Functional analysis of DNA methylation in lung cancer

B. LI^{1,2}, Q. LU³, Z.-G. SONG¹, L. YANG¹, H. JIN¹, Z.-G. LI¹, T.-J. ZHAO¹, Y.-F. BAI¹, J. ZHU¹, H.-Z. CHEN¹, Z.-Y. XU¹

Abstract. – OBJECTIVES: We aimed to explore the DNA methylation difference between lung cancer samples and non-cancer lung samples, and to investigate the role of DNA methylation in the mechanism of lung cancer development. Besides, we analyzed the transcriptional regulation network of DNA methylation and the miRNAs regulated by DNA methylation. This study provides a framework for DNA methylation in other tumors or diseases.

MATERIALS AND METHODS: DNA methylation and gene expression profiles used were obtained from Gene Expression Omnibus. Firstly, we identified differentially methylated genes (DMGs) by Student's *t*-test. Then we detected the biological processes and pathways changed in lung cancer by Gene Ontology (GO) and KEGG pathway enrichment analysis. The transcriptional factors in differential genes were identified and the microRNAs regulated by them were also obtained in TransmiR.

RESULTS: We obtained 108 DMGs between lung cancer samples and non-cancer samples. Besides development related biological processes and pathways were dramatically disordered. For the DMGs, we identified 11 transcriptional factors regulating them. Moreover, we screened out 21 relationships between DMGs and their transcriptional targets. Five microRNAs are reported to be regulated by DNA methylation genes. Finally a regulation network of DNA methylation was constructed.

CONCLUSIONS: DNA methylation participates in carcinogenesis at the transcriptional and post-transcriptional level. Aberrant DNA methylation will prevent its binding with the upstream regulatory proteins, inhibit the function of downstream target genes and regulate the expression of downstream miRNA, and consequently affect cell development, immuneresponse and apoptosis.

Key Words.

Lung cancer, Transcription factor, MicroRNA, Pathway, Gene Ontology, DNA methylation.

Introduction

Lung cancer is the uncontrolled growth of aberrant cells in one or both lungs¹. Survival of

lung cancer depends on stage, overall health, and other factors. Overall, 15% of people in the United States diagnosed with lung cancer survive five years after the diagnosis². Worldwide, lung cancer is the most common cause of cancer-related death in men and women, and is responsible for 1.38 million deaths annually in 2008³. Therefore, the research and treatment of lung cancer is of great significance to human health.

Aberrant DNA hypermethylation has been implicated as a component of an epigenetic mechanism that silences genes in lung cancer⁴. Park et al⁵ investigate the significance of DNA methylation in SLC5A8 (salute carrier family 5 iodide transporter member 8) expression in lung cancer cell lines and tissues and conclude that DNA methylation in the SLC5A8 promoter region may suppress the expression of SLC5A8 in lung tumor. Tekpli et⁶ al show DNA methylation at promoter regions of interleukin (IL) 1B, IL-6, and IL- 8 which are involved in the inflammatory response during lung cancer development⁶. Carvalho et al⁷ further report that more than 75% of the hypermethylated genes belong to the class of transcriptional regulators⁷. Aberrant DNA methylation of OLIG1 (oligodendrocyte transcription factor 1) is regarded as a novel prognostic factor in NSCLC (non-small cell lung cancer)8. Hypomethylation of intragenic LINE-1 (long interspersed nuclear element-1) represses transcription in cancer cells through AGO2 (argonaute 2)9. LDHC (lactate dehydrogenase C) gene expression in cancer cells is regulated by Sp1, CREB, and CpG island methylation¹⁰. In addition, it is reported that miRNA expression also can be regulated by DNA methylation. Let-7a-3 belongs to the archetypal let-7 miRNA gene family and is found to be methylated by the DNA methyltransferases DNMT1 and DNMT3B, resulting enhanced lung tumor phenotypes¹¹.

