

MiRNA153 induces pituitary tumor MMQ cell line apoptosis through down-regulating Skp protein expression

Z.-R. ZHAO, M. LI, P. SHI, P. ZHANG

Department of Neurology, Yantai Yeda Hospital, Yantai, Shandong, China

Z.-R. Zhao and M. Li contributed equally to this work

Abstract. – **OBJECTIVE:** Pituitary tumor seriously threatens patient's life. MicroRNAs regulate cell growth and apoptosis. This study aims to investigate the effects of miRNA153 on pituitary tumor MMQ cells proliferation and apoptosis.

MATERIALS AND METHODS: Synthetic miRNA153 and control miRNA were transfected to MMQ cell line. Cell proliferation and apoptosis were tested by MTT assay and flow cytometry, respectively. Skp protein expression was detected by Western blot assay. Skp siRNA or Skp plasmid was transfected to MMQ cells transfected by miRNA153 to evaluate the influence on MMQ cell apoptosis.

RESULTS: MMQ cell proliferation was inhibited and apoptosis was enhanced after miRNA153 transfection. Skp protein level decreased in MMQ cells transfected by miRNA153. Skp interference enhanced MMQ cell apoptosis induced by miRNA153. Skp overexpression restrained MMQ cell apoptosis triggered by miRNA153.

CONCLUSIONS: MiRNA153 transfection suppressed MMQ cell growth and induced apoptosis. MiRNA153 regulated MMQ cell apoptosis through down-regulating Skp protein.

Key Words:

miRNA153, Skp protein, Pituitary tumor MMQ cell line, Apoptosis.

Introduction

Pituitary tumor is an important type of neurological tumor¹. Though the pathological basis is still unclear, hormones abnormal secretion, aging, and genetic factors are considered to be the main decision factors². As a synergy factor of pituitary tumor infection, smoking may increase the risk of pituitary tumor. Also, mental pressure and poor immune condition can affect the occurrence of the disease. Pituitary tumor brings heavy physical and mental burden to patients^{3,4}. Surgery, radiotherapy, and chemotherapy are commonly

used for the treatment of pituitary tumor. However, they also have many kinds of shortcomings and deficiencies, such as bleeding and other side effects⁵⁻⁷. It is an important and difficult topic to improve the accuracy and success rate of pituitary tumor treatment. Molecular targeted therapy is a new way for tumor treatment⁸⁻¹⁰. Its difficulty and key point are the choices of molecular targets. Moreover, it showed poor efficacy on pituitary tumor¹¹. Therefore, more effective molecular targets for pituitary tumor are seriously needed in clinical application^{11,12}. More importantly, it is still a lack of miRNA as targets for the treatment of pituitary tumor¹². MiRNAs regulate multiple biological processes including cell cycle and survival. For instance, miRNA218 can suppress pituitary tumor growth, while miRNA34a is related to tumor metastasis^{13,14}, suggesting that miRNAs may also be involved in the occurrence and development of pituitary tumor¹³⁻¹⁵. Previous studies suggested that miRNA153 was significantly increased in pituitary tumor tissue compared with paracarcinoma tissue, revealing that miRNA153 may participate in the development of pituitary tumor^{16,17}. This study intended to explore the potential regulatory effect of miRNA153 on pituitary tumor MMQ cells. The strategy of anti-tumor is to kill tumor cells without affecting normal cells. Apoptosis is regulated by anti-apoptotic proteins and pro-apoptotic proteins^{18,19}. Ideal anti-cancer drug can reduce anti-apoptotic protein levels and up-regulate pro-apoptotic protein expressions. Skp protein is a widely investigated antiapoptotic molecule^{20,21}. At present, although many drugs targeted Skp protein, their effect in reducing Skp level is still unsatisfactory²². This study also tried to study the potential molecules that can target Skp protein. Therefore, it took pituitary tumor cell line MMQ to discuss the role and possible mechanism of miRNA153 on pituitary tumor.

