Jak-STAT signaling pathway	0.0306
Valine, leucine and isoleucine degradation	0.0306
beta-Alanine metabolism	0.0334
Bladder cancer	0.0446

Note: KEGG is Kyoto encyclopedia of genes and genomes and the KEGG PATHWAY database contains pathway maps for molecular systems in both normal and perturbed states.

Table II. The enriched potential target sites of the transcription factor with FDR < 0.05

Target	FDR
hsa_RTAAACA_V\$FREAC2_01	0.0003
hsa_AACTTT_UNKNOWN	0.0003
hsa_V\$PAX4_04	0.0008
hsa_CTTTGT_V\$LFE1_Q2	0.002
hsa_TTGTTT_V\$FOXO4_01	0.0029
hsa_V\$HFH3_01	0.0099
hsa_V\$FOX_Q2	0.0132
hsa_V\$NFkB_Q6	0.0132
hsa_V\$HSF2_01	0.0132
hsa_V\$CHX10_01	0.0137
hsa_AAAYWAACM_V\$HFH4_01	0.0137
hsa_V\$HFH4_01	0.0137
hsa_CAGCTG_V\$AP4_Q5	0.0137
hsa_V\$MYOD_Q6	0.0158
hsa_V\$PBX1_02	0.0158
hsa_AAANWWTGC_UNKNOWN	0.0186
hsa_V\$HNF4_Q6	0.0231
hsa_V\$AREB6_04	0.0231
hsa_WWTAAGGC_UNKNOWN	0.025
hsa_V\$SRF_Q4	0.0311
hsa_V\$MEF2_03	0.0311
hsa_WGGAATGY_V\$TEF1_Q6	0.0351
hsa_V\$SRF_Q5_01	0.0387
hsa_CATTGTYY_V\$SOX9_B1	0.0428
hsa_V\$SRF_Q6	0.0428
hsa_V\$AFP1_Q6	0.0457
hsa_V\$MYOGNF1_01	0.0495
hsa_V\$NFkB_C	0.0495
hsa_V\$SRF_C	0.0495
hsa_V\$FOXJ2_02	0.0495

Table III. List of glaucoma related small molecules.

CMap name	FDR	Enrichment
Genistein	0.0001	-0.506
Luteolin	0.0004	0.87
Thiamphenicol	0.00042	-0.825
Vancomycin	0.00044	-0.883
Eucatropine	0.0005	0.755
Cinchonine	0.00054	-0.875
Rimexolone	0.00058	0.855
Canadine	0.00068	-0.861
Metamizole sodium	0.00072	0.745
Naringenin	0.00092	-0.853
Monensin	0.00097	-0.72
Thiocolchicoside	0.00101	0.841
Ambroxol	0.00105	-0.844
Colchicine	0.00153	0.714
Felbinac	0.00173	-0.826
Eticlopride	0.00292	-0.803
Finasteride	0.003	-0.678
Zimeldine	0.00308	0.733
Adiphenine	0.0031	-0.729
Prestwick-1082	0.0032	-0.882

Note: CMap represents the Connectivity Map which can connect small molecules, genes and disease using gene-expression signatures.

cell state of glaucoma⁴⁴. Nine percent of the 68 glaucoma patients are treated with vancomycin⁴⁵. Twenty-one percent of glaucoma patients choose a combination of fortified aminoglycoside and vancomycin⁴⁶. All gram-positive patients are sensitive to vancomycin⁴⁷. Prestwick-1082 (enrichment = -0.882) was associated with high negative scores, which suggest that this small molecules is capable to cause opposite effect with glaucoma, that is, they can be used to improve therapeutic effect for ovarian cancer. Prestwick-1082 has recently garnered some attention for its possible therapeutic efficacy on ovarian cancer⁴⁸. Such small molecules provide new possibility for glaucoma treatment. It is also found that many DEGs have the same target sites of the transcription factor. These sites play vital roles in the regulation of gene expression.

Conclusions

Through the functional analysis of DEGs, we hypothesize that glaucoma may not merely caused by the optic nerve cells themselves, it might also be caused by infections due to immunity decline. The target sites of transcription factors play vital roles in the regulation of gene ex-

pression. In addition, although it may be premature to suggest that these screened small molecules might be ready for glaucoma clinical trials, it is clearly a direction that warrants additional consideration. Furthermore, we plan to test the treatment results of these molecules on glaucoma in our future research. All these results may facilitate glaucoma treatment with a new breakthrough.

Conflict of Interest

The Authors declare that there are no conflicts of interest.

