

Knockdown of long noncoding RNA DLX6-AS1 inhibits migration and invasion of thyroid cancer cells by upregulating UPF1

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Abstract. – **OBJECTIVE:** Recently, long non-coding RNAs (lncRNAs) have attracted much attention for their roles in tumor progression. The aim of this study was to investigate the effect of lncRNA DLX6 antisense RNA 1 (DLX6-AS1) in the development of thyroid cancer (TC), and to explore the underlying mechanism.

PATIENTS AND METHODS: DLX6-AS1 expression in both TC cells and tissue samples was detected by quantitative real-time-Polymerase Chain Reaction (qRT-PCR). Moreover, transwell assay and wound healing assay were conducted. QRT-PCR and Western blot assay were used to explore the underlying mechanism. Furthermore, a functional DLX6-AS1 was identified *in vitro*.

RESULTS: DLX6-AS1 expression level in TC tissues was significantly higher than that of the corresponding normal tissues. Moreover, TC cell migration and invasion were markedly inhibited after DLX6-AS1 was knocked down *in vitro*. The mRNA and protein expressions of UPF1 were both remarkably up-regulated after knockdown of DLX6-AS1. Meanwhile, the expression level of UPF1 was negatively correlated with the expression of DLX6-AS1 in TC tissues. Furthermore, knockdown of DLX6-AS1 significantly inhibited tumor metastasis *in vivo*.

CONCLUSION: Knockdown of DLX6-AS1 could inhibit TC cell migration and invasion via upregulating UPF1, which might be a potential therapeutic target in TC.

Keywords:

long non-coding RNA, DLX6-AS1, Thyroid cancer (TC), UPF1.

Introduction

Thyroid cancer (TC) is common cancer originated from follicular or parafollicular thyroid cells. The morbidity of TC has greatly increased in the past decades worldwide¹. Since 2007, the incidence of TC has increased by an average of 4.5% annually in America. Currently, TC is the eighth most common cancer in China². The prognosis of TC patients remains poor, which brings a huge burden to patients and the society³. Therefore, it is urgent to discover new biomarkers for TC diagnosis and treatment.

Long non-coding RNAs (lncRNA) are a subtype of non-protein coding RNAs with more than 200 nucleotides in length. Recent studies have indicated that lncRNA serves as a major contributor in a variety of cellular activities. For example, lncRNA CCAT2 facilitates the proliferation and metastasis of intrahepatic cholangiocarcinoma⁴. lncRNA FENDRR suppresses cell proliferation and malignancy in non-small cell lung cancer by sponging miR-761⁵. lncRNAAC132217.4 enhances the metastasis of oral squamous cell carcinoma cells *via* regulating IGF2⁶. lncRNA SNHG1 inhibits the differentiation of Treg cells impeding the immune escape of breast cancer⁷. In addition, down-regulated lncRNA UCA1 acts as a novel non-invasive diagnostic biomarker for bladder cancer⁸. However, the clinical role and biological mechanism of lncRNA DLX6 antisense RNA 1 (DLX6-AS1) in the development of TC have not been fully elucidated.

In this study, we found that the expression of DLX6-AS1 was remarkably up-regulated in TC tissues. Knockdown of DLX6-AS1 markedly inhibited the migration and invasion of TC cells *in vitro*. Furthermore, we explored the underlying mechanism of DLX6-AS1 function in TC metastasis.

Patients and Methods

Cell Lines and Clinical Samples

60 TC patients who received surgery at Harbin Medical University Cancer Hospital were enrolled in this study. Human tissues were collected from these patients. Before the operation, written informed consent was achieved. No radiotherapy or chemotherapy was performed for any patient before the operation. Tissues obtained from surgery were stored immediately at -80°C for use. All tissues were confirmed by an experienced pathologist. This study was approved by the Ethics Committee of the Harbin Medical University Cancer Hospital.