Materials and Methods

Materials and Cell Model

MMQ cell line was purchased from ATCC cell bank (Manassas, VA, USA). Fetal bovine serum (FBS) and Dulbecco's Modified Eagle's medium (DMEM) were bought from Gibco (Rockville, MD, USA). MiRNA153 (5'-TCACTA-TCGGAGGTCATCCAAT-3' and 5'-AGGT-GTAAAAGAAAGAACGCA-3'), scramble miRNA (5'-CCCATGGTTACCTATGAGGC-3' and 5'-TTTCGCACAATGTCAGATT-3'), Skp siRNA (5'-CTATCCATCGAGGCTAGTGC-3' and 5'-TCGATTATTCACTAAGCGT-3'), and Skp plasmid were got from GenePharma (Shanghai, China). Lipofectamine cell transfection kit was from Invitrogen (Carlsbad, CA, USA). MTT was bought from Beijing Dingguo Changsheng Biotechnology Co. Ltd. (Beijing, China). FITC-annexin, Caspase-3 detection kit, and Western blot related reagents were from Beyotime Biotechnology. Co. Ltd. (Shanghai, China). IgG mouse anti-human Skp and actin monoclonal antibodies were from Santa Cruz Biotechnology (Santa Cruz, CA, USA).

Cell Culture

MMQ cells were resuscitated and cultured in high glucose Dulbecco's Modified Eagle's medium (DMEM); Gibco (Rockville, MD, USA) medium⁹.

Transfection

MiRNA153 and scramble miRNA were transfected into MMQ cells using lipofectamine method. MMQ cells were seeded in 6-well plate at the density of 80%. A total of 1 μ l miRNA153 or scramble miRNA at 1 μ g/ μ l was suspended in lipo2000 and then transfected to MMQ cells⁹.

MTT Assay

MMQ cell viability was tested by MTT according to the conventional method¹⁰. MMQ cells were seeded in 24-well plate for 12 h. After transfected by miRNA153 or scramble miRNA, the cells were added with MTT at 2 mg/ml for 4 h. Next, the reaction was stopped by DMSO for 5 min. At last, the plate was read on microplate reader at 560 nm to obtain the absorbance value¹¹.

Flow Cytometry

MMQ cell apoptosis was tested by flow cytometry upon Annexin-V-FITC double staining method. MMQ cells were transfected with miRNA153 or scramble miRNA and collected after 48 h. The cells

in 500 μ l were mixed with 100 μ l buffer and 2 μ l Annexin-V-FITC at room temperature avoids of light for 25 min. At last, cells were tested on flow cytometry to evaluate cell apoptosis¹⁴.

Western Blot Assay

Total protein was extracted according to the kit instruction and quantified by BCA kit Gibco (Rockville, MD, USA). A total of 20 μ g protein was boiled for 6 min and separated by electrophoresis. After transferring and blocking, the membrane was incubated in antibody. At last, the membrane was washed with tris buffered saline-tween (TBST) and developed using enhanced chemiluminescence (ECL) reagent Amersham Biosciences (Piscataway, NJ, USA). The image was obtained by gel imaging system to analyze protein expression¹⁵.

Caspase-3 Activity Detection

Caspase-3 activity in MMQ cells was tested to evaluate cell apoptosis. After transfected by miRNA153 or scramble miRNA, MMQ cells were resuspended in Dulbecco's Modified Eagle Medium (DMEM) and added with chromophoric substrate at room temperature. Then the cells were put into 24-well plate and detected on microplate reader¹⁷. Caspase-3 relative activity was calculated as different value between absorbance in miRNA153 group and scramble miRNA group.

Skp Protein Interference and Overexpression

To test the impact of Skp protein interference or over-expression on MMQ cells after miRNA153 transfection, Skp siRNA or plasmid was transfected to MMQ cells upon lipo2000 according to the manual. Then miRNA153 or scramble miRNA was further transfected to MMQ cells. MMQ cells were seeded 24-well plate at the density of 70%. A total of 2 μ l Skp siRNA or plasmid at 0.5 μ g/ μ l was suspended in lipo2000 and transfected to MMQ cells. Next, a total of 4 μ l miRNA153 or scramble miRNA at 1 μ g/ μ l was suspended in lipo2000 and then transfected into MMQ cells transfected by Skp siRNA or plasmid.