¹Department of Cardiothoracic Surgery, Changhai Hospital, Shanghai, China

²Department of Cardiothoracic Surgery, the 411nd Hospital of PLA, Shanghai, China

³Department of Thoracic Surgery, Tangdu Hospital, Fourth Military Medical University, Xi'an, China

Gene Ontology (GO) and gene-set-enrichment analysis approach (GSEA) are utilized to characterize the underlying function of identified genes in lung cancer. Genes that differentially expressed between normal lung tissue and cancer show enrichment in gene ontology (GO) terms associated with mitosis and proliferation¹². The Fas signaling pathway and the antigen processing and presentation pathway are most significantly related with lung cancer susceptibility by GSEA¹³. DNA copy number aberrations in SCLC are significantly activated by the focal adhesion pathway¹⁴. MicroRNA signatures in tumor tissue are related to pathway "angiogenesis" in NSCLC evaluated by GSEA¹⁵.

In this study, we aim to integrate information of functional annotation, transcription factors and microRNAs to elaborate the role of DNA methylation in the mechanism of lung cancer development.

Materials and Methods

DNA Methylation Data

The DNA Methylation data we used was downloaded from National Center for Biotechnology Gene Expression Omnibus (NCBI GEO, http://www.ncbi.nlm.nih.gov/geo/) under accession number GSE32867¹⁶, which detects the DNA methylation status of 27,578 CpG (cytosine-phosphate-guanine) dinucleotides CpGs sites on 28 lung cancer samples and 27 adjacent non-lung cancer tissue samples. The number of probes is 27,578, corresponding to 14,495 protein-coding genes.

Gene Expression Data

The gene expression data we used was downloaded from NCBI GEO (http://www.ncbi.nlm.nih.gov/geo/) under accession number GSE32863¹⁷, which is based on the platform of Illumina HumanWG-6 v3.0 expression beadchip. This profile is compromised by 58 lung cancer samples and 58 adjacent non-cancer samples. The number of probes is 48,803, corresponding to 25,188 protein-coding genes.

Date Preprocessing

For DNA methylation data, when multiple probes were mapped to the same gene, the level of DNA methylation β for this gene was represented by the mean value of all the probes. For gene expression, when one probe was mapped to multiple genes, these probes were removed. For

multiple probes that were mapped to the same gene, expression level of this gene was represented by the median expression value of all the probes.

Differential Analysis

By comparing the methylation pattern of lung cancer samples and non-cancer lung samples, we identified differentially methylated genes (DMGs). The statistical method used was Student's *t*-test (FDR: False Discovery Rate = 0.01), with the threshold of differential methylation level at 10%. In this manner, we identified genes and methylation loci with significant DNA methylation under disease status.

Gene Ontology and Pathway Analysis

In order to analyze the biological function that is regulated by methylation, we performed Gene Ontology (biological processes, BP) analysis for DMGs. These genes were also analyzed from the perspective of KEGG (Kyoto Encyclopedia of Genes and Genomes) pathway. Both processes utilize DAVID (Database for Annotation, Visualization and Integration Discover)¹⁸, the parameter "count" is set to 2 and corrected *p*-value is set to 0.01 for multiple testing.

Results

DNA Methylation Analysis

After mapping the DNA methylation probes to the genome, 72.3% of the methylated loci were less than 500 bp (base pair) from TSS (Transcription Start Site) of genes and 92% of the methylated loci were less than 1,000 bp from TSS of genes (Figure 1). Significant methylation difference was identified in 108 genes between lung cancer and non-cancer samples with multiple testing corrected FDR < 0.01. As shown in Figure 2, 84 genes had high methylation level near their TSS. The DNA methylation level was low for 14 genes in tumor samples.

Functional Analysis of Differentially Methylated Genes

The differentially methylated genes (DMGs) were annotated to biological processes and pathways. As shown in Table I, 108 DMGs were significantly enriched in 19 biological processes (including apoptosis process, the cell death process, ion transfer and the immune response regulation process) or four pathways (Calcium

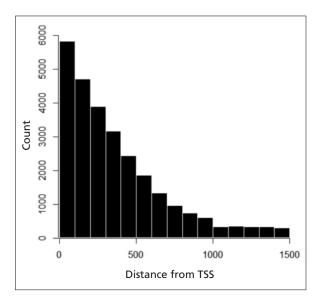


Figure 1. DNA methylation loci are significantly concentrated in the vicinity of the promoter of the gene. A total of 19,948 (72.3%) sites locate at the core promoter of gene. A total of 25,368 (92%) sites locate within 1kb from TSS of gene. The x-axis represents the distance between methylation site and TSS of gene, and the y-axis represents the number of methylation loci.

signaling pathway, Cardiac muscle contraction, Alzheimer's disease, and Huntington's disease).