Cell Culture

Human thyroid carcinoma cell lines (TPC-1, K1, SW579) and normal human thyroid cells (Nthy-ori 3-1) were provided by the Institute of Biochemistry and Cell Biology, Chinese Academy of Science (Shanghai, China). Cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM; HyClone, Logan, UT, USA) consisting of 10% fetal bovine serum (FBS; Gibco Technologies, Gaithersburg, MD, USA) and penicillin. Besides, the cells were maintained in an incubator with 5% CO_2 at 37°C .

Cell Transfection and Grouping

Lentiviral small hairpin RNA (shRNA) targeting DLX6-AS1 was synthesized and cloned into pLentiCMVla-EGFP-F2A-Puro vector (Biossetia Inc., San Diego, CA, USA). Subsequently, TPC-1 cells were used for packaging viruses, DLX6-AS1 shRNA (sh-DLX6-AS1) and empty vector (control).

Quantitative Real Time-Polymerase Chain Reaction (qRT-PCR)

Total RNA from tissues and cells was extracted using TRIzol reagent (Invitrogen, Carlsbad, CA, USA). Subsequently, extracted total RNA was reverse transcribed into complementary deoxyribonucleic acids (cDNAs) through the Re-

verse Transcription Kit (TaKaRa Biotechnology Co., Ltd., Dalian, China). Primers used for Real Time-Polymerase Chain Reaction (RT-PCR) were as follows: DLX6-AS1 primers forward: 5'-AGTTTCTCTCTAGATTGCC-3', reverse: 5'-ATTGACATGTTAGTGCC-3'; Glycerinaldehyde 3-phosphate dehydrogenase (GAPDH) primers forward: 5'-CCAATCAAGTGG-CAATGCTGG-3' and reverse: 5'-TGATC-GGACTGTGGTCAT-3'. The thermal cycle was as follows: 30 sec at 95°C , 40 cycles at 95°C , and 35 sec at

Wound Healing Assay

Cells were transferred into 6-well plates, followed by culture in DMEM medium overnight. After scratching with a plastic tip, the cells were cultured in serum-free DMEM. Wound closure was viewed at specific time points. Each assay was independently repeated three times.

Transwell Assay

After transfection, 2×10^5 cells in 100 μL serum-free DMEM were transformed to the upper chamber of an 8- μm culture insert (Corning, Lowell, MA, USA) coated with or without 50 μg laminin (BD Biosciences, Franklin Lakes, NJ, USA). Meanwhile, 20% FBS-DMEM was added to the lower chamber of culture inserts. 24 h later, these inserts were treated with methanol for 30 min and stained with hematoxylin for 20 min. Migrated and invaded cells were observed under an inverted microscope ($\times 40$), and the number of cells was counted. Three fields were randomly selected for each sample.

Western Blot Analysis

Radioimmunoprecipitation assay (RIPA; Beyotime, Shanghai, China) was utilized to extract the total protein in cells. The concentration of extracted protein was quantified by the bicinchoninic acid (BCA) protein assay kit (TaKaRa, Dalian, China). Target proteins were separated by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). Then, the cells were incubated with primary and corresponding secondary antibodies. Cell Signaling Technology (CST, Danvers, MA, USA) provided us rabbit anti-GAPDH and rabbit anti-UPF1, as well as goat anti-rabbit secondary antibody. Image J software (NIH, Bethesda, MD, USA) was applied for the assessment of the protein expression.

Xenograft Model

For tumor metastasis assay, transfected TPC-1 cells were injected into the tail vein of NOD/SCID mice (4-5 weeks old). After 4 weeks, the mice were sacrificed, and lung tissues were extracted. The number of metastatic nodules in lung tissue was then counted. The animal experiments were approved by the Animal Ethics Committee of Harbin Medical University.

Statistical Analysis

Statistical Product and Service Solutions (SPSS) 19.0 (SPSS, Chicago, IL, USA) was used for all statistical analysis. Data were presented as mean \pm Standard Deviation (SD). The Student's *t*-test was selected when appropriate. $p < 0.05$ was considered statistically significant.