Statistical Analysis

All data analysis was performed on SPSS 11.0 software (SPSS Inc. Chicago, IL, USA). The data was presented as mean \pm standard deviation and compared by *t*-test. $p < 0.05$ was depicted as statistical significance.

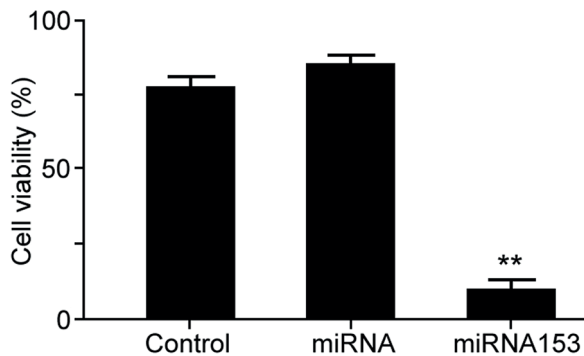


Figure 1. MiRNA153 suppressed MMQ cell viability. ** $p < 0.01$ compared with miRNA group.

Results

MiRNA153 Suppressed MMQ Cell Viability

MTT assay results showed that compared with MMQ cells transfected by 0.5 μg scramble miRNA, MMQ cell viability significantly decreased after 0.5 μg miRNA153 transfection ($p = 0.0047$) (Figure 1). Since no statistical difference was observed in cell viability between scramble miRNA transfection and normal control ($p > 0.05$), cells transfected with scramble miRNA were treated as control in the following experiments.

MiRNA153 Induced MMQ cell Apoptosis

Annexin-V-FITC staining method was applied to test MMQ cell apoptosis. It was revealed that phosphatidylserine eversion in MMQ cells transfected by 1 μg miRNA153 was enhanced compared to MMQ cells transfected by 1 μg scramble miRNA ($p = 0.0082$) (Figure 2).

MiRNA153 Activated Caspase-3 in MMQ Cells

As shown in Figure 3, caspase-3 activity in MMQ cells transfected by 0.5 μg miRNA153 was markedly higher than that in MMQ cells transfected by scramble miRNA ($p = 0.029$).

MiRNA153 Reduced Skp Protein Expression in MMQ cells

Western blot was adopted to determine Skp protein expression in MMQ cells. As shown in Figure 4, Skp protein level in MMQ cells transfected by 0.5 μg miRNA153 was significantly lower than that in MMQ cells transfected by scramble miRNA ($p = 0.056$).

Skp Knockdown Enhanced MMQ Cell Apoptosis Induced by miRNA153

To evaluate the influence of Skp protein on miRNA153 induced MMQ cell apoptosis, Skp siRNA was transfected to MMQ cells to observe the phenomenon. Western blot results demonstrated that Skp level declined after Skp siRNA transfection. Skp protein expression in Skp siRNA + miRNA153 group was lower than that in miRNA153 group (Figure 5A). Caspase-3 detection exhibited that caspase-3 activity apparently enhanced in Skp siRNA + miRNA153 group compared with miRNA153 group ($p = 0.026$) (Figure 5B).

Skp Protein Overexpression Suppressed MMQ Cell Apoptosis Induced by miRNA153

To determine the impact of Skp protein on miRNA153 induced MMQ cell apoptosis, Skp plasmid was transfected to MMQ cells to overexpress Skp. Western blot revealed that Skp level up-regulated after Skp plasmid transfection. Skp protein expression in Skp + miRNA153 group was significantly higher than that in miRNA153 group (Figure 6A). Caspase-3 detection suggested