Transcriptional Regulation of Methylation

In mammals, DNA methylation is widespread in the genome epigenetic modification. A methyl (-CH3), which can block the binding of the protein, is added to cytosine of CG diad on DNA by DNA methylation transferase enzyme. It is reported that DNA methylation near TSS can regulate gene expression. Thus, aberrant DNA methylation has been found in embryonic development and genomic imprinting 19,20 . In this study, we obtained gene expression of 47 DMGs between lung samples and adjacent non-cancer lung samples. Among these 47 genes, we further identified 18 genes that differentially expressed in lung cancer based on KS (Kolmogorov-Smirnov)-test method (p < 0.05).

Regulation Network of DNA Methylation

The TRANSFAC (Transcription Factor database) provides abundant data on eukaryotic transcription factors, their experimentally-proven binding sites, consensus binding sequences and regulated genes²¹. For the DMGs, we picked out the transcriptional factors and extracted their target genes based on TRANSFAC version 11.4. Fi-

nally we obtained 21 relationships between DMGs and their corresponding transcriptional targets, involving five DMGs. Moreover, we identified the transcriptional factors that target DMGs. As result, 13 relationships between upstream transcriptional factors and DMGs were found to regulate aberrant DNA methylation, involving 11 transcriptional factors. Since the initial transcript of miRNA is unknown, it is not possible to directly determine its upstream proteins that regulate the transcription process. TransmiR (transcription factor-microRNA regulation) database stores the relationship between miRNA and transcriptional factors by literature mining²². From TransmiR database we identified the transcriptional relationship between two DNA hypermethylated genes and six miRNAs (pre-mir-302a/b/c/d, pre-mir-367, and let-7a). We constructed a lung cancer-related methylation regulatory network by integrating three types of DNA methylation regulation. As shown in Figure 3, as epigenetic modification markers on the genome, DNA methylation participates in carcinogenesis at the transcriptional and post-transcriptional level. In the process of carcinogenesis aberrant DNA methylation will prevent its binding with the upstream regulatory proteins and in-

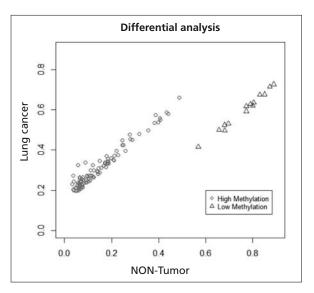


Figure 2. Differential analysis of DNA methylation in lung cancer. The red circles represent differentially methylated genes in lung cancer which locate above the diagonal. The blue triangles represent genes lowly methylated in lung cancer. The x-axis represents methylation level of gene in non-cancer lung samples and the y-axis represents the DNA methylation level of gene in lung cancer. When multiple genes are mapped to one gene, the DNA methylation level of the gene is represented by mean value of all the probes.

Table I. GO and KEGG annotation of differentially methylated genes (DMGs).

Category	GO ID	ρ value	Term
GOTERM_BP	GO:0065008	3.18E-04	Regulation of biological quality
GOTERM_BP	GO:0006816	7.82E-04	Calcium ion transport
GOTERM_BPL	GO:0015674	0.002029	di-, tri-valent inorganic cation transport
GOTERM_BP	GO:0003013	0.00258	Circulatory system process
GOTERM_BP	GO:0008015	0.00258	Blood circulation
GOTERM_BP	GO:0002026	0.00294	Regulation of the force of heart contractio
GOTERM_BP	GO:0006915	0.003402	Apoptosis
GOTERM_BP	GO:0012501	0.003752	Programmed cell death
GOTERM_BP	GO:0006811	0.005388	Ion transport
GOTERM_BP	GO:0048518	0.00542	Positive regulation of biological process
GOTERM_BP	GO:0050776	0.005901	Regulation of immune response
GOTERM_BP	GO:0048584	0.00706	Positive regulation of response to stimulus
GOTERM_BP	GO:0002684	0.007309	Positive regulation of immune system proces
GOTERM_BP	GO:0006959	0.007627	Humoral immune response
GOTERM_BP	GO:0032501	0.007766	Multicellular organismal process
GOTERM_BP	GO:0051240	0.008095	Positive regulation of multicellular organi
GOTERM_BP	GO:0007188	0.009322	G-protein signaling, coupled to cAMP nucleo
GOTERM_BP	GO:0006958	0.00953	Complement activation, classical pathway
GOTERM_BP	GO:0051605	0.009624	Protein maturation by peptide bond cleavage
KEGG_PATHWAY	hsa04020	1.45E-05	Calcium signaling pathway
KEGG_PATHWAY	hsa04260	4.31E-04	Cardiac muscle contraction
KEGG_PATHWAY	hsa05010	5.08E-04	Alzheimer's disease
KEGG_PATHWAY	hsa05016	7.83E-04	Huntington's disease