Results

DLX6-AS1 Expression Level in TC Tissues and Cell Lines

QRT-PCR was first conducted to detect DLX6-AS1 expression in 60 patients' tissues and 10 TC cell lines. As a result, DLX6-AS1 was significantly upregulated in TC tissues and cell lines (Figure 1).

Knockdown of DLX6-AS1 Inhibited Migration and Invasion of TC Cells

DLX6-AS1 expression in TC cells was significantly higher than that of normal cells (Figure 2A). According to DLX6-AS1 expression

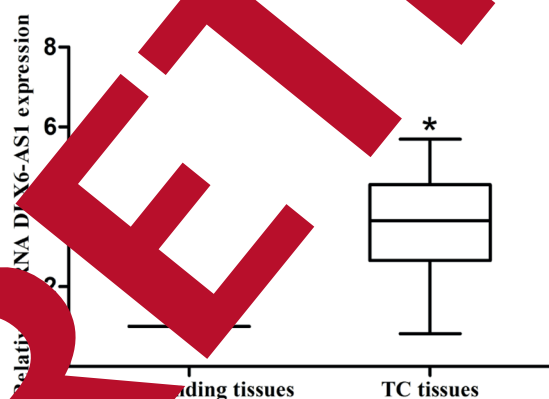


Figure 1. Expression level of DLX6-AS1 increased significantly in TC tissues. DLX6-AS1 expression increased markedly in TC tissues compared with the corresponding normal tissues. Data were presented as mean \pm standard error of the mean. * $p < 0.05$.

TC cells, TPC-1 cell line was chosen for knock-down of DLX6-AS1 *in vitro*. QRT-PCR was utilized to detect DLX6-AS1 expression (Figure 2B). Subsequent wound healing assay found that knockdown of DLX6-AS1 markedly inhibited TC cell migration (Figure 2C). The results of the transwell assay showed that the migration and invasion of TC cells were remarkably inhibited after DLX6-AS1 was knocked down (Figure 2D and 2E).

The Interaction Between UPF1 and DLX6-AS1 in TC Cells

UPF1 was predicted as the target protein of DLX6-AS1 through Starbase (http://starbase.sysu.edu.cn/starbase2/rbpLncRNA.php). The results of qRT-PCR showed that, compared with the empty vector (control) group, the expression level of UPF1 was significantly higher in TC cells of DLX6-AS1 shRNA (sh-DLX6-AS1) group (Figure 3A). Western blot assay demonstrated that after DLX6-AS1 was knocked down, the protein expression of UPF1 was significantly up-regulated (Figure 3B). Furthermore, we found that UPF1 expression in TC tissues was markedly lower than that of the corresponding normal tissues (Figure 3C). Correlation analysis demonstrated that UPF1 expression was negatively correlated with DLX6-AS1 expression in TC tissues (Figure 3D).

DLX6-AS1 Knockdown Inhibited Tumor Metastasis In Vivo

The ability of DLX6-AS1 in tumor metastasis was detected *in vivo*. The number of metastatic nodules in lung tissues of the sh-DLX6-AS1 group was significantly reduced when compared with the control group (Figure 4A). The expression level of DLX6-AS1 and UPF1 in dissected nodules tissues was detected by RT-qPCR. The results showed that DLX6-AS1 was lowly-expressed in the shRNA group when compared with the control group (Figure 4B). However, UPF1 was highly expressed in the shRNA group when compared with the control group (Figure 4C). The above results suggested that DLX6-AS1 could induce tumor metastasis *via* down-regulating UPF1 *in vivo*.