References

- 1) PFLUGFELDER SC, BAUDOIN C. Challenges in the clinical measurement of ocular surface disease in glaucoma patients. *Clin Ophthalmol* 2011; 5: 1575-1583.
- 2) HOWELL GR, MACALINAO DG, SOUSA GL, WALDEN M, SOTO I, KNEELAND SC, BARBAY JM, KING BL, MARCHANT JK, HIBBS M, STEVENS B, BARRES BA, CLARK AF, LIBBY RT, JOHN SW. Molecular clustering identifies complement and endothelin induction as early events in a mouse model of glaucoma. *J Clin Invest* 2011; 121: 1429-1444.
- 3) KINGMAN S. Glaucoma is second leading cause of blindness globally. *Bull World Health Organ* 2004; 82: 887-888.
- 4) TANITO M, KAIJZU S, TAKAI Y, OHIRA A. Status of systemic oxidative stresses in patients with primary open-angle glaucoma and pseudoexfoliation syndrome. *PLoS One* 2012; 7: e49680.
- 5) PASQUALE LR, KANG JH. Female reproductive factors and primary open-angle glaucoma in the Nurses' Health Study. *Eye (Lond)* 2011; 25: 633-641.
- 6) ALMASIEH M, WILSON AM, MORQUETTE B, CUEVA VARGAS JL, DI POLO A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res* 2012; 31: 152-181.
- 7) JOHNSON EC, DOSER TA, CEPURNA WO, DYCK JA, JIA L, GUO Y, LAMBERT WS, MORRISON JC. Cell proliferation and interleukin-6-type cytokine signaling are implicated by gene expression responses in early optic nerve head injury in rat glaucoma. *Invest Ophthalmol Vis Sci* 2011; 52: 504-518.
- 8) MIAO H, CHEN L, RIORDAN SM, LI W, JUAREZ S, CRABB AM, LUKAS TJ, DU P, LIN SM, WISE A, AGAPOVA OA, YANG P, GU CC, HERNANDEZ MR. Gene expression and functional studies of the optic nerve head astrocyte transcriptome from normal African Americans and Caucasian Americans donors. *PLoS One* 2008; 3: e2847.
- 9) PAYLAKHI SH, YAZDANI S, APRIL C, FAN JB, MOAZZENI H, RONAGHI M, ELAHI E. Non-housekeeping genes expressed in human trabecular meshwork cell cultures. *Mol Vis* 2012; 18: 241-254.