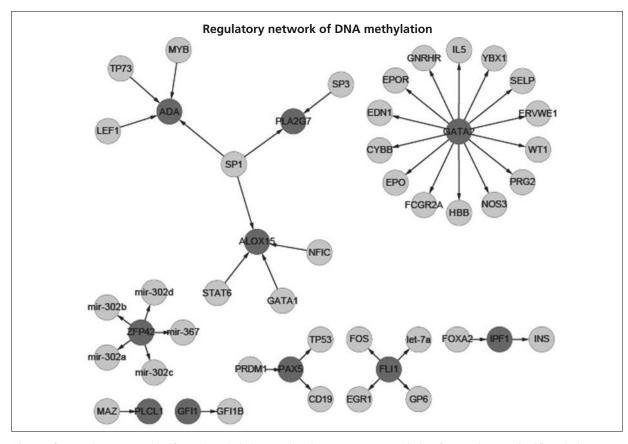


Figure 3. Regulatory network of DNA methylation. Red nodes represent transcription factors that are significantly hypermethylated in lung cancer. Blue nodes represent microRNA and target genes of transcriptional factors. Edges represent transcriptional regulation of DNA methylation.

hibit the function of gene regulated by them. At the same time, the aberrant DNA methylation will regulate the expression of downstream genes and miRNAs, and consequently affect cell development, differentiation and apoptosis.

Discussion

DNA methylation profiles of lung cancer have suggested that the methylation is an important marker in genome modification²³. In this study, we identified 108 DMGs based on DNA methylation profiles of lung cancer samples and non-caner lung samples. These DMGs were predicted to be involved in lung cancer by cell development, immuneresponse and apoptosis pathways. Further we constructed the regulatory network of DNA methylation by integrating transcriptional and posttranscriptional information. Some DMGs regulated the downstream target genes and microR-NAs and on the contrary they were regulated by upstream transcriptional factors. We will discuss them based on previous studies.

Paired box protein Pax-5 (PAX5) encodes a potent transcription factor that plays a key role in B-cell development and carcinogenic events²⁴. PAX5 expression is regulated by its upstream factor PRDM1²⁵ or methylation in NSCLC patients²⁶. The anti-tumorigenic function of PAX5 is mediated by upregulating downstream targets of tumor protein 53 (p53) and mesenchymal-epithelial transition factor (MET)²⁷.

Adenosine deaminase (ADA) is an enzyme of purine catabolism which catalizes adenosine into inosine and the deficit of ADA causes the inhibition of DNA synthesis and repair²⁸. ADA is also demonstrated to be a transcriptional target of p73 and p63 that sustain cellular growth by transcriptional activation of cell cycle progression genes²⁹. ASD is inversely associated with the proliferation and differentiation of lymphocytes, especially T lymphocytes³⁰ and thus is also important for immune response. Patients suffering from lung cancer have lower sputum ADA activity compared with pulmonary tuberculosis³¹.

GATA-binding protein 2 (GATA-2) is shown to be directly recruited to the promoter region of the androgen receptor upon androgen stimulation of prostate cancer cells and induce androgen receptor and key androgen-regulated gene AZGP1 expression which is associated with the progression to metastatic disease³². In this study, we also found that GATA2 was methylated in lung cancer and

GATA2 might be involved in lung cancer progression by regulating gonadotropin-releasing hormone receptor (GNRHR) expression, which is in line with results described by Schang et al³³. In addition, recent study indicates that KRAS mutant lung cancer cells depend on the transcription factor GATA2 and GATA2 depletion inhibits suppresses KRAS mutant cell growth by suppressing proteasome activity and inactivating expression of the IL-1 signaling pathway or Rho signaling^{34,35}.