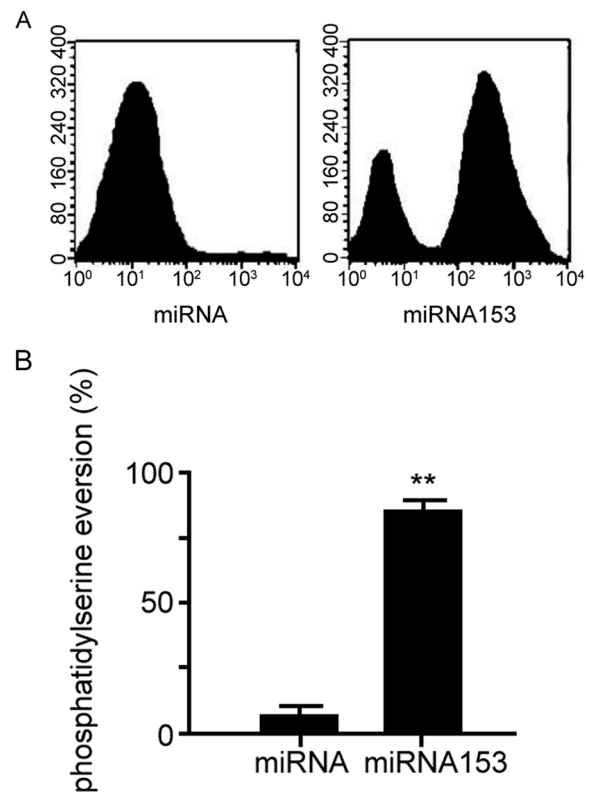


Figure 2. MiRNA153 induced MMQ cell apoptosis. ** $p < 0.01$ compared with miRNA group.

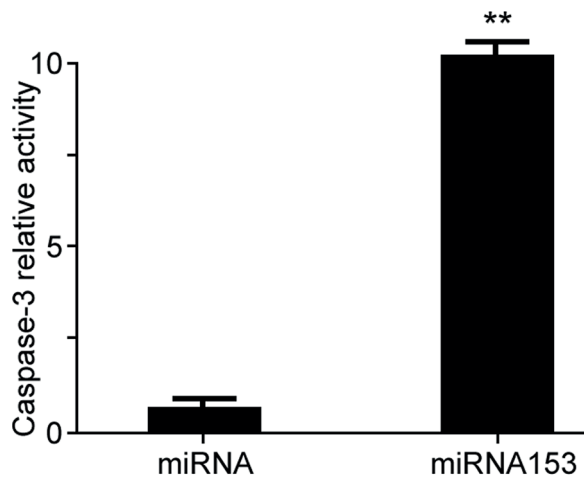


Figure 3. MiRNA153 activated Caspase-3 in MMQ cells. * $p < 0.05$ compared with miRNA group.

that caspase-3 activity apparently declined in Skp + miRNA153 group compared with miRNA153 group ($p = 0.022$) (Figure 6B).

Discussion

This study investigated the regulatory role and the possible mechanism of miRNA153 on pituitary tumor MMQ cells from molecule and protein levels. It was showed that miRNA153 transfection reduced MMQ cell viability, suppressed cell growth, and induced cell apoptosis. It was consistent with previous report, as that



Figure 4. MiRNA153 reduced Skp protein expression in MMQ cells.

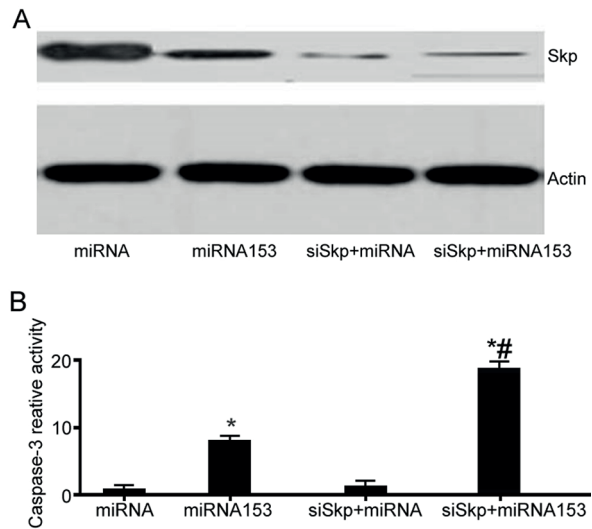


Figure 5. Skp knockdown enhanced MMQ cell apoptosis induced by miRNA153. * $p < 0.05$ compared with miRNA group. # $p < 0.05$ compared with miRNA153 group.