Discussion

Plenty of lncRNAs have been proved to play important roles in the occurrence and progres-

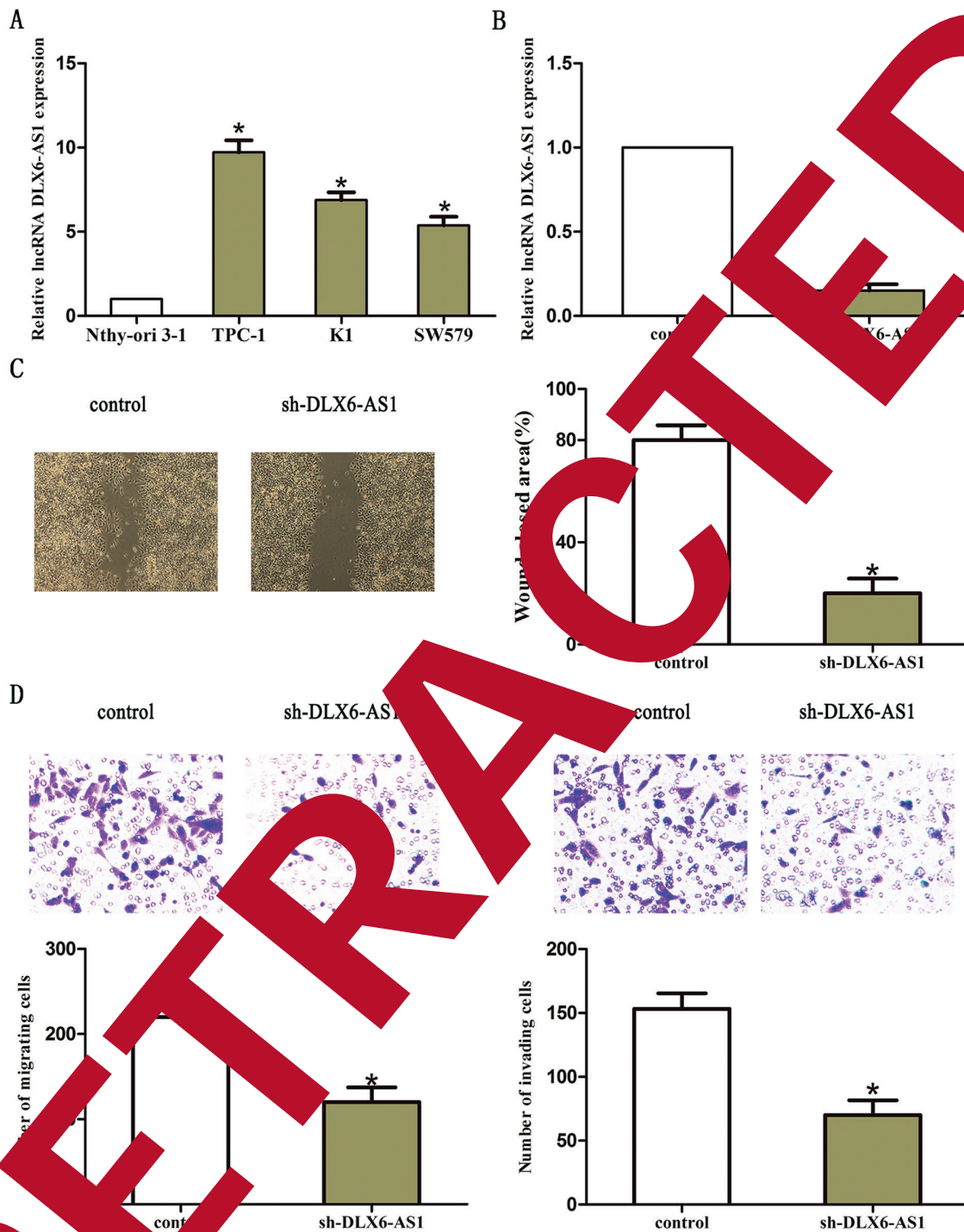


Fig. 2. Knockdown of DLX6-AS1 inhibited TC cell migration and invasion. **A**, Expression level of DLX6-AS1 relative to GAPDH in human TC cell lines and Nthy-ori 3-1 (normal human thyroid cell line) was determined by qRT-PCR. **B**, DLX6-AS1 expression in TPC-1 TC cells transduced with DLX6-AS1 shRNA (sh-DLX6-AS1) and empty vector (control) was determined by qRT-PCR. GAPDH was used as an internal control. **C**, Wound healing assay showed that the migrated length of TPC-1 cells in the sh-DLX6-AS1 group decreased significantly when compared with the control group (magnification: 40 \times). **D**, Wound healing assay showed that the migrated length of TPC-1 cells in the sh-DLX6-AS1 group decreased significantly when compared with the control group (magnification: 40 \times). **E**, Transwell assay showed that the number of migrated cells decreased markedly *via* knockdown of DLX6-AS1 in TPC-1 TC cells (magnification: 40 \times). **F**, Transwell assay showed that the number of invaded cells decreased remarkably *via* knockdown of DLX6-AS1 in TPC-1 TC cells (magnification: 40 \times). The results represented the average of three independent experiments (mean \pm standard error of the mean). * p <0.05, compared with control cells.