Friend leukemia virus integration 1 (FLI1), an Ets transcription factor family member, plays a central and rate-limiting role in the pathogenesis of Ewing's sarcoma (EWS) by fusion with EWS gene. Recent studies also suggest that FLI1 is a candidate gene for squamous lung cancer by the transcriptome network analysis³⁶. Increasing evidence indicates that EWS-FLI-1 can regulate miRNA expression³⁷. For example, De Vito show that EWS-FLI-1 can directly suppress let-7a expression in EWS family tumors development and subsequently, let-7a expression regulates EWS growth by its target gene HMGA2, an oncogene in a variety of human tumors³⁸. In this study, we also found that FLI-1 might be involved in lung cancer by modulating let-7a expression.

However, there are some limits in this study. Since the current regulatory relationships between miRNAs and transcriptional factors are limited, therefore, additional data in existing databases is needed to construct the regulating network of DNA methylation. Moreover, apart from the markers of DNA methylation, histone modification is also an important dynamic marker of the genome³⁹. Subsequent analysis will integrate data from multiple aspects and, thus, interpret lung cancer at a system perspective.

Conflict of Interest

None declared.

References

- STERNER-KOCK A, THOREY IS, KOLI K, WEMPE F, OTTE J, BANGSOW T, KUHLMEIER K, KIRCHNER T, JIN S, KESKI-OJA J, VON MELCHNER H. Disruption of the gene encoding the latent transforming growth factor-beta binding protein 4 (LTBP-4) causes abnormal lung development, cardiomyopathy, and colorectal cancer. Genes Dev 2002; 16: 2264-2273.
- COLLINS LG, HAINES C, PERKEL R, ENCK RE. Lung cancer: diagnosis and management. Am Family Phys 2007; 75: 56-63.

- FERLAY J, SHIN HR, BRAY F, FORMAN D, MATHERS C, PARKIN DM. Estimates of worldwide burden of cancer in 2008: Globocan 2008. Int J Cancer 2010; 127: 2893-2917.
- 4) LOKK K, VOODER T, KOLDE R, VÄLK K, VÖSA U, ROOSIPUU R, MILANI L, FISCHER K, KOLTSINA M, UR-GARD E. Methylation markers of early-stage nonsmall cell lung cancer. PLoS One 2012; 7: e39813.
- 5) PARK JY, KIM D, YANG M, PARK HY, LEE SH, RINCON M, KREAHLING J, PLASS C, SMIRAGLIA DJ, TOCKMAN MS, KIM SJ. Gene silencing of SLC5A8 identified by genome-wide methylation profiling in lung cancer. Lung Cancer 2012; 79: 198-204.
- 6) TEKPLI X, LANDVIK NE, ANMARKUD KH, SKAUG V, HAUGEN A, ZIENOLDDINY S. DNA methylation at promoter regions of interleukin 1b, interleukin 6, and interleukin 8 in non-small cell lung cancer. Cancer Immunol Immunother 2013; 62: 337-345.
- CARVALHO RH, HABERLE V, HOU J, VAN GENT T, THONGJUEA S, VAN IJCKEN W, KOCKX C, BROUWER R, RUKERS E, SIEUWERTS A. Genome-wide DNA methylation profiling of non-small cell lung carcinomas. Epigenetics Chromatin 2012; 5: 9.
- Brena RM, Morrison C, Liyanarachchi S, Jarjoura D, Davuluri RV, Otterson GA, Reisman D, Glaros S, Rush LJ, Plass C. Aberrant DNA methylation of OLIG1, a novel prognostic factor in non-small cell lung cancer. PLoS Med 2007; 4: e108.
- APORNTEWAN C, PHOKAEW C, PIRIYAPONGSA J, NGAM-PHIW C, ITTIWUT C, TONGSIMA S, MUTIRANGURA A. Hypomethylation of intragenic LINE-1 represses transcription in cancer cells through AGO2. PLoS One 2011; 6: e17934.
- TANG H, GOLDBERG E. Homo sapiens lactate dehydrogenase c (Ldhc) gene expression in cancer cells is regulated by transcription factor Sp1, CREB, and CpG island methylation. J Androl 2009; 30: 157-167.
- 11) BRUECKNER B, STRESEMANN C, KUNER R, MUND C, MUSCH T, MEISTER M, SÜLTMANN H, LYKO F. The human let-7a-3 locus contains an epigenetically regulated microRNA gene with oncogenic function. Cancer Res 2007; 67: 1419-1423.
- 12) URGARD E, VOODER T, VOSA U, VALK K, LIU M, LUO C, HOTI F, ROOSIPUU R, ANNILO T, LAINE J, FRENZ CM, ZHANG L, METSPALU A. Metagenes associated with survival in non-small cell lung cancer. Cancer Inform 2011; 10: 175-183.
- 13) WEI S, WANG LE, MCHUGH MK, HAN Y, XIONG M, AMOS CI, SPITZ MR, WEI QW. Genome-wide geneenvironment interaction analysis for asbestos exposure in lung cancer susceptibility. Carcinogenesis 2012; 33: 1531-1537.
- 14) OCAK S, YAMASHITA H, UDYAVAR AR, MILLER AN, GONZALEZ AL, ZOU Y, JIANG A, YI Y, SHYR Y, ESTRADA L, QUARANTA V, MASSION PP. DNA copy number aberrations in small-cell lung cancer reveal activation of the focal adhesion pathway. Oncogene 2010; 29: 6331-6342.