miRNA participated in cell growth and survival³. There is still a lack of study to investigate the impact of miRNA to the pituitary tumor³. Therefore, it has both the theoretical significance and practical value to explore miRNA regulation of tumor cell growth and survival. Skp protein is an anti-apoptotic protein²³. It is still controversy whether Skp protein is regulated by miRNA153 to mediate MMQ cell growth^{24,25}. Our results demonstrated that the miRNA153 transfection reduced Skp protein level in MMQ cells. MMQ cell apoptosis increased after miRNA153 transfection and Skp protein knockdown, while it reduced after Skp plasmid transfection. In this study, three different results proved the role of Skp protein in pituitary tumor MMQ cell apoptosis induced by miRNA153. Skp protein declined in MMQ cells after miRNA153 transfection. Skp interference enhanced MMQ cell apoptosis induced by miRNA153. Skp over-expression restrained MMQ cell apoptosis triggered by miRNA153. It suggested the role of Skp protein in MMQ apoptosis induced by miRNA153. Moreover, targeting Skp might be a new strategy for pituitary tumor treatment²⁶. At present, Skp protein also plays an anti-apoptotic role in other types of cancer cells^{22,25,27,28}. However, there is still no report about the regulation between miRNA153 and Skp protein in pituitary tumor. This study also had the following deficiencies: firstly, we did not collect the clinical specimen of pituitary tumor, thus can't discuss the rela-

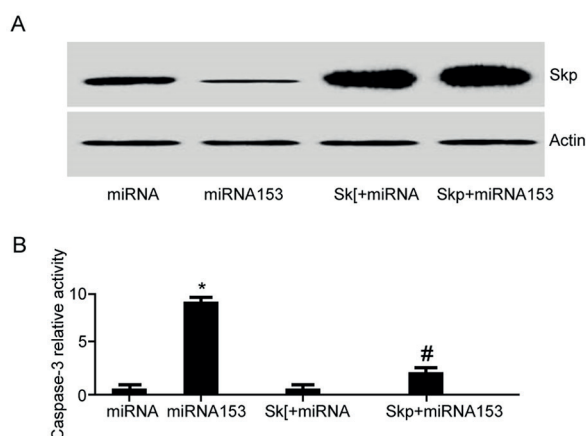


Figure 6. Skp over-expression reduced MMQ cell apoptosis induced by miRNA153. (A) Western blot detection of Skp protein expression. (B) Caspase-3 relative activity. * $p < 0.05$ compared with miRNA group. # $p < 0.05$ compared with miRNA153 group.

relationship between Skp protein and pituitary tumor in clinic; secondly, we did not obtain the pituitary tumor tissue from patients received therapy, thus we can't observe Skp protein level changes after treatment to confirm the role of Skp in pituitary tumor; lastly, we did not establish the pituitary tumor animal model to observe the curative efficacy of targeting miRNA153 on pituitary tumor *in vivo*.

Conclusions

MiRNA153 transfection suppressed pituitary tumor MMQ cell viability. MiRNA153 induced MMQ cell apoptosis through down-regulating Skp protein level. It suggested that Skp might be a potential treatment target for pituitary tumor, which may provide theoretical basis for its application.

Conflict of interest

The authors declare no conflicts of interest.

References

- 1) JIE G, GUOZHENG X, YING L, YI Z, BO D. Expression of LRIG1 in pituitary tumor and its clinical significance. *Eur Rev Med Pharmacol Sci* 2016; 20: 1969-1973.
- 2) LIANG HQ, WANG RJ, DIAO CF, LI JW, SU JL, ZHANG S. The PTTG1-targeting miRNAs miR-329, miR-300, miR-381, and miR-655 inhibit pituitary tumor cell

tumorigenesis and are involved in a p53/PTTG1 regulation feedback loop. *Oncotarget* 2015; 6: 29413-29427.