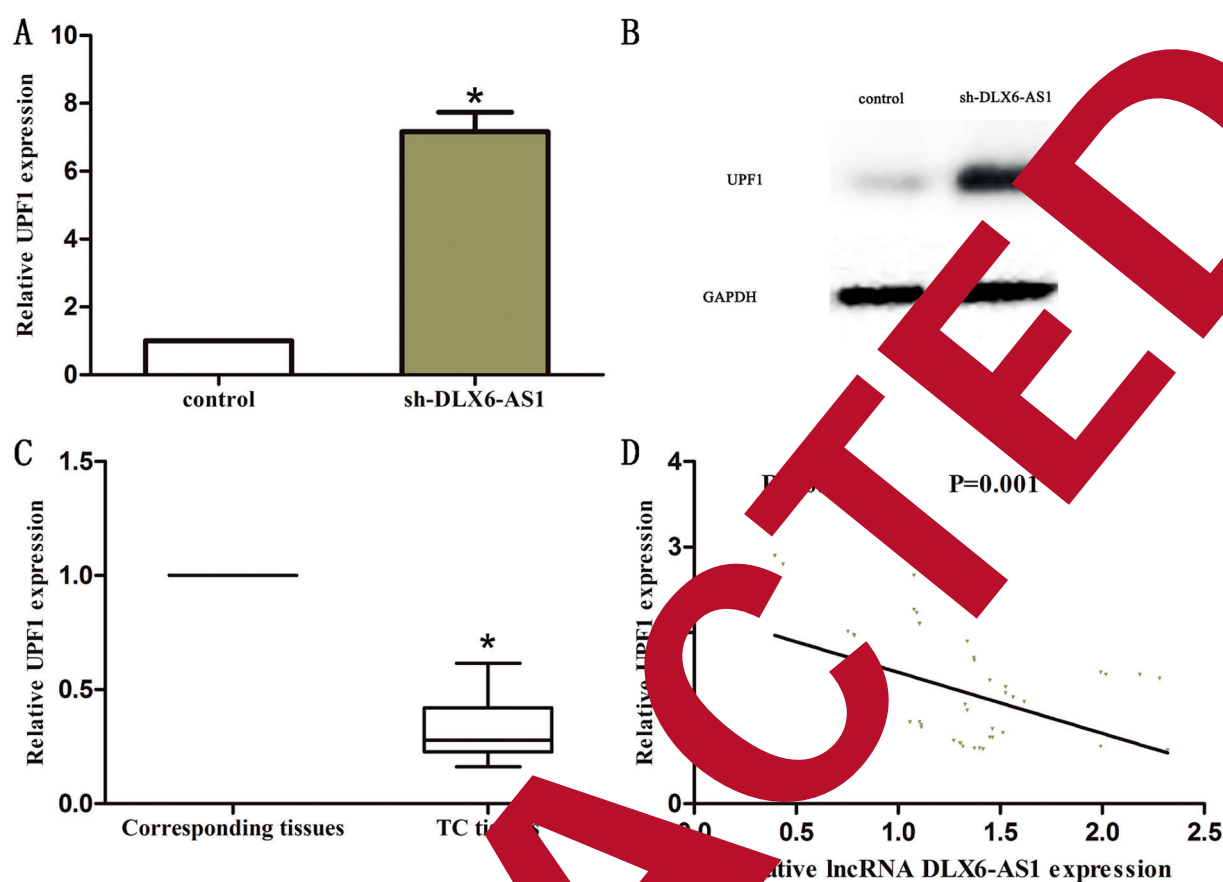


Figure 3. Interaction between DLX6-AS1 and UPF1. **A**, RT-PCR results showed that UPF1 expression was significantly higher in the DLX6-AS1 shRNA (sh-DLX6-AS1) group compared with the empty vector (control) group. **B**, Western blot assay revealed that protein expression of UPF1 was up-regulated in the DLX6-AS1 shRNA (sh-DLX6-AS1) group compared with the empty vector (control) group. **C**, UPF1 expression was significantly down-regulated in TC tissues when compared with the corresponding normal tissues. **D**, There is a negative linear correlation between the expression level of UPF1 and DLX6-AS1 in TC tissues. The results represented the average of three independent experiments. Data were presented as mean \pm standard error of the mean. * $p < 0.05$.

sion of TC. For instance, lncRNA-SHOG7 regulates the proliferation, apoptosis and invasion of bladder cancer cells⁹. Down-regulated lncRNA ANRIL promotes tumor metastasis in thyroid cancer via TGF-beta/Smad signaling pathway¹⁰. LncRNA HOTAIR promotes the development of cervical cancer by regulating Notch pathway¹¹. Meanwhile, lncRNA ENST00000537266 and ENST00000426465 are important regulators of cell proliferation in papillary thyroid cancer¹². In addition, lncRNA MEG3 functions as a tumor suppressor in cervical cancer, which leads to the inhibition of tumor growth¹³.

Among non-coding RNA DLX6 antisense RNA (DLX6-AS1), located in 7q21.3, has been recently explored. Scholars^{14,15} have found that DLX6-AS1 is abnormally expressed in several

cancers, and is related to tumor progression. For example, DLX6-AS1 promotes the proliferation and invasion of renal cell carcinoma cells by targeting miR-26a axis¹⁶. DLX6-AS1 induces cell invasion by regulating miR-181b in pancreatic cancer¹⁷. DLX6-AS1 promotes the proliferation and metastasis of non-small cell lung cancer by regulating miR-144¹⁸. In this study, we found that DLX6-AS1 was significantly up-regulated both in TC tissues and cells. After DLX6-AS1 was knocked down, TC cell migration and invasion was found markedly inhibited. The above results indicated that DLX6-AS1 promoted tumorigenesis of TC and might act as an oncogene.

UPF1 (UPF1 RNA helicase and ATPase) was predicted as the target protein of DLX6-AS1 through Starbase v2.0. It has been indicated that