- 15) DONNEM T, FENTON CG, LONVIK K, BERG T, EKLO K, ANDERSEN S, STENVOLD H, AL-SHIBLI K, AL-SAAD S, BREMNES RM, BUSUND LT. Microrna signatures in tumor tissue related to angiogenesis in non-small cell lung cancer. PloS One 2012; 7: e29671.
- 16) SELAMAT SA, CHUNG BS, GIRARD L, ZHANG W, ZHANG Y, CAMPAN M, SIEGMUND KD, KOSS MN, HAGEN JA, LAM WL, LAM S, GAZDAR AF, LAIRD-OFFRINGA IA. Genome-scale analysis of DNA methylation in lung adenocarcinoma and integration with mrna expression. Genome Res 2012; 22: 1197-1211.
- 17) SHARMA S, STOLINA M, LUO J, STRIETER RM, BURDICK M, ZHU LX, BATRA RK, DUBINETT SM. Secondary lymphoid tissue chemokine mediates T cell-dependent antitumor responses in vivo. J Immunol 2000; 164: 4558-4563.
- HUANG DA W, SHERMAN BT, LEMPICKI RA. Bioinformatics enrichment tools: Paths toward the comprehensive functional analysis of large gene lists. Nucleic Acids Res 2009; 37: 1-13.
- 19) YIN LJ, ZHANG Y, LV PP, HE WH, WU YT, LIU AX, DING GL, DONG MY, QU F, XU CM, ZHU XM, HUANG HF. Insufficient maintenance DNA methylation is associated with abnormal embryonic development. BMC Med 2012; 10: 26.
- Li E, Beard C, Jaenisch R. Role for DNA methylation in genomic imprinting. Nature 1993; 366: 362-365.
- 21) MATYS V, KEL-MARGOULIS OV, FRICKE E, LIEBICH I, LAND S, BARRE-DIRRIE A, REUTER I, CHEKMENEV D, KRULL M, HORNISCHER K, VOSS N, STEGMAIER P, LEWICKI-POTAPOV B, SAXEL H, KEL AE, WINGENDER E. Transfac and its module transcompel: transcriptional gene regulation in eukaryotes. Nucleic Acids Res 2006; 34: D108-110.
- 22) WANG J, Lu M, QIU C, CUI Q. Transmir: a transcription factor-microrna regulation database. Nucleic Acids Res 2010; 38: D119-122.
- 23) NOUSHMEHR H, WEISENBERGER DJ, DIEFES K, PHILLIPS HS, PUJARA K, BERMAN BP, PAN F, PELLOSKI CE, SULMAN EP, BHAT KP, VERHAAK RG, HOADLEY KA, HAYES DN, PEROU CM, SCHMIDT HK, DING L, WILSON RK, VAN DEN BERG D, SHEN H, BENGTSSON H, NEUVIAL P, COPE LM, BUCKLEY J, HERMAN JG, BAYLIN SB, LAIRD PW, ALDAPE K. Identification of a CpG island methylator phenotype that defines a distinct subgroup of glioma. Cancer Cell 2010; 17: 510-522.
- 24) O'BRIEN P, MORIN P, OUELLETTE RJ, ROBICHAUD GA. The Pax-5 gene: a pluripotent regulator of B-cell differentiation and cancer disease. Cancer Res 2011; 71: 7345-7350.
- 25) DESAI S, MAURIN M, SMITH MA, BOLICK SC, DESSUREAULT S, TAO J, SOTOMAYOR E, WRIGHT KL. PRDM1 is required for mantle cell lymphoma response to bortezomib. Mol Cancer Res 2010; 8: 907-918.
- 26) PESEK M, KOPECKOVA M, BENESOVA L, MESZAROSOVA A, MUKENSNABL P, BRUHA F, MINARIK M. Clinical significance of hypermethylation status in NSCLC: Evaluation of a 30-gene panel in patients with advanced disease. Anticancer Res 2011; 31: 4647-4652.