- 10) WANG DY, RAY A, RODGERS K, ERGORUL C, HYMAN BT, HUANG W, GROSSKREUTZ CL. Global gene expression changes in rat retinal ganglion cells in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2010; 51: 4084-4095.
- 11) LAURILA K, LAHDESMÄKI H. Systematic analysis of disease-related regulatory mutation classes reveals distinct effects on transcription factor binding. *In Silico Biol* 2009; 9: 209-224.
- 12) WITTMER J, MARTI-JAUN J, HERSBERGER M. Functional polymorphism in ALOX15 results in increased allele-specific transcription in macrophages through binding of the transcription factor SPI1. *Hum Mutat* 2006; 27: 78-87.
- 13) MATSUDA M, SAKAMOTO N, FUKUMAKI Y. Delta-thalassemia caused by disruption of the site for an erythroid-specific transcription factor, GATA-1, in the delta-globin gene promoter. *Blood* 1992; 80: 1347-1351.
- 14) ERB C. [Importance of the nuclear factor kappaB for the primary open angle glaucoma--a hypothesis]. *Klin Monbl Augenheilkd* 2010; 227: 120-127.
- 15) PACINI C, IORIO F, GONCALVES E, ISKAR M, KLABUNDE T, BORK P, SAEZ-RODRIGUEZ J. DvD: An R/Cytoscape pipeline for drug repurposing using public repositories of gene expression data. *Bioinformatics* 2013; 29: 132-134.
- 16) YEH CT, WU AT, CHANG PM, CHEN KY, YANG CN, YANG SC, HO CC, CHEN CC, KUO YL, LEE PY, LIU YW, YEN CC, HSIAO M, LU PJ, LAI JM, WANG LS, WU CH, CHIOU JF, YANG PC, HUANG CY. Trifluoperazine, an antipsychotic agent, inhibits cancer stem cell growth and overcomes drug resistance of lung cancer. *Am J Respir Crit Care Med* 2012; 186: 1180-1188.
- 17) CHEN MH, LIN KJ, YANG WL, KAO YW, CHEN TW, CHAO SC, CHANG PM, LIU CY, TZENG CH, CHAO Y, YEH CN, HUANG CY. Gene expression-based chemical genomics identifies heat-shock protein 90 inhibitors as potential therapeutic drugs in cholangiocarcinoma. *Cancer* 2013; 199: 293-303.
- 18) CLAERHOUT S, LIM JY, CHOI W, PARK YY, KIM K, KIM SB, LEE JS, MILLS GB, CHO JY. Gene expression signature analysis identifies vorinostat as a candidate therapy for gastric cancer. *PLoS One* 2013; 6: e24662.
- 19) HERNANDEZ MR, AGAPOVA OA, YANG P, SALVADOR-SILVA M, RICARD CS, AOI S. Differential gene expression in astrocytes from human normal and glaucomatous optic nerve head analyzed by cDNA microarray. *Glia* 2002; 38: 45-64.
- 20) TEAM RDC, R: A language and environment for statistical computing. 2005, ISBN 3-900051-07-0. R Foundation for Statistical Computing, Vienna, Austria, 2013. url: <http://www.R-project.org>.
- 21) IRIZARRY RA, HOBBS B, COLLIN F, BEAZER-BARCLAY YD, ANTONELLIS KJ, SCHERF U, SPEED TP. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* 2003; 4: 249-264.
- 22) SMYTH GK. Linear models and empirical bayes methods for assessing differential expression in microarray experiments. *Stat Appl Genet Mol Biol* 2004; 3: Article3.
- 23) BENJAMINI Y, HOCHBERG Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol* 1995; 57: 289-300.
- 24) ASHBURNER M, BALL CA, BLAKE JA, BOTSTEIN D, BUTLER H, CHERRY JM, DAVIS AP, DOLINSKI K, DWIGHT SS, EPPIG JT, HARRIS MA, HILL DP, ISSEL-TARVER L, KASARSKIS A, LEWIS S, MATESE JC, RICHARDSON JE, RINGWALD M, RUBIN GM, SHERLOCK G. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet* 2000; 25: 25-29.
- 25) ZHENG Q, WANG XJ. GOEAST: a web-based software toolkit for Gene Ontology enrichment analysis. *Nucleic Acids Res* 2008; 36: W358-363
- 26) KOTERA M, HIRAKAWA M, TOKIMATSU T, GOTO S, KANEHISA M. The KEGG databases and tools facilitating omics analysis: latest developments involving human diseases and pharmaceuticals, in *Next Generation Microarray Bioinformatics*. Springer, 2012; pp. 19-39.
- 27) ZHANG B, KIROV S, SNODDY J. WebGestalt: an integrated system for exploring gene sets in various biological contexts. *Nucleic Acids Res* 2005; 33: W741-748.
- 28) DEXTER DUNCAN NPABZ. WebGestalt2: an updated and expanded version of the Web-based Gene Set Analysis Toolkit. *BMC Bioinformatics* 2010; Suppl 4: 10.
- 29) LAMB J, CRAWFORD ED, PECK D, MODELL JW, BLAT IC, WROBEL MJ, LERNER J, BRUNET JP, SUBRAMANIAN A, ROSS KN, REICH M, HIERONYMUS H, WEI G, ARMSTRONG SA, HAGGARTY SJ, CLEMONS PA, WEI R, CARR SA, LANDER ES, GOLUB TR. The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and disease. *Science* 2006; 313: 1929-1935.
- 30) GREGORY MS, HACKETT CG, ABERNATHY EF, LEE KS, SAFF RR, HOHLBAUM AM, MOODY KS, HOBSON MW, JONES A, KOLOVOU P, KARRAY S, GIANI A, JOHN SW, CHEN DF, MARSHAK-ROTHSTEIN A, KSANDER BR. Opposing roles for membrane bound and soluble Fas ligand in glaucoma-associated retinal ganglion cell death. *PLoS One* 2011; 6: e17659.
- 31) SCHLOTZER-SCHREHARDT U. New pathogenetic insights into pseudoexfoliation syndrome/glaucoma: Therapeutically relevant? *Ophthalmologie* 2012; 109: 944-951.
- 32) SACCA SC, CENTOFANTI M, IZZOTTI A. New proteins as vascular biomarkers in primary open angle glaucomatous aqueous humor. *Invest Ophthalmol Vis Sci* 2012; 53: 4242-4253.
- 33) NAGABHUSHANA A, CHALASANI ML, JAIN N, RADHA V, RANGARAJ N, BALASUBRAMANIAN D, SWARUP G. Regulation of endocytic trafficking of transferrin receptor by optineurin and its impairment by a glaucoma-associated mutant. *BMC Cell Biol* 2010; 11: 4.