- 3) TANG J, WANG Z, CHEN L, HUANG G, HU X. Gossypol acetate induced apoptosis of pituitary tumor cells by targeting the BCL-2 via the upregulated microRNA miR-15a. *Int J Clin Exp Med* 2015; 8: 9079-9085.
- 4) YUAN B, YU WY, DAI LS, GAO Y, DING Y, YU XF, CHEN J, ZHANG JB. Expression of microRNA26b and identification of its target gene EphA2 in pituitary tissues in Yanbian cattle. *Mol Med Rep* 2015; 12: 5753-5761.
- 5) FAN X, MAO Z, HE D, LIAO C, JIANG X, LEI N, HU B, WANG X, LI Z, LIN Y, GOU X, ZHU Y, WANG H. Expression of somatostatin receptor subtype 2 in growth hormone-secreting pituitary adenoma and the regulation of miR-185. *J Endocrinol Invest* 2015; 38: 1117-1128.
- 6) RENJIE W, HAIOIAN L. MiR-132, miR-15a and miR-16 synergistically inhibit pituitary tumor cell proliferation, invasion and migration by targeting Sox5. *Cancer Lett* 2015; 356: 568-578.
- 7) LIAO C, CHEN W, FAN X, JIANG X, QIU L, CHEN C, ZHU Y, WANG H. MicroRNA-200c inhibits apoptosis in pituitary adenoma cells by targeting the PTEN/Akt signaling pathway. *Oncol Res* 2013; 21: 129-136.
- 8) GENTILIN E, TAGLIATI F, FILIERI C, MOLE D, MINOIA M, ROSARIA AMBROSIO M, DEGLI UBERTI EC, ZATELLI MC. miR-26a plays an important role in cell cycle regulation in ACTH-secreting pituitary adenomas by modulating protein kinase C delta. *Endocrinology* 2013; 154: 1690-1700.
- 9) LI J, YUAN J, YUAN XR, ZHANG C, LI HY, ZHAO J, ZHANG ZP, LIU JP. Induction effect of microRNA-449a on glioma cell proliferation and inhibition on glioma cell apoptosis by promoting PKC-alpha. *Eur Rev Med Pharmacol Sci* 2015; 19: 3587-3592.
- 10) SHI X, TAO B, HE H, SUN Q, FAN C, BIAN L, ZHAO W, LU YC. MicroRNAs-based network: a novel therapeutic agent in pituitary adenoma. *Med Hypotheses* 2012; 78: 380-384.
- 11) CHINTHARLAPALLI S, PAPANENI S, LEI P, PATHI S, SAFE S. Betulinic acid inhibits colon cancer cell and tumor growth and induces proteasome-dependent and -independent downregulation of specificity proteins (Sp) transcription factors. *BMC Cancer* 2011; 11: 371.
- 12) MAO ZG, HE DS, ZHOU J, YAO B, XIAO WW, CHEN CH, ZHU YH, WANG HJ. Differential expression of microRNAs in GH-secreting pituitary adenomas. *Diagn Pathol* 2010; 5: 79.
- 13) STILLING G, SUN Z, ZHANG S, JIN L, RIGHI A, KOVACS G, KORBONITS M, SCHEITHAUER BW, KOVACS K, LLOYD RV. MicroRNA expression in ACTH-producing pituitary tumors: up-regulation of microRNA-122 and -493 in pituitary carcinomas. *Endocrine* 2010; 38: 67-75.
- 14) AOELIAN RI, CALIN GA, CROCE CM. miR-15a and miR-16-1 in cancer: discovery, function and future perspectives. *Cell Death Differ* 2010; 17: 215-220.