- 27) LI X, CHEUNG K, MA X, TIAN L, ZHAO J, GO M, SHEN B, CHENG A, YING J, TAO Q. Epigenetic inactivation of paired box gene 5, a novel tumor suppressor gene, through direct upregulation of p53 is associated with prognosis in gastric cancer patients. Oncogene 2011; 31: 3419-3430.
- 28) Tullo A, Mastropasqua G, Bourdon JC, Centonze P, Gostissa M, Costanzo A, Levrero M, Del Sal G, Saccone C, Sbisa E. Adenosine deaminase, a key enzyme in DNA precursors control, is a new p73 target. Oncogene 2003; 22: 8738-8748.
- 29) LEFKIMMIATIS K, CARATOZZOLO MF, MERLO P, D'ERCHIA AM, NAVARRO B, LEVRERO M, TULLO A. P73 and p63 sustain cellular growth by transcriptional activation of cell cycle progression genes. Cancer Res 2009; 69: 8563-8571.
- OCANA I, MARTINEZ-VAZQUEZ J, SEGURA R, FERNANDEZ-DE-SEVILLA T, CAPDEVILA J. Adenosine deaminase in pleural fluids. Test for diagnosis of tuberculous pleural effusion. Chest 1983; 84: 51.
- 31) SAMARA I, BAKAKOS P, ORPHANIDOU D, LATSI P, KATSIM-POULA S, PAPAETIS GS, DIMAKOU K. Sputum adenosine deaminase activity in patients with pulmonary tuberculosis and lung cancer. Adv Clin Exp Med 2007; 16: 533.
- 32) BÖHM M, LOCKE W, SUTHERLAND R, KENCH J, HENSHALL S. A role for GATA2 in transition to an aggressive phenotype in prostate cancer through modulation of key androgen-regulated genes. Oncogene 2009; 28: 3847-3856.

- 33) SCHANG AL, GRANGER A, QUÉRAT B, BLEUX C, COHEN-TANNOUDJI J, LAVERRIÈRE J-N. GATA2-induced silencing and lim-homeodomain protein-induced activation are mediated by a bi-functional response element in the rat gnrh receptor gene. Mol Endocrinol 2013; 27: 74-91.
- 34) Kumar MS, Downward J. Abstract a26: GATA2 is necessary for oncogene-mutant non-small cell lung tumorigenesis. Clin Cancer Res 2012; 18: A26-A26.
- 35) Kumar MS, Hancock DC, Molina-Arcas M, Steckel M, East P, Diefenbacher M, Armenteros-Monterroso E, Lassailly F, Matthews N, Nye E. The GATA2 transcriptional network is requisite for RAS oncogene-driven non-small cell lung cancer. Cell 2012; 149: 642-655.
- 36) BAI J, Hu S. Transcriptome network analysis reveals potential candidate genes for squamous lung cancer. Int J Mol Med 2012; 29: 95-101.
- 37) BAN J, JUG G, MESTDAGH P, SCHWENTNER R, KAUER M, ARYEE D, SCHAEFER K, NAKATANI F, SCOTLANDI K, REITER M. Hsa-mir-145 is the top EWS-FLI1-repressed microRNA involved in a positive feedback loop in Ewing's sarcoma. Oncogene 2011; 30: 2173-2180.
- 38) DE VITO C, RIGGI N, SUVA ML, JANISZEWSKA M, HORLBECK J, BAUMER K, PROVERO P, STAMENKOVIC I. Let-7a is a direct EWS-FLI-1 target implicated in Ewing's sarcoma development. PLoS One 2011; 6: e23592.
- Berger SL. Histone modifications in transcriptional regulation. Curr Opin Genet Dev 2002; 12: 142-148.