- 34) NAKAMURA M. New insights into the pathogenesis of glaucomatous optic neuropathy and refinement of the objective assessment of its functional damage. *Nihon Ganka Gakkai Zasshi* 2012; 116: 298-344; discussion 345-296.
- 35) MESSINA-BAAS OM, GONZALEZ-HUERTA LM, CHIMA-GALAN C, KOFMAN-ALFARO SH, RIVERA-VEGA MR, BABAYAN-MENA I, CUEVAS-COVARRUBIAS SA. Molecular analysis of the CYP1B1 gene: identification of novel truncating mutations in patients with primary congenital glaucoma. *Ophthalmic Res* 2007; 39: 17-23.
- 36) MANSERGH FC, KENNA PF, AYUSO C, KIANG AS, HUMPHRIES P, FARRAR GJ. Novel mutations in the TIGR gene in early and late onset open angle glaucoma. *Hum Mutat* 1998; 11: 244-251.
- 37) RICHARDS JE, RITCH R, LICHTER PR, ROZSA FW, STRINGHAM HM, CARONIA RM, JOHNSON D, ABUNDO GP, WILLCOCKSON J, DOWNS CA, THOMPSON DA, MUSARELLA MA, GUPTA N, OTHMAN MI, TORREZ DM, HERMAN SB, WONG DJ, HIGASHI M, BOEHNKE M. Novel trabecular meshwork inducible glucocorticoid response mutation in an eight-generation juvenile-onset primary open-angle glaucoma pedigree. *Ophthalmology* 1998; 105: 1698-1707.
- 38) KWON HS, TOMAREV SI. Myocilin, a glaucoma-associated protein, promotes cell migration through activation of integrin-focal adhesion kinase-serine/threonine kinase signaling pathway. *J Cell Physiol* 2011; 226: 3392-3402.
- 39) EDWARD DP, MORALES J, BOUHENNI RA, PATIL J, EDWARD PR, CUMMINGS TJ, CHAUDHRY IA, ALKATAN H. Congenital ectropion uvea and mechanisms of glaucoma in neurofibromatosis type 1: new insights. *Ophthalmology* 2012; 119: 1485-1494.
- 40) SCHLUNCK G, HAN H, WECKER T, KAMPIK D, MEYER-TERVEHN T, GREHN F. Substrate rigidity modulates cell matrix interactions and protein expression in human trabecular meshwork cells. *Invest Ophthalmol Vis Sci* 2008; 49: 262-269.
- 41) DING OJ, COOK AC, DUMITRESCU AV, KUEHN MH. Lack of immunoglobulins does not prevent C1q binding to RGC and does not alter the progression of experimental glaucoma. *Invest Ophthalmol Vis Sci* 2012; 53: 6370-6377.
- 42) AGAPOVA OA, KAUFMAN PL, HERNANDEZ MR. Androgen receptor and NFkB expression in human normal and glaucomatous optic nerve head astrocytes in vitro and in experimental glaucoma. *Exp Eye Res* 2006; 82: 1053-1059.
- 43) BERRY FB, O'NEILL MA, COCA-PRADOS M, WALTER MA. FOXC1 transcriptional regulatory activity is impaired by PBX1 in a filamin A-mediated manner. *Mol Cell Biol* 2005; 25: 1415-1424.
- 44) CHAN CC, HOLLAND EJ. Infectious endophthalmitis after Boston type 1 keratoprosthesis implantation. *Cornea* 2012; 31: 346-349.
- 45) CHIAM PJ, ARASHVAND K, SHAIKH A, JAMES B. Management of blebitis in the United Kingdom: a survey. *Br J Ophthalmol* 2012; 96: 38-41.
- 46) REYNOLDS AC, SKUTA GL, MONLUX R, JOHNSON J. Management of blebitis by members of the American Glaucoma Society: a survey. *J Glaucoma* 2001; 10: 340-347.
- 47) LENG T, MILLER D, FLYNN HW, JR., JACOBS DJ, GEDDE SJ. Delayed-onset bleb-associated endophthalmitis (1996-2008): causative organisms and visual acuity outcomes. *Retina* 2011; 31: 344-352.
- 48) SUN N, ZANG W, LI W. Bioinformatics analysis reveals potential candidate drugs for psychological stress in ovarian cancer. *Eur Rev Med Pharmacol Sci* 2012; 16: 1362-1366.