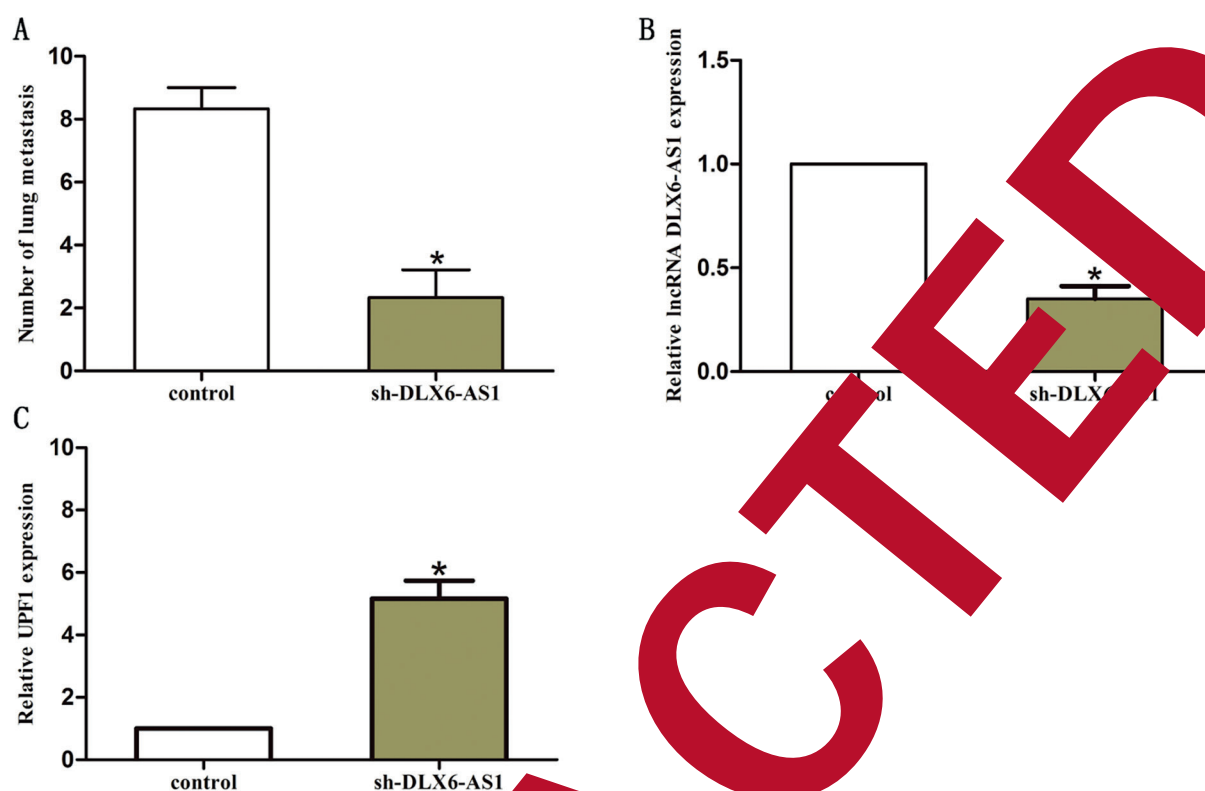


Figure 4. Knockdown of DLX6-AS1 inhibited metastasis *in vivo*. **A**, The number of metastatic nodules in lung tissues of sh-DLX6-AS1 group was significantly reduced compared with the control group. **B**, DLX6-AS1 was lowly-expressed in dissected nodules of the sh-DLX6-AS1 group compared with the control group. **C**, UPF1 was highly-expressed in dissected nodules of the sh-DLX6-AS1 group compared with the control group. The results represented the average of three independent experiments (mean \pm standard error of the mean). * $p < 0.05$ compared with control cells.

UPF1 plays a crucial role in tumor growth and invasion in multiple cancers and is negatively regulated by lncRNA MALAT1. MALAT1 participates in the development of gastric cancer and UPF1 also represses the development of hepatocellular carcinoma *via* targeting miR-101-3p [17²⁰. Moreover, SNHG6 acts as an oncogene in pancreatic carcinoma by regulating miR-101-3p [17].

In the present work, UPF1 expression was markedly up-regulated after knockdown of DLX6-AS1. Moreover, UPF1 expression in TC tissues was negatively correlated with DLX6-AS1 expression. All the above results suggested that DLX6-AS1 might promote tumor growth of TC *via* down-regulating UPF1. Further knockdown of DLX6-AS1 significantly inhibited metastasis *in vivo*.

Conclusions

The present results identified that DLX6-AS1 was remarkably up-regulated in TC tissues and

cell lines. Besides, DLX6-AS1 remarkably enhanced TC cell migration and invasion by down-regulating UPF1. We suggest that DLX6-AS1 might contribute to therapy for TC as a candidate target.

Conflict of Interests

The Authors declare that they have no conflict of interests.

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