- 15) BOTTONI A, ZATELLI MC, FERRACIN M, TAGLIATI F, PICCIN D, VIGNALI C, CALIN GA, NEGRINI M, CROCE CM, DEGLI UBERTI EC. Identification of differentially expressed microRNAs by microarray: a possible role for microRNA genes in pituitary adenomas. *J Cell Physiol* 2007; 210: 370-377.
- 16) JIN S, DAI Y, LI C, FANG X, HAN H, WANG D. MicroRNA-544 inhibits glioma proliferation, invasion and migration but induces cell apoptosis by targeting PARK7. *Am J Transl Res* 2016; 8: 1826-1837.
- 17) XUE H, YUAN G, GUO X, LIU Q, ZHANG J, GAO X, GUO X, XU S, LI T, SHAO Q, YAN S, LI G. A novel tumor-promoting mechanism of IL6 and the therapeutic efficacy of tocilizumab: hypoxia-induced IL6 is a potent autophagy initiator in glioblastoma via the p-STAT3-MIR155-3p-CREBRF pathway. *Autophagy* 2016; 12: 1129-1152.
- 18) STOJCHEVA N, SCHECHTMANN G, SASS S, ROTH P, FLOREA AM, STEFANSKI A, STUHLER K, WOLTER M, MULLER NS, THEIS FJ, WELLER M, REIFENBERGER G, HAPPOLD C. MicroRNA-138 promotes acquired alkylator resistance in glioblastoma by targeting the Bcl-2-interacting mediator BIM. *Oncotarget* 2016; 7: 12937-12950.
- 19) MENACHEM A, MAKOVSKI V, BODNER O, PASMANIK-CHOR M, STEIN R, SHOMRON N, KLOOG Y. Intercellular transfer of small RNAs from astrocytes to lung tumor cells induces resistance to chemotherapy. *Oncotarget* 2016; 7: 12489-12504.
- 20) TOKUDOME T, SASAKI A, TSUJI M, UDAKA Y, OYAMADA H, TSUCHIYA H, OGUCHI K. Reduced PTEN expression and overexpression of miR-17-5p, -19a-3p, -19b-3p, -21-5p, -130b-3p, -221-3p and -222-3p by glioblastoma stem-like cells following irradiation. *Oncol Lett* 2015; 10: 2269-2272.
- 21) CHAKRABARTI M, RAY SK. Anti-tumor activities of luteolin and silibinin in glioblastoma cells: overexpression of miR-7-1-3p augmented luteolin and silibinin to inhibit autophagy and induce apoptosis in glioblastoma *in vivo*. *Apoptosis* 2016; 21: 312-328.
- 22) ZHANG L, ZHANG S, YAO J, LOWERY FJ, ZHANG Q, HUANG WC, LI P, LI M, WANG X, ZHANG C, WANG H, ELLIS K, CHEERATHODI M, MCCARTY JH, PALMIERI D, SAUNUS J, LAKHANI S, HUANG S, SAHIN AA, ALDAPE KD, STEEG PS, YU D. Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth. *Nature* 2015; 527: 100-104.
- 23) FENG R, DONG L. Knockdown of microRNA-127 reverses adriamycin resistance via cell cycle arrest and apoptosis sensitization in adriamycin-resistant human glioma cells. *Int J Clin Exp Pathol* 2015; 8: 6107-6116.
- 24) WU H, LIU Q, CAI T, CHEN YD, WANG ZF. Induction of microRNA-146a is involved in curcumin-mediated enhancement of temozolomide cytotoxicity against human glioblastoma. *Mol Med Rep* 2015; 12: 5461-5466.
- 25) CHAKRABARTI M, RAY SK. Direct transfection of miR-137 mimics is more effective than DNA demethylation of miR-137 promoter to augment anti-tumor mechanisms of delphinidin in human glioblastoma U87MG and LN18 cells. *Gene* 2015; 573: 141-152.
- 26) GWAK HS, KIM TH, JO GH, KIM YJ, KWAK HJ, KIM JH, YIN J, YOO H, LEE SH, PARK JB. Silencing of microRNA-21 confers radio-sensitivity through inhibition of the PI3K/AKT pathway and enhancing autophagy in malignant glioma cell lines. *PLoS One* 2012; 7: e47449.
- 27) FAN B, JIAO BH, FAN FS, LU SK, SONG J, GUO CY, YANG JK, YANG L. Downregulation of miR-95-3p inhibits proliferation, and invasion promoting apoptosis of glioma cells by targeting CELF2. *Int J Oncol* 2015; 47: 1025-1033.
- 28) JIANG ZH, DONG XW, SHEN YC, QIAN HL, YAN M, YU ZH, HE HB, LU CD, QIU F. DNA damage regulates ARID1A stability via SCF ubiquitin ligase in gastric cancer cells. *Eur Rev Med Pharmacol Sci* 2015; 19: 3194-3200.